



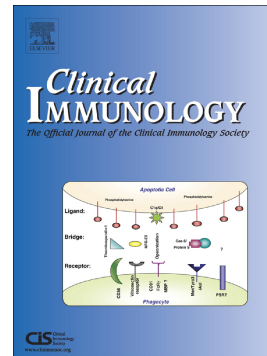
Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Journal Pre-proof

A hitchhiker's guide through the COVID-19 galaxy

Susanna Felsenstein, Andreas Otto Reiff



PII: S1521-6616(21)00186-8

DOI: <https://doi.org/10.1016/j.clim.2021.108849>

Reference: YCLIM 108849

To appear in: *Clinical Immunology*

Received date: 29 May 2021

Accepted date: 4 September 2021

Please cite this article as: S. Felsenstein and A.O. Reiff, A hitchhiker's guide through the COVID-19 galaxy, *Clinical Immunology* (2021), <https://doi.org/10.1016/j.clim.2021.108849>

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2021 Published by Elsevier Inc.

Title: A hitchhiker's guide through the COVID-19 Galaxy

Authors: **1. Susanna Felsenstein, MSc, DTM&H**
 University of Liverpool, Faculty of Health and Life Sciences
 Brownlow Hill, Liverpool, L69 3GB
 United Kingdom
 Honorary Senior Research Associate, WPRO/WHO

 Tel.: +49 17670985464
 Email: sumakafe@liverpool.ac.uk

2. Andreas Otto Reiff, MD*
 Adjunct Professor
 Arthritis & Rheumatic Diseases
 Oregon Health & Science University
 3181 SW Sam Jackson Park Rd.
 Portland, OR 97239
 United States

Senior Vice President of Medical Sciences, Immunology and Inflammation Parexel

Business Address	8 Federal Street, Billerica, MA 01821
Business Phone	+1 978 313 3900
Home Address	10105 Southwest Morrison St., Portland, OR 97225, USA
Home Phone	(626) 264 2734

Email: reiffa@ohsu.edu
 * corresponding author

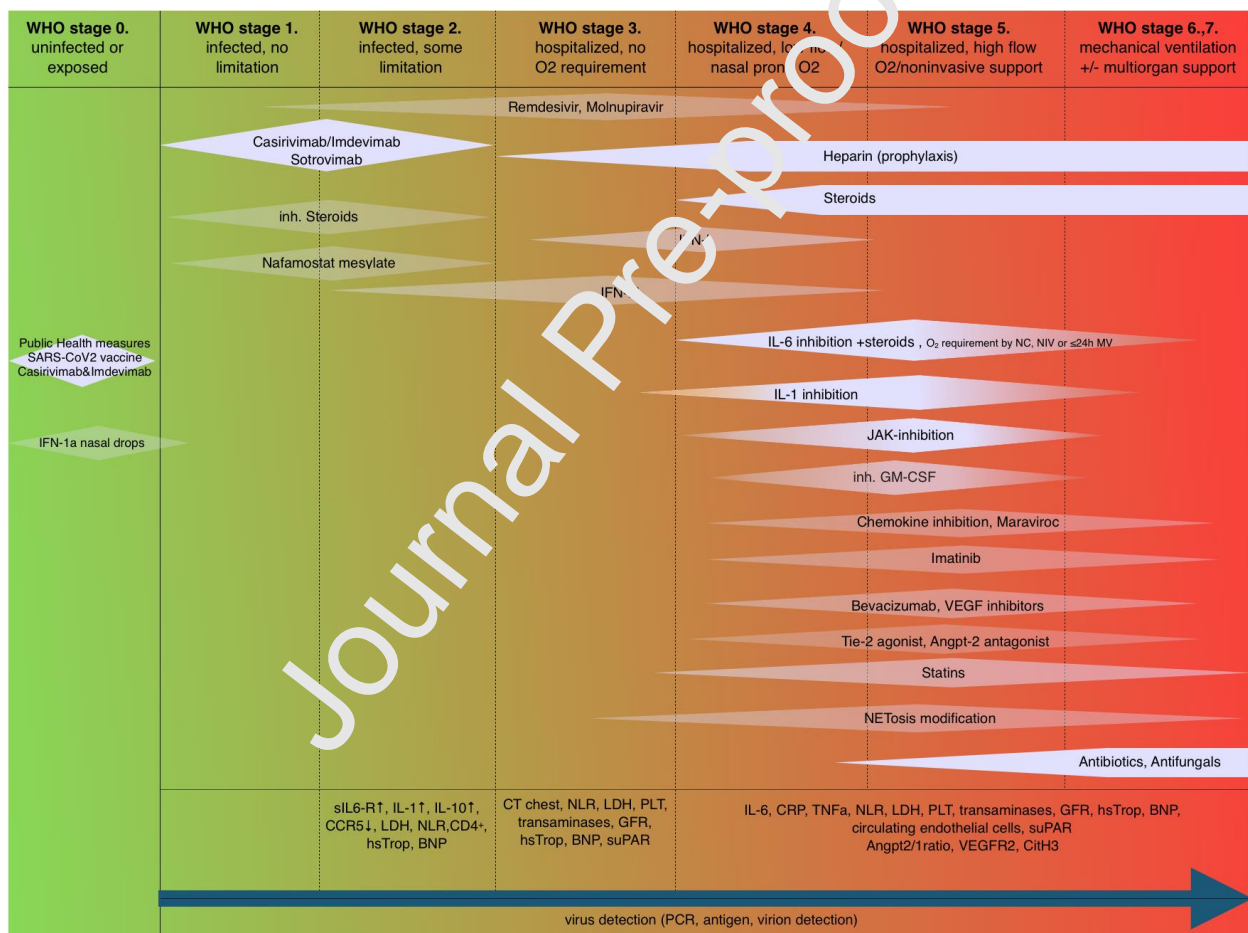
Word count: 11581

Abstract: Numerous reviews have summarized the epidemiology, pathophysiology and the various therapeutic aspects of Coronavirus disease 2019 (COVID-19), but a practical guide on “how to treat whom with what and when” based on an understanding of the immunological background of the disease stages remains missing.

This review attempts to combine the current knowledge about the immunopathology of COVID-19 with published evidence of available and emerging treatment options.

We recognize that the information about COVID-19 and its treatment is rapidly changing, but hope that this guide offers those on the frontline of this pandemic an understanding of the host response in COVID-19 patients and supports their ongoing efforts to select the best treatments tailored to their patient's clinical status.

Figure 1.



Introduction

Since SARS-CoV-2 was first identified in December 2019 in Wuhan, China [1], coronavirus disease 2019 (COVID-19) has evolved into a pandemic resulting in 223 million infections and almost 4.6 million deaths [2]. Due to the rapid global spread of the virus and lack of adequate worldwide vaccine coverage, novel viral variants differing in transmission dynamics and pathogenicity have continued to evolve and now dominate among patients requiring hospitalization [3, 4]. After exposure to the virus, typically through aerosol or droplet particles, SARS-CoV-2 binds to the angiotensin-converting enzyme 2 (ACE2) receptor, enriched on the surfaces respiratory [5-9] and intestinal epithelia [9]. Expression of ACE-2 on endothelium remains controversial, but some data supports that endothelial infection takes place [10-12].

The incubation period averages 3 (2-14) days [13-15], subject to host factors [16-21] and viral variant involved [22]. A recent metanalysis of 350 studies found that approximately a third of infected individuals remain asymptomatic [23] but can still shed virus and transmit the disease [24-27]. Most who do develop symptoms experience a mild disease course that may include fever, cough, myalgia, diarrhea, sore throat, and a loss of smell and taste [28, 29]. However, since the emergence of new variants and more rigorous testing, there has been a shift in the hospitalization risk. Between November 2020 and January 2021, the absolute risk of hospitalization overall was 4.7% in individuals testing positive for the alpha variant, reaching 21.4% in those over 80 years of age [30]. A more recent study, including over 43,000 SARS-CoV-2 positive individuals, approximately half of whom were asymptomatic, found a hospitalization rate of 2.3% following infection with the delta variant, which after adjustment, is twice the hospitalization risk when compared to the alpha variant [31].

Of those hospitalized, 20-30% [32] progress to acute respiratory distress syndrome (ARDS), which remains the leading cause of death. Among the 4.3%–22.5% of hospitalized patients [32-36], one to two-thirds of those requiring intensive care [37-39], and as many as 75% with COVID-19-associated ARDS may not survive [33].

Changes in patient management have had a significant impact on outcomes. Inpatient mortality reportedly decreased from 26% [40-42] at the beginning of 2020 to 7.6% [41] by mid-2020. Notably, much of this development is owed to improved outcomes in hospitalized patients who never progressed to mechanical ventilation (MV), whereby there has been little change in the prognosis of those with severe disease [36].

Vaccinations have reduced the risk of severe disease even more significantly. Recent CDC data showed that the risk of infection and hospitalization were 4.9 and 29.2 times lower in vaccinated when compared to unvaccinated individuals, respectively. When hospitalization did occur, progression to severe disease was significantly less likely in vaccinated patients [43].

The reported overall case fatality ranges from 0.4%-1% [30, 44], with individual risk determined by a relatively well-defined set of parameters [45, 46]. Patients at highest risk for disease progression are [47-53]:

- unvaccinated
- male
- of older age
- have comorbidities including obesity ($\text{BMI} \geq 30 \text{ kg/m}^2$), hypertension, diabetes
- have other chronic pre-existing conditions involving the cardiovascular, respiratory or renal systems

Moderate to severe COVID-19 is characterized by a dysregulated immune response resulting in a multisystem process dominated by endothelial activation and a prothrombotic state [54-56] and involving the cardiovascular, hepatic, renal and neurological systems [57-61]. The multisystem nature of the vascular involvement has been illustrated on whole body or lung PET-CTs of COVID-19 patients [62] and may even persist in survivors experiencing ongoing symptoms [63].

Therefore, a thorough understanding of the immunopathology in COVID-19 is critical for selecting the most appropriate therapeutic interventions and preventing patient exposure to unnecessary or potentially harmful treatments.

The **key immunologic processes of COVID-19** include:

- an initial rapid increase in viral load
- excessive and prolonged innate immune activation
- epi- and endothelial barrier dysfunction
- a pro-coagulant state
- excessive pulmonary neutrophil recruitment and formation of neutrophil extracellular traps (NETs)

These processes are also implicated in other infectious and inflammatory conditions. It remains to be determined if and to what extent the immune mechanisms observed in COVID-19 indeed differ from infectious and non-infectious conditions such as SIRS, inflammatory AKI, and other systemic hyperinflammatory states.

To classify disease severity and assist in standardizing of research protocols, the **WHO** has developed an **ordinal 9 point scale** (Figure 1) reflecting the various stages of disease progression [64, 65]. Applying this scale, this article attempts to match the underlying immunopathology of COVID-19 with evidence-based treatment modalities published in the peer-reviewed literature. We recognize that during the progression of the disease to severe COVID-19, these processes overlap, influence one another, and are causally linked. As the clinical picture evolves, different processes emerge and therapeutic targets change. Our knowledge of the immunopathology and therapeutic options in COVID-19 is expanding daily. Best up to date advice will be found online through resources, such as the regularly revised websites of the NIH and WHO.

1. WHO 9 point Scale, Patient Stage 0. No clinical or virological evidence of infection

Until vaccines achieve protection at a population level, social distancing, face masks, and hand hygiene are effective and necessary measures mitigating infection risk [66].

Over 114 vaccine candidates utilizing a diverse set of technologies are currently in clinical development[67]. Vaccination with mRNA constructs targeting influenza, rabies, zika or chikungunya virus have been subject to research efforts for some time and are now applied to SARS-CoV-2 [68-70]. Of those, two mRNA based vaccines, mRNA1273 from Moderna, Tozinameran from the BioNTech/Pfizer partnership and two adenovirus-vector vaccines, AZD1222 from AstraZeneca and the single-dose Janssen/Johanson & Johnson vaccine, have been granted Emergency Use Authorization (EUA) as COVID-19 vaccines in the US since December 2020.

In addition, an adjuvanted inactivated virus vaccine by Sinovac and the heterologous recombinant adenovirus vaccine Sputnik V have been in widespread use.

Vaccines provide high-level protection from SARS-CoV-2 infection and severe disease and elicit a robust antibody and B- and T- cell response [71, 72]. However, despite the effective initial humoral vaccine response, neutralization activity declines over time. To what extent serum antibody titers are a proxy for reinfection risk remains to be determined, but evidence for neutralizing activity and protection from (re)infection is emerging[73].

A recent large study demonstrated that antibody titers in response to the two most widely used mRNA vaccines decreased significantly after six months [74]. In addition, vaccine-induced efficacy against emerging viral variants is reduced [75, 76], supporting recent discussions for the need for booster vaccines.

In summary, the observation of breakthrough infections in vaccinated people, decreasing antibody titers following vaccination and emergence of new escape variants all highlight the ongoing need for close surveillance of this highly dynamic situation.

Based on published evidence, therapeutic recommendations include

Since vaccines have become available, other prophylactic measures have become less relevant. However, they may remain of importance for select high-risk individuals, especially when suboptimal vaccine responses may be expected, such as in the immunocompromised.

1. Vaccines

as discussed above

2. Casirivimab and Imdevimab

The use of the monoclonal antibody combination casirivimab plus imdevimab (see below) as post-exposure prophylaxis has been shown to result in a significant reduction of symptomatic SARS-CoV-2 infections compared with placebo (1.5% vs 7.8%; OR 0.17; $p < 0.001$) [77]. As a result of these findings, the Food and Drug Administration (FDA) has issued an Emergency Use Authorization (EUA) for this combination as post-exposure prophylaxis within seven days [78].

3. Topical Interferon-1 α

Type 1 Interferon is critically involved in the early antiviral response [79, 80] (see below). Prophylactic use of IFN-1 α nasal drops four times daily in 3000 uninfected health care workers (HCWs) resulted in no symptomatic SARS-CoV-2 infections in any of the patient-facing staff [81]. Controlled studies investigating the role of IFN-1 α in preventing COVID-19 are underway (NCT04552379, NCT04320238) [82].

Take home messages for this stage:

1. Social distancing, wearing face masks, eye protection and hand hygiene are effective measures mitigating an infection risk
2. Vaccination is the primary prophylactic measure. Until final data analysis of future phase III/IV trials are available, the duration of protection from clinical disease will remain undetermined.
3. Combination treatment of casirivimab and imdevimab is effective postexposure prophylaxis
4. Other prophylactic measures such as IFN-1 α and monoclonal antibody preparations might be of value in certain high risk groups

2. WHO 9 point Scale, Patient Stage 1. Infection, Ambulatory, no limitation of activities

During the incubation period, patients are asymptomatic, and many will never develop symptoms as described above. In others, epithelial infection and local inflammation may result in symptoms consistent with a mild viral infection [83].

As in most the disease does not progress further, the critical question here is if treatment is required at all and, if so, for whom.

High-risk patients should be monitored closely to initiate therapeutic interventions at the first signs of disease progression.

SARS-CoV-2 replication peaks early, *at symptom onset*, so the timing of virostatic therapies is critical. Delayed antiviral treatment may shorten viral shedding but not significantly affect the viral load (VL) [84]. Outpatients with a higher VL one week after symptom onset are more likely to be hospitalized and prolonged shedding of replication-competent virus is associated with more severe disease [21, 85, 86]. This suggests that early antiviral treatment may curb the rapid early replication and possibly influence the risk of disease progression.

Based on published evidence about this disease stage, therapeutic recommendations include

1. Antiviral therapy:

Nucleotide analogs - remdesivir, favipiravir, galidesivir and others [87] - mainly act by inhibiting the viral RNA-dependent RNA polymerase and thereby viral replication.

- a. **Remdesivir (RDV)** is an adenosine analogue initially developed as a treatment against Ebolavirus [88-91]. It is administered intravenously (iv.) as oral bioavailability is poor. Lipid analogues [92] and dry powder preparations for inhalation [93] addressing this shortcoming are under development. Treatment duration in trials range from 5 to 10 days, dosed at 200 mg OD on day one followed by 100mg. The primary dose-limiting effect is hepatotoxicity, and monitoring of liver function and coagulation is recommended.

Key trials assessing RDV use in COVID-19 [94] have limited enrolment to hospitalized patients. In ACTT-1 (Adaptive COVID-19 Treatment Trial), a double-blinded and placebo-controlled trial [95], RDV accelerated clinical recovery (10d vs 15d, $p < 0.001$) and reduced 28 day mortality, driven by patients at WHO stage 4 (HR 0.30 [0.14-0.64]) [95]. In SIMPLER-1 [96], five days of RDV in addition to standard of care was associated with clinical improvement at day 11 in hospitalized patients, mainly at WHO stage 3 (OR 1.65; [1.09-2.48], $p = 0.02$) [96, 97].

In the much larger WHO-led Solidarity trial (11,266 hospitalized patients of varying severity), RDV did not impact 28 day mortality (HR 0.95; [0.81-1.11] overall, HR 0.86; [0.67-1.11] not ventilated, HR 1.2; [0.80-1.80] ventilated), progression to MV or length of hospital stay. This included patients without oxygen requirement WHO stage 3, as well as 4f [98].

As a result of the above, the WHO no longer recommends RDV for the treatment of COVID-19 [99]. On the other hand, the NIH advises to include RDV for hospitalized patients receiving noninvasive O₂ supplementation or those at high risk for disease progression. An already initiated RDV course should be completed in patients progressing to WHO stages 5 and beyond [100]. Starting RDV in mechanically ventilated patients is not recommended.

- b. **Molnupiravir (EIDD-2801)** is currently undergoing phase II/III trials. Earlier work has shown effective inhibition of viral replication of SARS-CoV-2 *in vitro* and in animal models [101]. In two dose-escalation studies in outpatients with mild COVID-19, molnupiravir was safe, well-tolerated, and shortened viral shedding compared to placebo [102, 103]. While molnupiravir did not benefit hospitalized patients, a phase II/III study is currently investigating its impact on hospitalization rate, clinical characteristics and mortality in outpatients with mild to moderate COVID-19. Its oral bioavailability may be an asset in the ambulant setting [104].
- c. **Favipiravir** has been evaluated in mild to moderate COVID-19 patients, most not requiring oxygen [105], was well-tolerated, and accelerated viral clearance. It is now undergoing further study in outpatients [106].

Novel antiviral agents continue to be developed [107], such as PF-07304814, a SARS-CoV-2 protease inhibitor for which phase 1 results are awaited (NCT04535167). Several agents are in pre-clinical development, and more data is likely to become available over the following months.

2. Blocking (co)-receptors, preventing viral entry into host cells.

- a. Recombinant human ACE2 (**rhACE2**) receptor [108] as decoy therapy has been used, to some encouraging effect, in a small case series of patients with non-COVID-19 associated ARDS [109], suggesting a mechanism of action other than viral neutralization. Instead, rhACE2 may restore homeostasis of the ACE2/Ang1-7/MasR system, as lack of ACE2 mediates both epi- and endothelial inflammation (see below). Concerns for negatively impacting pulmonary autoregulation have not been substantiated [482].
- b. In addition to ACE2 binding, viral entry requires proteolysis of the spike protein by the host-enzyme TMPRSS2 [110], which is androgen-dependent, which may account for some of the observed risk

disparity. Serine protease (TMPRSS2)-inhibitors such as **nafamostat** and **camostat mesylate** [111, 112] are being explored for use in mild COVID-19 [113]. The latter expedited recovery by 40% in outpatients with mild disease by day five [114] but had no impact on clinical improvement, admission rate to intensive care or mortality in hospitalized patients [115]. Since nafamostat also inhibits fibrinogen proteolysis, it has been proposed as a short-acting anticoagulant at later disease stages [116-118]. Single reports of cerebral bleeds on this treatment require careful consideration [483].

- c. **Maraviroc**, an inhibitor of chemokine receptor CCR5, is used widely in HIV therapy. Maraviroc inhibits the viral SARS-CoV-2 protease *in vitro* [119]; and is currently being evaluated in phase II trials in ventilated COVID-19 patients (300mg BD for 14days, NCT04441385, NCT04435522) as well as in patients with moderate disease (NCT04710199). Animal data suggest that this compound may also have additional benefits by reducing neutrophil recruitment to the lung in severe COVID-19 [120].

None of these treatments is recommended outside clinical trials yet.

3. Anti-SARS-CoV2 monoclonal antibody preparations

While recommended in the beginning of the pandemic, bamlanivimab and etesevimab, the recent emergence of escape variants has led to their replacement by new antibody preparations.

- a. REGN-CoV2 contains two anti-spike receptor-binding-domain (RBD)-antibodies, **casirivimab** and **imdevimab**. In SARS-CoV-2 positive outpatients, one dose accelerated viral clearance and symptom resolution (13 vs 6 days) among seronegatives [121]. The effect on seroconverted individuals was less pronounced. An RCT assessing 2.4g or 8g of casirivimab/imdevimab, administered within 7 days from symptom onset in SARS-CoV-2-positive outpatients reduced medically attended visits in the combined treatment group compared to placebo by half (6% vs. 3% overall), and from 15% to 6% in seronegative patients [122]. Data indicates that REGN-CoV2 benefits outpatients with mild COVID-19, who are at risk for disease progression.
- b. Results for **VIR-7831** (Sotrovimab), a monoclonal antibody with Xtend technology prolonging its half-life and expected to enhance pulmonary absorption, has been assessed in SARS-CoV-2 infected outpatients with mild or moderate illness (COMET-ICE trial). A single i.v. dose of 500mg resulted in a subsequent reduction of relative risk for hospitalization or death by 85% compared to placebo ($p=0.002$) [123].
- c. **Nanobodies** are antibody fragments consisting of a single monomeric variable antibody domain occurring naturally in camelids and sharks. Nanobodies with a high affinity for spike protein, effectively competing with ACE-2 and recognizing epitopes that are structurally not accessible to conventional antibodies are being explored as neutralizing antiviral agents, currently at the pre-clinical stage [124, 125].

4. Hydroxychloroquine (HCQ)

Among its many anti-inflammatory and anti-thrombotic effects, HCQ interferes with viral uptake and intracellular transport by altering the endosomal pH. However, HCQ failed to demonstrate an impact on clinical outcome or survival in exposed presymptomatic individuals [126, 127], including those with mild disease [128, 129], those hospitalized with or without O₂ requirement and with severe COVID-19 [130, 131]. HCQ prolongs the QT interval, which, particularly in patients with underlying cardiac problems, is another argument against its widespread use [132]. Two meta-analyses on the effect of HCQ in combination with Azithromycin demonstrated an increase in mortality among hospitalized patients (RR 1.27 [1.04-1.54]; RR 1.11 [1.02,1.20]) [134, 133].

5. Ivermectin (IVM)

This anthelmintic agent has received attention as an inhibitor of intracellular viral transport *in vitro*, however at MICs well above what would be safely achievable *in vivo* [135]. Heterogeneity of data available has complicated their interpretation [137]. A recent meta-analysis of 10 RCTs in 1173 patients

evaluating its use in COVID-19 has not identified a clinical or survival benefit [136]. The use of IVM is not recommended outside of clinical trials.

Take home messages for this stage:

1. Timing of antiviral therapies is likely critical but due to lack of data no recommendations for their use in outpatients can be made
2. Post exposure prophylaxis with selected anti-SARS-CoV2 monoclonal antibody preparations are recommended in high risk individuals
3. Agents blocking (co)-receptors, preventing viral entry into host cells remain under investigation with some having shown clinical benefit
4. Hydroxychloroquine has failed to demonstrate any clinical or survival benefit for all disease stages
5. The use of ivermectin is not recommended outside of clinical trials

3. WHO 9 point Scale, Patient Stage 2. Infection, Ambulatory, limitation of activities

At this disease stage, patients may display signs of a lower respiratory tract infection or mild pneumonitis with cough and fever.

Alveolar macrophages (AMs) are the first line of defense and respond to PAMP/TLR signaling triggered by infected alveolar epithelial cells (AEC)[138]. Both produce pro-inflammatory cytokines (IL1 β , IL8, IL18, TNF α , IFN γ) and chemokines (CXCL2) that recruit peripheral immune cells to the lung. Epithelial infection also downregulates regulatory ligands, removing the tolerizing epithelial interaction with, and disinhibiting, AMs [139, 140].

The viral receptor ACE2 is part of the ACE2/Angiotensin-(1-7)/MAS axis of the Renin-Angiotensin-System[141], which counteracts the pro-inflammatory and vasoconstrictive effects of Angiotensin 2 (AT2) by cleaving it to Ang1-7 [142]. After binding SARS-CoV-2, ACE2 is internalized [143-145], and AT2 will accumulate as a result. Mediated by the Angiotensin 2 receptor 1 (AT1R)[146-148], AT2 upregulates endothelial adhesion molecules, facilitates leukocyte recruitment [141, 149], and polarizes macrophages towards a pro-inflammatory M1 phenotype [150-153]. The conversion of AT2 by ACE2 into anti-inflammatory Ang1-7 is impaired, and excess AT2 damages epi- and endothelial integrity through its inflammatory, vasoconstrictive and pro-fibrotic effects [154]. ACE2 downregulation induced by SARS-CoV-2 infection exacerbates a pro-inflammatory state, causing lung damage that may exceed the initial viral cytopathic effect [144, 145].

The key questions at this disease stage are:

- a. **how likely the patient will progress to more severe disease based on his/her risk profile and**
- b. **which biomarkers should be measured to assess the risk for progression**

Most risk scores have been validated in hospitalized patients, and little is available to help with stratifying risk in outpatients [155-157].

An acuity score predicting hospitalization, intensive care admission, or mortality risk in COVID-19 patients based on 30 parameters performed well. Blood pressure, respiratory rate and SaO₂ were the most relevant predictors, feasible in most outpatient settings [158].

Biomarkers indicative of innate immune cell activation and epithelial damage are now useful to predict disease progression. CCR5, IL1ra and IL10 may predict a severe disease course up to a week prior to clinical deterioration [159]. Until such specific biomarkers become widely available, it is important to consider vital signs and laboratory parameters that are accessible without delay. These include hsTroponin, proBNP, IL-1, LDH, transaminases, renal function, inflammatory markers and coagulation

testing which indicate early extrapulmonary end organ involvement and have been shown to assist with clinical assessment and guide management decisions (discussed below).

Based on published evidence about this disease stage, therapeutic recommendations include:

1. Antiviral therapy

As discussed above, antivirals may theoretically be of benefit but have not been sufficiently studied in outpatients. The development of RDV preparations for inhalation in outpatients considered at risk of progression may add therapeutic options before admission becomes necessary [93, 160].

2. Anti-SARS-CoV2 monoclonal antibody preparations

The recommendations for the use of anti-SARS-CoV2 monoclonal antibody preparations as discussed above apply for this disease stage as well.

3. Interferon III (IFN- λ)

IFN- λ is exclusively expressed by respiratory and gastrointestinal epithelium. Hematopoietic cells lack IFN- λ receptors, and therefore it has little systemic pro-inflammatory effect. With a favorable safety profile observed in phase II hepatitis D trials [161,162], IFN- λ seems an attractive candidate for COVID-19 therapy. Initial data on IFN- λ use in outpatients (180mcg once daily) showed accelerated viral clearance if IFN- λ was administered within five days of symptom onset compared to placebo [163]. Others, administering IFN- λ within three days of symptom onset, did not find such benefit [164]. The side effect profile was favorable, with transient transaminitis being the main reported adverse event.

4. Budesonide

GCs may downregulate ACE2 in respiratory epithelium [165] and reduces airway inflammation, possibly impacting the beginning of epithelial and macrophage-driven host response. The STOIC trial of an age-stratified cohort with mild COVID-19 symptoms for less than seven days. Intervention was open-label, 800mcg Budesonide dry powder inhalation BID until symptom resolution compared to SOC. Medically attended visits and hospitalizations were fewer (14% vs 1%; $p=0.004$), and symptom resolution faster (7 vs 8 days, $p=0.007$) [166]. The treatment was well-tolerated, encouraging larger placebo-controlled trials that target mildly affected outpatients.

- **Convalescent plasma (CP)**

CP has been widely administered to patients with COVID-19, often with advanced disease. Patients may have already seroconverted and have neutralizing anti-SARS-CoV2 concentrations equivalent to those contained in CP [167] (Table 1). CP may contain pro-inflammatory and pro-coagulant factors [168]. Further, SARS-CoV2 specific antibody titers vary greatly [169]. Antibody kinetics in COVID-19 differ: nonsurvivors have a delayed antibody response, whereas survivors produce neutralizing antibodies more rapidly [170]. Based on this observation and considering the abovementioned caveats, the *timing* of exogenous antibody administration seems critical.

As the majority of studies on CP use have been uncontrolled, it is not surprising that efficacy assessments of a metanalysis including 30 studies and RCTs with 17,225 patients [171] were inconclusive (“very uncertain”) and found no effect on mortality or clinical improvement at 28 days.

CP outside of clinical trials is no longer recommended, except for patients with impaired humoral immunity. A recently published open-label RCT on CP use in 921 hospitalized patients was terminated early for futility. The risk for intubation or death by day 30 did not differ (32.4% in the CP group, 28.0% in the SOC group; RR 1.16; [0.94–1.43] $P=0.18$) and patients receiving CP experienced more serious adverse events (33.4% versus 26.4%; RR = 1.27, 95% CI 1.02–1.57, $P=0.03$) [172].

- **AT1R blockers, ACE-inhibitors (ACEi)**

This drug class was initially hypothesized to impact COVID-19 outcomes either by restoring homeostasis of the ACE2/Ang1-7/Mas-R system; or conversely by upregulating tissue-resident ACE2. A metanalysis

of 21 studies[173] did not support a difference in risk of death (pooled OR 1.29 [0.89-1.87] $p=0.18$) or disease severity (pooled OR 0.94 [0.59-1.50] $p=0.81$) in patients who had been receiving ACEi when contracting SARS-CoV-2. Since then, several studies assessing the impact of discontinuing ACEi treatment upon COVID-19 diagnosis have not identified a difference in disease severity or death. Discontinuation of ACEi/ARB treatment in those already using these agents is therefore not justified.

- ***Azithromycin (AZM)***

Besides its antimicrobial properties, AZM has immunomodulatory effects. It repolarizes macrophages towards tissue-restorative M2 and inhibits pro-inflammatory NF κ B and STAT1 signaling [174, 175]. However, in patients with a moderate oxygen requirement (WHO stage 4), AZM did not impact progression to MV or death [176]. As macrolides prolong the QTc interval, their use should be carefully monitored, especially in older patients or in combination with other pro-arrhythmogenic agents. Most studies have investigated AZM in combination with HCQ and repeatedly identified an increased mortality risk associated with this combination [133]. AZM is therefore not recommended in the treatment of COVID-19.

Take home messages for this stage:

1. Risk assessment in mildly symptomatic outpatients should integrate demographic factors, extent of respiratory symptoms, neutrophil/lymphocyte ration, inflammatory markers and biomarkers of extrapulmonary tissue injury
2. Anti-SARS-CoV-2 monoclonal antibody preparations are recommended in high risk individuals
3. Inhaled budesonide in ambulatory patients not requiring oxygen may be beneficial but requires more detailed assessment
4. Evidence does not support the use of azithromycin and, especially in combination with HCQ, may inflict harm.

4. WHO 9 point Scale, Patient Stage 3 Hospitalized, no O₂ requirement

Hospitalization becomes necessary in approximately 4.7% of infected individuals. The risk in patients over 60 years is higher – approximately between 10 and 20% [30]. The decision to admit patients not requiring O₂ will be informed by a comprehensive assessment of clinical, laboratory and imaging findings [177], with more pro-active management of risk groups and the availability of healthcare resources.

Several clinical scores have been developed to distinguish those at risk for disease progression at the time of hospitalization (**Table 2**). A moderately accurate prediction of future severe COVID-19 disease can be achieved by combining the results of CT findings of the lung, inflammatory markers (C-reactive protein, ferritin, neutrophils, lymphocytes, albumin), evidence of tissue injury (transaminases, LDH, Troponin, D-Dimer) and evidence of electrolyte imbalance (blood urea, electrolytes)[178]. Lymphopenia and neutrophilia, expressed as elevated NLR (neutrophil/lymphocyte ratio) on admission are consistently associated with disease progression and death [179, 180].

A metanalysis of 5699 patients showed that an elevated NLR on admission increased the risk of death almost threefold (RR2.74 [0.98-7.66][181]. Leukocytosis, elevated LDH, procalcitonin, and transaminitis were associated with increased risk of ICU admission and death [32], while lymphopenia, elevated CRP and fibrinogen on admission predicted an O₂ requirement [182]. Another metanalysis including 4969 patients found that neutrophilia and lymphopenia on admission was associated with a significantly increased risk of progression to severe COVID-19 (OR 7.99; 1.77-36.14 resp. OR 4.2; 3.46-5.09,) and death (OR 7.87; 1.75-35.4, resp. OR 3.71; 1.63-8.44) [183].

Biomarkers that may be helpful to assess risk for disease progression at this stage reflect activation of innate immunity, immune cell recruitment, and beginning damage to epithelial and endothelial barriers and tissue injury.

Blood samples of COVID-19 patients show significantly higher levels of circulating endothelial cells (CECs) on admission than those with other respiratory infections, demonstrating **early and extensive endothelial injury** [184]. Epithelial and endothelial damage begins long before a patient is admitted to the ICU, and CECs, if available, may be of prognostic value now [185]. Other markers of endothelial activation with discriminatory value at this stage are von Willebrand Factor (vWF), angiopoietin (Angpt-1/Angpt-2 ratio, see below) and soluble urokinase plasminogen activator receptor (suPAR). Early discharge and mild disease trajectory have been predicted by a suPAR of $\leq 2\text{ng/mL}$ with high specificity [186].

Of all cytokines measured in over 1400 COVID-19 patients at hospitalization [187], IL-6 and TNF α levels independently predicted disease severity and death, outperforming CRP, D-Dimers and ferritin. Higher CRP, IL-6, IL-8, IL-10, TNF α and IL-2R levels on admission were found in those patients later progressing to critical illness and/or death [188].

Hospitalization and progression to severe disease could also be predicted by a decision algorithm integrating demographic risk factors and comorbidities with immune cell profiling [189].

At this stage, replicating virus may rarely be present in blood [190, 191]. Viremia and RNAemia in COVID-19 increase the risk of critical disease and death six to elevenfold [192-194].

Considering more widely available markers, the combination of elevated LDH, CRP and decreased lymphocyte counts predicted ten-day mortality [195]. The combined analysis of the patient's age, CD4⁺ lymphocyte counts and LDH was a clinically useful composite for disease progression (AUC 0.92) [196]. In summary, markers of inflammation (CRP, ferritin), cardiac (troponin, BNP), epithelial (Angpt-2) and endothelial injury (CECs), combined with pre-existing clinical risk factors, may provide the best assessment for disease progression. Angpt-2 and CECs may also be helpful biomarkers in patients at risk for disease progression before an O₂ requirement develops but may not be widely available.

The Lung Injury Prediction Score (LIPS) assessed the risk of ARDS at time of hospitalization in a variety of conditions [197-201]. Even though not validated for COVID-19 ARDS, its positive predictive value for this indication was enhanced significantly when Angiopoietin 2 (Angpt-2), CRP, and the FiO₂/SpO₂ ratio within 6h of admission were included.

Multiorgan involvement, including coagulopathy, myocardial, liver, intestinal and kidney injury, may all precede respiratory manifestations [202, 203]. Myocardial injury on admission in particular predicts poor outcome, especially if both troponin and proBNP are elevated. Higher troponin levels on admission are accompanied by higher D-Dimers, fibrinogen, creatinine, WBC, and procalcitonin levels, reflecting organ involvement beyond the respiratory and cardiac systems.

In a metaanalysis published by Figliozzi et al., evidence of acute cardiac injury was by far most predictive for poor outcome (OR 10 [5-22.4]), followed by renal injury and low platelet and lymphocyte count [204]. Metadata from 10 clinical studies generated two predictive equations including CRP, neutrophil, lymphocyte count +/- D dimer, resulting in a sensitivity of 0.76 (0.68) and specificity of 0.79 (0.83) when applied to a cohort of patients [205].

Future works must emphasize parameters that predict deterioration at a time point when therapeutic interventions can counteract disease progression. Based on a recent UK study on COVID-19 patients presenting to the emergency department, strict implementation of simple clinical observations while considering demographic risk factors outperforms the prognostic value of laboratory biomarkers [206].

Finally, a recent study reports that Anti-DNA and anti-phosphatidylserine antibodies, determined at hospital admission, correlated strongly with progression to severe disease (PPV 85.7% and 92.8%). Antiphospholipid antibodies have been observed in COVID-19 patients since the very beginning of the pandemic [207]. This suggests that autoantibodies following the initial viral insult contribute to the pathology at later stages of COVID-19.

Based on published evidence about this disease stage, therapeutic recommendations include:

1. Antiviral therapy.

The WHO no longer recommends antivirals for hospitalized patients. NIH guidelines however suggest that RDV may be used in hospitalized patients at high risk of disease progression with or without oxygen requirement (WHO stage 3, 4).

2. Corticosteroids:

RECOVERY assessed dexamethasone in hospitalized patients of varying severity. There was no benefit seen in patients who did not require ventilatory support (OR 1.19; 0.91-1.55) [208] or in those with early disease (symptom duration <7days) [208]. Concerns for early steroid use would include immunosuppression at a time when viral replication may still be very active [209, 210]. In a metanalysis of five RCTs including 7692 patients, steroid use in patients without O_2 requirement was even associated with an increased mortality risk (RR 1.23 [1.00-1.62]; $p=0.05$) [211]. In summary, there is presently no evidence to support the use of steroids at WHO stage 3.

2. Interferons

Interferons (IFN), produced by lymphocytes (Type II: IFN- γ) and epithelia (Type III: IFN- λ) are some of the most effective antiviral defense mechanisms. Type I IFNs (IFN α , IFN β) initiate an antiviral response through their receptors INFR1/2, widely expressed on epithelial, endothelial and myeloid cells. INFR engagement activates Janus Kinase (JAK1), which mediates inflammation and antiviral effects [212]. While the use of a pro-inflammatory signaling molecules seems counterintuitive initially, *the timing of IFN-I administration in relation to viral replication is critical*. The replication of SARS-CoV-2 is reported to peak already at symptom onset. A rapid IFN-I response controls viral replication, whereas a delayed IFN-I rise results in excessive inflammation and tissue damage instead [82, 213-215].

In critically ill COVID-19 patients, IFN-1 α and β responses are impaired, virus persistence is prolonged and systemic inflammatory markers are comparatively high [216, 217]. SARS-CoV-2 produces only a weak early IFN-I response *in vitro* [217]. A suppressed early IFN-I response may allow viral replication to peak unopposed and contributes to the excessive inflammation seen in patients with severe disease [213, 214]. It follows that exogenous IFN-I should be beneficial early, while delayed administration could easily be harmful [218].

Results of important IFN trials are summarized in **Table 3**. The Solidarity trial assessed IFN- β 1a therapy at WHO stages 3-6. It failed to demonstrate a survival benefit overall and suggested worse outcomes among ventilated patients.

Three trials in hospitalized patients (WHO stages 3-5) treated with either IFN- β 1b s.c. for two weeks or nebulized IFN- β 1a resp. IFN- α 2b within five days of admission suggested accelerated clinical improvement, reduced ICU admissions and lower mortality [219]. Treatment more than five days after admission however *increased* mortality (aHR 0.05 [0.01-0.37] early treatment, 6.8 [1.41-40.8] $p=0.2$ late treatment) [220].

In a phase II placebo-controlled study of nebulized IFN- β 1a [221] in hospitalized patients, at WHO stages 3 and 4, IFN treatment still reduced the risk of severe disease or death significantly even though median symptom duration was ten days (OR 0.21 [0.04-0.97]; $p=0.046$). IFN-I may therefore retain a benefit for longer than suggested, at least in the noncritically ill [222].

3. Heparin

The International Society for Thrombosis and Hemostasis (ISTH) recommends low molecular weight heparin prophylaxis for all hospitalized patients with COVID-19 and supports its continuation for 2-6 weeks following discharge [223, 224].

The benefit of heparinization leading to improved organ support free survival in noncritically ill hospitalized patients has now been backed up by results from ATTACC/ACTIV-4a/REMAP-CAP and CORIST studies (see below). In the noncritically ill hospitalized group, therapeutic anticoagulation may be superior to prophylactic dosing, but more data is required [225, 226].

- ***Anti-SARS-CoV2 monoclonal antibody preparations***

Monoclonal antibodies failed to demonstrate a benefit in hospitalized patients [227, 228], and are no longer recommended regardless of oxygen requirement, except in patients with humoral immunodeficiency [229].

Take home messages for this disease stage:

1. Patient risk stratification for disease progression is a critical step during this disease stage. Clinical risk scoring systems could assist this, in conjunction with immune cell profiling, imaging results and appropriate biomarkers
2. Interferon therapy, administered within 3-5 days of admission, may be of benefit at this stage but more evidence is needed for a recommendation to be made.
3. Heparin prophylaxis should be initiated in all hospitalized patients with COVID-19
4. The use of GCs and monoclonal antibodies at this stage is not recommended

5. WHO 9 point Scale, Patient Stage 4 Hospitalized, O₂ requirement by mask or nasal prongs

The reported rate of patients progressing to stage 4 varies widely, but a large proportion of those admitted will require oxygen supplementation. Mortality in this group can be significant, even in those not dyspneic at presentation [230].

In a subset of patients, the controlled antiviral response transitions to a dysregulated immune response during this WHO stage, possibly even earlier. The clinical presentation is now characterized by ongoing respiratory epithelial and endothelial damage, followed by excessive recruitment of activated innate and adaptive immune cells. The most relevant immunopathologic processes, which in our opinion characterize stage 4 and overlap in many aspects with stages 3 and 5, are outlined below.

a. Disrupted AT2/ACE2 homeostasis

The downregulation of ACE2 in cells infected by SARS-CoV2 leads to elevated AT2 levels, vasomotor disturbance, increased ventilation-perfusion (V/Q) mismatch (ventilation of non-perfused lung areas), microcapillary leaks, and epithelial apoptosis [143-145]. AT2's pro-inflammatory effects via NFκB [141] enhance leukocyte-endothelial interactions through upregulation of ICAM-1 and VCAM-1, setting the stage for NETosis and thrombotic complication (see below) [231, 232].

b. Macrophage activation and polarization

Monocytes and macrophages are key elements of the early antiviral response, dominate the developing dysregulated inflammatory process, and are the drivers for cytokine excess, neutrophil and lymphocyte recruitment, development of barrier dysfunction and tissue fibrosis [233, 234].

Depending on their environment, macrophages exist on a spectrum from pro-inflammatory M1, responsible for pathogen killing, production of reactive oxygen species (ROS) and proinflammatory cytokines (IL1b, TNFα, IL6, IL18) [235], to M2 cells with a focus on phagocytic activity, promoting immune tolerance, fibrosis and tissue repair [236-238]. Non-inflammatory removal of apoptotic

immune cells, *efferocytosis*, is a unique feature of M2 macrophages [239]. Activated alveolar macrophages (AM) [138, 140] recruit bone-marrow derived monocytes to the lung [240, 241], where they adopt an M1 phenotype, complementing the antiviral response but also amplifying tissue damage [242] and initiate massive neutrophil recruitment and activation of Th1 and Th17 cells [243]. Histopathology of autopsied lungs of patients with COVID-19 ARDS implies a crucial role for macrophage activation and the subsequent neutrophil migration [244, 245]. The persistence and prolonged activation of M1 macrophages result in an excess of pro-inflammatory mediators, reactive oxygen species, enzymes and accumulating cellular debris all of which is detrimental to epi/and endothelial integrity [235, 246-248]. Once the inflammatory stimulus is removed, M1 must revert to M2 macrophages to begin a “clean up and repair program” and deactivate the previous “pro-inflammatory program”. Otherwise, the inflammatory process will persist [249, 250]. One of the factors inhibiting the repolarization to M2 is netosis, thereby exacerbating tissue damage [251].

c. Activation of the VEGF-Angpt-1/2-Tie2 system

High Angpt-2 levels predict ICU admission at the time of hospitalization [252]. Patients with Angpt-2 levels above 5000pg/mL were 10 times more likely to require ICU care (OR 9.33 [2.35-44.9]). Angpt-2 was the only blood parameter correlating with compliance measures during MV (mL/cmH₂O, $r = -0.46$, $p = 0.01$) and renal function, emphasizing the prognostic relevance of biomarkers of endothelial activation and microvascular damage during this stage.

Pulmonary neutrophil recruitment may be associated with further significant clinical deterioration and escalation of respiratory support [244]. Therefore, a high NLN as well as markers of epithelial and endothelial damage (low VEGF2R levels and low Angpt-1/2 ratio (see below) is expected to have prognostic value at this stage [202, 253-256].

Based on published evidence about this disease stage, therapeutic recommendations include:

1. Antiviral therapy: see recommendations as detailed under prior WHO stages

2. Steroids: GCs have many anti-inflammatory properties, including the repolarization of macrophages towards M2 and inhibition of neutrophil recruitment [257, 258].

The RECOVERY trial yielded landmark data on the role of GCs in COVID-19, and its results emphasize the importance of timing of therapeutic interventions. It studied hospitalized patients at WHO stages 3, 4 and 5 treated with **dexamethasone** (6mg OD i.v./p.o.), for 10 days (n= 2104) to SOC (n=4321) and demonstrated a 28 day survival benefit in mechanically ventilated (29.3% vs 41.4%; HR 0.64 [0.51-0.81]) or O₂ dependent patients at WHO stage 4/5 (23.3% vs 26.3%; HR 0.82[0.71-0.94]); but no benefit in those without O₂ requirement (11.8% vs 14.0%; HR 1.19 [0.91-1.55]) [208]. GCs were only beneficial if the symptom duration was longer than 7 days.

A metanalysis of seven studies (n=1703) [259] addressed GCs in COVID-19 patients with an at least moderate O₂ requirement; most were ventilated. GCs decreased the 28 day mortality (HR 0.66 [0.52-0.83], $p < 0.001$), in those mechanically ventilated or on noninvasive ventilation (noninvasive O₂: HR 0.41 [0.19-0.88]; MV: HR 0.69 [0.55-0.86]), whereas *patients requiring inotropes did not benefit* (HR 0.55 [0.34-0.88] vs 1.05 [0.65-1.69]; $p = 0.06$). Another metanalysis of 7692 patients similarly identified a benefit of steroids, limited to patients requiring MV (RR 0.85 [0.72; 1.00, $p = 0.05$)] [211]. In summary, data is consistent showing that steroids are beneficial at later disease stages, in patients requiring oxygen or MV (see below).

3. IL-6 inhibition

Increased IL-6 expression by monocytic cells in COVID-19 [260] provides a rationale for the use of IL-6 blockers (Sarilumab, Siltuximab, Tocilizumab (TCZ)). An IL-6 level of >30pg/mL at hospitalization indicated a future need for MV in a cohort of 146 patients [261].

Table 4 summarizes relevant studies on IL-6 inhibition in hospitalized patients specific to WHO stages at recruitment. The results indicate in most that risk of progression to MV is decreased when IL-6 inhibition is initiated at WHO stage 4 or 5.

Recovery has been the largest trial investigating IL-6 inhibition [265]. It recruited hospitalized patients mainly at WHO stages 4, 5 and 6, most (82%) received concomitant GCs. In patients at stage WHO 4 and 5, 28-day mortality (HR 0.81 [0.67-0.99]; HR 0.96 [0.74-1.00]), respectively and the risk of progression to MV was reduced (15% vs 19%; HR 0.79; 0.69-0.92; $p=0.002$). At WHO stage 6, a survival benefit was not as evident (HR 0.93 [0.74-1.18])[265] and overall was only present when GCs were given concomitantly (RR 0.79 [0.7-0.89] vs 1.16 [0.91-1.48]).

A recent metanalysis of 27 trials including 10,930 patients at WHO stages 3, 4, 5, IL-6 blockade (TCZ $n=18$, sarilumab $n=9$) compared to placebo or SOC confirmed these findings. 28-day mortality (22% vs 25%; OR 0.86 [0.79-0.95]) and risk of progression to MV were both reduced in the IL-6 inhibitor group. Again, the benefit was limited to a combination with GCs (OR 0.78 [0.63-0.98]). IL-6 blockade alone did not achieve a mortality reduction (OR 1.09 [0.91-1.30]) [266]. Another more recent metanalysis of 28 cohort studies and 8 RCTs showed again that the risk for progression to MV was reduced (RR 0.84 [0.76-0.93]) in WHO stages 4 and 5 and a survival benefit limited to those receiving concomitant GCs [267].

In summary, Tocilizumab is recommended *in combination with steroids* for recently hospitalized patients at WHO stage 4-5, with rapid disease progression or who require MV for less than 24 hours [268].

A double-blinded RCT including 457 and 1365 patients randomized and treated in phases 2 and 3, respectively, assessed the use of sarilumab. Among the 20% of phase 3 patients receiving MV, a third of whom also received steroids, the proportion with ≥ 1 -point improvement in clinical status at day 22 was 43.2% for sarilumab and 35.5% for placebo (RRR 24.7%). In analyses combining phase 2 and 3 patients requiring MV, the mortality risk was reduced, though non-significantly (HR 0.76; [0.51 to 1.13]). Again patients receiving GCs concomitantly showed more pronounced risk reduction (OR 0.49 [0.25 to 0.94]).

4. IL-1-inhibitors:

IL-1-inhibitors in the form of the endogenous receptor antagonist IL-1ra (anakinra) or as monoclonal antibody against IL-1 β (canakinumab) showed promise in cohort and observational studies [269-274] that triggered further investigations. Evidence remains controversial, but the timing of administration yet again seems crucial.

A randomized trial [275] compared the addition of anakinra to SOC in patients at WHO stage 4ff. No difference was seen between the groups in mortality by 28 days (22% vs 24%, aHR 0.77 [0.33-1.77]), oxygen wean, or time to discharge.

When patients requiring oxygen were randomized to receiving anakinra within ≤ 4 days from admission, early treatment reduced 28-day mortality by 74% (aHR 0.26 [0.1-0.66], $p<0.001$) compared to SOC. No survival benefit was seen in patients not in the early treatment group who may have received anakinra as late rescue therapy (aHR 0.82, $p=0.7$). These results allow some attribution of benefit to use at earlier disease stages [276] and illustrate how critical the clinical status at the time of treatment allocation is. A recent metanalysis of IL-1 inhibition in COVID-19 could not proceed due to the data heterogeneity between studies [277]. A suPAR level of >6 ng/mL heralds the development of respiratory failure in COVID-19 [278] and may assist biomarker-guided IL-1 inhibition [279].

Two recent studies failed to demonstrate a benefit of IL-1 inhibition with canakinumab compared to SOC. Patients were included at WHO stages 4 and 5, and neither MV free survival nor risk of COVID-19 related death differed significantly [280]. Additional reasons for the lack of canakinumab benefit in COVID-19 are likely based on the pharmacokinetic profile of this drug and its selective inhibition of IL-1 β , leaving IL-1 α unopposed [281].

At present, pending further data collection, IL-1 inhibition is not recommended as SOC in COVID-19 management.

5. *Janus-kinase-inhibitors (JAK inhibitors)*

Many immune reactions responsible for the inflammatory response in COVID-19 (including IFN-1a,b) are transcriptionally regulated by the JAK-STAT pathway [282, 283]. A meta-analysis [284] of five studies investigating JAK inhibition in COVID-19 demonstrated a significant reduction in mortality (HR 0.12 [0.03-0.39]), and ICU admission (OR 0.05 [0.01-0.26]). **Table 5.**

In two early studies in hospitalized patients, most of whom with an O₂ requirement but not requiring MV, treatment with **Baricitinib**, an oral JAK1/JAK2 inhibitor, for seven days on LPV/r +/- HCQ background, demonstrated a faster reduction in O₂ requirement and a lower mortality rate (1/20 (5%) vs 25/56 (45%) compared to SOC [285]. A follow-up study mainly included patients at WHO stages 3/4 [286]. Here, the need for intensive level care at 14 days was significantly reduced in the treatment group, and patients were more likely to be discharged by two weeks (77.8% vs 12.8%, $p < 0.0001$).

TACTIC-R [289] is assessing the combination of baricitinib with **ravulizumab** (a C5 inhibitor) in WHO stages 3-5. Although treatment with **roxolitinib**, an oral JAK1/2 inhibitor, was shown to be safe, it did not reduce mortality or progression to MV in patients at WHO stages 4 and 5 [290].

In a recent study assessing **tofacitinib** in the treatment of hospitalized patients at WHO stages 3, 4 and 5 (including high flow O₂ only) [291], the cumulative incidence of death or respiratory failure through day 28 was reduced by 37% (RR 0.63; [0.41 to 0.97] $p = 0.04$). All-cause mortality was observed in 2.8% of tofacitinib and 5.5% of placebo-treated patients, but the effect was not significant (HR 0.49; 95% CI, 0.15 to 1.63). Serious adverse events were not significantly more common in the treatment group (14.1% vs 12.0%). Potential safety concerns for JAKi include a rise in creatinine kinase, transaminases, and myelosuppression, which may increase the risk of opportunistic infections. The complete blood count should be monitored during treatment.

6. *TNF α inhibitors (TNFi)*

Data on the use of TNFi in COVID-19 is limited. In a small study including seven patients, three of which were already mechanically ventilated, Infliximab at a dose of 5mg/kg iv on days one and three [292], resulted in a rapid decrease of pro-inflammatory cytokines and a clinical improvement in six of seven patients. In comparison, the mortality rate in the 17 control patients at a similar stage of hospitalization was 35%. The ACTIV trial (NCT04513940) recruits hospitalized patients with moderate to severe COVID-19 (WHO stage 4ff) and will, in addition to infliximab, assess abatacept and cenicriviroc, an inhibitor of chemokine receptors CXCR2 and CCR5, for this indication.

7. *GM-CSF inhibition – or supplementation?*

GM-CSF, among other functions as overall pro-inflammatory cytokine and growth factor, polarizes macrophages towards M1 and upregulates integrin expression by neutrophils, mediating their adhesion to and migration across endothelium. Higher serum levels of GM-CSF in ARDS correlate with a higher risk of death [293]. Antagonizing GM-CSF, therefore, appears to be an attractive target in COVID-19 [213]. The best time for GM-CSF inhibition, based on immunopathology, would be *prior* to the recruitment of peripheral monocytes. GM-CSF inhibition has an established safety record [294], but neutropenia, alveolar proteinosis, and impaired viral clearance remain concerns. In addition, lack of GM-CSF inhibits phagocytosis, efferocytosis by M2 macrophages and impairs the removal of NETs which may delay macrophage repolarization.

Conversely, GM-CSF is critical for AM survival, surfactant removal, epithelial protection and the antiviral response. Higher GM-CSF levels in ARDS bronchoalveolar lavage fluid are associated with better outcomes [295-297], contrasting the association of higher serum levels with a worse prognosis [298, 299]. Despite initial concerns for excessive granulocyte mobilization and recruitment of neutrophils to the lung [295], first data assessing inhaled GM-CSF (sargramostim 125mcg, BD, for 5 days) in hypoxemic patients are encouraging [300].

Addition of sargramostim for five days to SOC in patients at WHO stages 4 and 5 was associated with a P(A-a)O₂ improvement by $\geq 33\%$ compared to SOC alone (54% vs 26%, $p < 0.001$, NCT04326920). In a second cohort, including patients at WHO stage 4 and those requiring high flow oxygen but not NIV, oxygenation was also improved (treatment group 84%, SOC group 64% $p = 0.02$) [301].

Amplifying pulmonary neutrophil recruitment might worsen the patient's respiratory status. Under this premise, GM-CSF receptor blockade is also under investigation in COVID-19. **Mavrilimumab** (i.v. 6mg/kg once) showed some promise in a small prospective cohort study from Italy in patients at WHO stages 4 and 5 [302].

A double-blinded RCT recruited 40 patients in WHO stages 4 and 5 ($n = 21$ receiving mavrilimumab) and found no significant difference in mortality or oxygen wean to placebo. However, mortality was high overall (43% and 53%, respectively) [303]. An ongoing study comparing mavrilimumab to placebo in hospitalized patients at WHO stages 4 and 5 reported in an interim analysis of $n = 166$ that MV-free survival was higher in the treatment arm (86.7% vs 74.4%, $p = 0.1$), equivalent to a 65% risk reduction, with final results outstanding [485].

8. Interventions targeting NETosis. Netosis is probably one of the most important yet underrecognized mechanisms in the pathophysiology of COVID-19. The release of Neutrophil Extracellular Traps, or NETosis, is a defense system utilized by neutrophils against bacteria, viruses or protozoa. During the formation of neutrophil extracellular traps (NETs), the neutrophil nuclear membrane is dissolved and NETs consisting of chromatin, citrullinated histones (CitH3), neutrophil elastase (NE) and oxidative enzymes such as myeloperoxidase are released into the extracellular space [304-306]. Excessive NETosis damages epithelial [307] and endothelial [308] cells. NET removal by two extracellular enzymes, DNase I and DNaseIL3, expressed by dendritic cells and macrophages, is critical for tissue homeostasis [309]. NETs promote M1 persistence in COVID-19 and delay macrophage repolarization, which prevents the degradation of cellular debris by M2, facilitated by C1q [251]. As a result, efferocytosis, a hallmark feature of M2 cells, cannot occur effectively. Pro-inflammatory cytokines continue to be released, which prevents a timely switch to tissue-restorative repair processes [247, 248, 310]. NETs are also highly prothrombotic. They entrap erythrocytes and platelets and can form intravascular NET clots [309, 311]. Autopsies of COVID-19 victims show this featuring thrombotic occlusion of pulmonary, cardiac, renal, and hepatic vasculature by aggregated NETs [312, 313]. NETosis can be quantified by measuring specific biomarkers (cell-free DNA, myeloperoxidase [MPO]-DNA, and citrullinated histone H3 [Cit-H3]) [314]. These correlate closely with SOFA scores [315, 316] and may be useful for risk stratification at earlier disease stages.

Dornase alfa is commonly used in inhaled form for patients with cystic fibrosis where it cleaves extracellular DNA, mainly from leukocytes, thereby decreasing the viscosity of respiratory secretions [317]. Beneficial effects on recovery in small case series in critically ill COVID-19 patients with ARDS have been published, additional trials are underway [318], [319], [320, 321]. Other DNase enzymes for the treatment of hospitalized patients with acute moderate to severe SARS-CoV-2 infection are currently in development.

9. Heparin

The ATTACC trial compared therapeutic-dose heparinization as an initial strategy in *noncritically* ill patients, most at WHO stage 4 with SOC thromboprophylaxis. There was a trend favoring therapeutic-dose heparinization (survival to discharge: 76.4% vs 80.2%), exclusive to this earlier disease stage [225], but more data is required.

10. 2-deoxy-2-Glucose:

2-DG was granted EUA by the Indian authorities for moderate and severe COVID-19 when faced by the overwhelming pandemic impact on the Indian subcontinent [322].

It inhibits glycolytic ATP production and is used to sensitize tumor tissue to chemo- and radiotherapeutic agents. 2-DG administration followed by low dose radiation was suggested as a means to reduce lung inflammation in COVID-19 [323]. The agent accumulates in metabolically active, virus-infected cells and results in their apoptosis. Phase 3 trials recruited patients at WHO stage 4ff, without adding radiation. Early oxygen wean was more frequently possible (42% vs 31%), but more evidence to support this treatment is needed, and detailed data on safety is lacking.

Take home messages for this disease stage:

1. Data strongly support the use of GCs at this stage. Careful monitoring for secondary infections in these patients is critical.
2. JAK-inhibitors offer a benefit in terms of preventing progression to MV and survival
3. IL-6 inhibition, in combination with GCs, is recommended at this and later disease stages
4. While results from larger trials with IL-1 inhibitors are lacking, data available from observational cohorts suggests that they may have a benefit on clinical outcome and survival in this but not later disease stages.
5. The administration of GM-CSF antibodies can currently not be recommended while the use of inhaled GM-CSF may be of benefit at this and later stages
6. Enzymatic therapy with DNase 1 or recombinant DNase 1L3 to counteract Netosis may play an important role in preventing progression of COVID-19 in this disease stage. However, data of clinical trials are still pending.

6. WHO 9 point Scale, Patient Stage 5: Noninvasive ventilation or high flow oxygen

Driven by inflammatory cell recruitment and barrier dysfunction, patients at this stage have progressed to severe pneumonia, and their gas exchange is more severely affected. They require high flow oxygen, and approximately one fifth will require noninvasive pressure support [324].

The three main immunologic mechanisms during this stage include:

1. Disruption of endothelial and epithelial integrity

Worsening capillary leakage and alveolar edema now contribute to poor gas exchange [325, 326].

The main determinants of endothelial and epithelial permeability are the VEGF and Ang/Tie2 systems.

The primary stimulant of VEGF production by AECs is IL-1 β [327-329]. Under normal physiologic conditions, pulmonary VEGF levels of capillary and alveolar lumens are strictly compartmentalized [330]. During an infection with SARS-CoV-2 this compartmentalization is lost, resulting in worsening epithelial damage [331] and release of alveolar-side VEGF into the bloodstream across the damaged barrier [332]. This promotes endothelial Angpt-2 release [331], amplifying capillary leakage [333].

Therefore, an increase of VEGF in the alveolus (as detectable in bronchoalveolar lavage fluid) indicates improved barrier function and predicts recovery from ARDS [334] while increasing plasma levels are associated with worsening pulmonary edema [335, 336].

Angpt-1 is the main agonist of the endothelial Tie2 receptor [337, 338]. Their interaction seals endothelial tight junctions and protects against capillary leakage [339-345]. It opposes Angpt-2 action on Tie2 [342, 346], which increases capillary permeability [342, 347, 348] and leads to epithelial apoptosis [325, 346, 349-353].

Increased Angpt-2 and low VEGF2R levels in plasma predict ARDS severity and 28d mortality [336]. In mechanically ventilated patients, serum Angpt-2 correlates with the severity of pulmonary vascular leakage and predicts the likelihood of ICU admission, development of ARDS and resulting fatality in COVID-19 [252, 354-360]. A low Angpt-1/Angpt-2 ratio is a marker for endothelial dysfunction and a consistent feature of adverse outcomes in sepsis, DIC and ARDS [361-369].

2. Neutrophil Recruitment and Amplification of Inflammation

Much of COVID-19-associated inflammatory pulmonary damage is mediated by M1 macrophages and the neutrophils they recruit [55, 370-372]. Neutrophilia, especially in the BAL fluid, is a consistent feature of severe COVID-19 and predicts mortality [28, 190, 202, 253, 255, 373-375]. Autopsies of COVID-19 patients have demonstrated the accumulation of neutrophils and M1 macrophages associated with microangiopathic and thrombotic changes in pulmonary capillaries [376-378]. Especially in patients who require respiratory support, the neutrophil population contains immature, lower density granulocytes (LDGs) [256]. LDGs are ineffective phagocytes [256, 312, 379-382], produce large amounts of pro-inflammatory cytokines (IL17, IFN-I) and have a propensity to form NETs [383, 384].

CXCL5 concentration in BAL fluid correlates with the extent of neutrophil infiltration of lung parenchyma [385, 386].

The damaged alveolar epithelium, in turn, activates the endothelium, which upregulates adhesion molecules [387-389], and mechanically entraps primed neutrophils [390-392]. This close interaction with the activated endothelium activates the neutrophils, which causes them to release inflammatory mediators, form NETs [312, 390] and enter the alveolus [371].

In summary, neutrophils home to the COVID-19 lung, interact with the damaged endothelium and contribute to tissue damage. Because of NETosis-induced impairment of macrophage repolarization, efferocytosis is defective. Accumulating NETs may not be adequately removed and sustain inflammation and neutrophil recruitment, further exacerbating inflammatory tissue injury.

3. Immune thrombosis

Thromboembolism complicates up to a third of COVID-19 admissions to ICU [393-397]. Generalized endothelial damage and thrombotic microvascular injury of lungs, kidneys, liver and heart and frequent pulmonary embolism [396] and stroke [398], characterize severe disease.

Evidence for endothelial dysfunction is present as early as WHO stage 3. Levels of FVIII, vWF:Ag, D-Dimers at the time of hospitalization correlate with risk of thromboembolic complications and mortality in COVID-19 patients [182, 399, 400].

Not all markers of endothelial damage have equal prognostic value, and more data are required in this area. Thrombomodulin, selectin, Angpt-2 and CEC levels were all significantly elevated in patients with more severe COVID-19, but in a comparative analysis, only vWF antigen discriminated disease severity of outpatients, non-critical (WHO stage 3,4,5) and critical (WHO stages 5,6,7) COVID-19 [401, 402]. Other selected markers of endothelial damage may predict inpatient mortality (glycocalyx damage (AUC 0.74), ADAMTS13 (AUC 0.72) and VEGFA (AUC 0.73)), but will not be readily accessible to most clinicians [403].

4. Complement activation

The complement system has antiviral properties [404] but can also result in tissue injury through activation of Netosis and pro-coagulant effects. The pivotal role of complement activation in COVID-19 was identified early [405]. Histopathology of skin, kidney and lung biopsies from COVID-19 patients (n=5) showed extensive deposition of C5b-9 in the microvasculature [406]. Complement pathways are highly induced in the COVID-19 lung, which correlate with disease severity [407-409].

Based on published evidence about this disease stage, therapeutic recommendations include:

1. Antiviral therapy: remains indicated as discussed above

2. Steroids: remains indicated as discussed above

3. Heparin: remains indicated as discussed above

4. Cytokine inhibitors: As discussed above, IL-6 inhibition can be expected to be of benefit. The data for IL-1 inhibition is less clear but on balance would favor earlier use (WHO stage 4)

5. JAK inhibitors: Based on the ACTT-2 and COV-barrier results, JAK inhibition has most impact at this stage.

ACTT-2, a double-blinded, placebo-controlled RCT enrolled over 1,000 inpatients at WHO stage 4ff to assess efficacy and safety of **baricitinib** 4mg OD for 14 days in addition to RDV versus RDV alone. Patients receiving GCs were excluded. Baricitinib addition made progression to MV or death less likely (HR 0.69; [0.5-0.95]). Patients on high flow O₂ or NIV (WHO stage 5) benefitted most. Here, time to clinical recovery was shortened from 18 to 10 days and clinical improvement by two weeks was twice as likely (OR 2.2 [1.4-3.6]). In patients at WHO stage 3, 4 or 6 however, baricitinib did not impact time to recovery. Secondary infections were less frequent in the treatment arm [287].

The COV-barrier trial [484], a recently published double-blinded, placebo-controlled phase 3 RCT assessed baricitinib in addition to SOC among hospitalized COVID-19 patients, over 90% of who also received GCs. Overall 28-day mortality in the treatment group was significantly reduced (8% vs 13%, HR 0.57 [95% CI 0.41–0.78], $p=0.002$), and clinical improvement at day 4 through 14 was more likely. Patient at WHO stage 5 (NIV or high flow O₂) again benefited most (28-day mortality HR 0.52 (95% CI 0.33–0.80); $p=0.006$). The baricitinib benefit was maintained in those who did not receive concomitant GCs or RDV, and persisted when mortality risk was re-analyzed at 60 days (HR 0.62 [0.47-0.83] $p=0.005$).

In summary, baricitinib appears to have its most significant benefit at WHO stage 5. It is currently recommended in combination with remdesivir only which, given recent evidence, may be revised [288].

6. Angiopoietin 2 inhibitors, VEGF inhibitors

Vanucizumab, a bispecific monoclonal antibody directed against Angpt-2 and VEGF, usually used as an angiogenesis inhibitor in solid tumors [410], is currently in ongoing trials in COVID-19.

Similarly, inhibition of VEGF as the main factor stimulating Angpt-2 release may be of value, especially as it enters the circulation in severe lung injury. **Bevacizumab**, a monoclonal VEGF-A antibody, has now been repurposed for use in COVID 19 (NCT04270414; NCT04305106) in patients meeting ARDS criteria.

In a study of 26 patients, treatment with i.v. bevacizumab resulted in improved PaO₂/FiO₂ within 24h and rapid normalization of inflammatory markers [411]. However, the clinical status of the cohort was very diverse, complicating the interpretation of these findings. A case series in COVID-19 patients requiring ICU level care [412] included $n=25$ receiving bevacizumab, and $n=21$ receiving a combination of TCZ/ bevacizumab. 23/25 (93%) of bevacizumab treated individuals recovered to discharge, as did 14/21 patients receiving a combination treatment. Dosing and WHO stages of the patients were not reported, and more research is required before an assessment of its benefit can be made.

7. Tie-2 mimetics: Vasculotide, a Tie2 mimetic improved survival in animal models of viral pneumonia and ARDS and reduced pulmonary edema and endothelial apoptosis [413-415]. Clinical trials investigating AV-001/Vasculotide and similar products in human ARDS and COVID-19 are planned.

8. Complement inhibition: Monoclonal antibodies targeting specific complement factors, **eculizumab** and **ravulizumab** inhibiting C5, or AMY-101 inhibiting C3, are currently undergoing assessment in COVID-19 studies. So far, available data is limited to uncontrolled smaller case series.

At WHO stage 5ff (>6L/min O₂ requirement, severe pneumonia, or ARDS), eculizumab 900 mg on D1, 8, 15, and 22 in addition to SOC was associated with lower 28-day mortality (7/35 (20%) vs 23/45 (51%), $p=0.005$), and respiratory support could be weaned faster [416]. A trial assessing ravulizumab in 122 patients with severe COVID-19 (WHO stage 6ff) was halted after interim analysis did not support efficacy [419]. Assessment of patients not yet requiring MV (WHO stage 5) is being evaluated.

Selective C5a inhibition in severe COVID-19 has been investigated by Vlaar and colleagues[420]. C5a is a strong chemoattractant of neutrophils, leads to endothelial activation and is central to neutrophil tissue-factor dependent pro-coagulant activity [421, 422]. Administration of seven i.v. doses of C5a inhibitor **vilobelimab** in 15 patients with severe COVID-19, mainly at WHO stages 5 and 6 did not impact early

oxygen wean or mortality compared to SOC (aHR 0.65 [95% CI 0.10–4.14]). Thromboembolic complications though were less frequent (2/15 vs 6/15). Given these initial results, vilobelimab is undergoing further assessment in severe COVID-19 (NCT04333420). In summary, despite some studies showing rapid decline of inflammatory markers, sufficient evidence supporting the use of complement inhibitors outside of clinical trials is lacking [416–418]

9. Statins: Statins inhibit MyD88, upstream of NF κ B, and have several anti-inflammatory and immunomodulatory effects. Earlier metadata suggests a risk reduction of 30% for progression to severe COVID-19 or death with the use of statins [423]. A more recent metaanalysis including seven retrospective cohort studies (2398 patients) found that COVID-19 patients taking statins had nearly 40% lower odds of progressing to the composite endpoint of severe/critical illness or death (OR: 0.59; [0.35–0.99]). This was even more pronounced in patients taking statins pre-admission (OR 0.51 [0.41–0.64]). The addition of simvastatin to SOC in patients with ARDS due to a variety of pathologies showed that only those with a hyperinflammatory phenotype, defined by IL-6 and sTNF α levels, benefited from statins. In this subgroup, the improvement achieved in 28 day mortality and ventilator-free survival was significant [424]. While this does not address whether or not adding statins acutely would be of benefit, these findings may be relevant to future research on COVID-19 related ARDS.

10. Imatinib is a Bcr-Abl tyrosine kinase inhibitor and approved chemotherapeutic agent for Philadelphia chromosome positive CML and ALL. Experimental and early clinical evidence suggests that imatinib protects the integrity of the vascular barrier [425, 426]. It has been studied in severe COVID-19 with the rationale of mitigating damage to the barrier of the alveolo-capillary unit. In a double-blinded placebo-controlled RCT [427], 400 patients at WHO stages 4ff were assigned to either placebo or imatinib at a loading dose of 800mg followed by 400mg OD for nine days. Three-quarters of participants received concomitant GCs, a fifth RDV; no other immunomodulatory agents were used. Time to discontinuation of MV or oxygen wean did not differ, while time spent on MV was shorter (survivors 7 vs 12 days, $p=0.02$) and 28-day survival improved (mortality risk aHR 0.52 [0.26–1.05]; $p=0.068$).

Take home messages for this disease stage

1. Risk stratification based on clinical findings and biomarkers is critical
2. Currently available data strongly support the use of GCs in patients at this disease stage.
3. Heparin: remains indicated as discussed above
4. JAK inhibitors remain indicated as discussed above
5. Although data remain limited, monoclonal antibody directed against Angpt-2 and VEGF may play a role in preventing the progression to MV in this disease stage
6. IL-6 inhibitors are recommended under certain conditions at this stage
7. Data is not sufficient to recommend the use of complement inhibitors or imatinib at this disease stage, but new data on a potential role for these agents is emerging

7. WHO 9 point Scale, Patient Stage 6 – Intubation and Mechanical Ventilation

At this stage, patients progress from requiring high flow oxygen to intubation and MV. The clinical deterioration at this stage is a direct consequence of the inflammatory and immunologic mechanisms initiated at stages 3 and 4 that are now leading to respiratory failure.

In over 10,000 hospitalized COVID-19 patients from Germany, mortality was 53% among those who progressed to MV, compared to 16% who did not [324, 428].

Autopsy results in mechanically ventilated patients who had rapidly progressed to severe respiratory failure demonstrated neutrophilic invasion of the alveolar spaces and microvasculature, epithelial injury and microthrombi, likely related to excessive neutrophil recruitment to the lung [244].

Based on published evidence about this disease stage, therapeutic recommendations include:

1. Steroids are beneficial in COVID-19 patients requiring MV (see under WHO stage 4).

2. Antibiotic and Antifungal treatment:

Prolonged immunosuppression in the critically ill must be navigated with caution. Secondary bacterial and fungal superinfections frequently complicate severe COVID-19, and patients must be closely monitored. Increasingly, COVID-19 associated invasive mycoses are being recognized, due to profound lymphopenia, prolonged significant illness, and immunosuppressive therapies [429].

3. Heparin

There is a high incidence of isolated pulmonary artery thrombi in critically ill COVID-19 patients suggesting the possibility that some thrombotic events in these patients are formed in situ rather than representing dislodged emboli [430]. While thromboembolism is very common in COVID-19, heparinization does not completely abolish this risk [431-433], and thromboembolic events despite prophylactic, and even therapeutic heparinization occur.

Biomarkers of NETosis such as cell-free DNA are significantly elevated in patients at WHO stage 5. Many factors contribute to the prothrombotic state in severe COVID-19, with NET formation and antiphospholipid antibodies emerging as important contributors [312]. Lastly, heparin resistance is not uncommon in severe COVID-19 [434], and alternative strategies for anticoagulation may have to be pursued, such as direct thrombin inhibition with argatroban [435].

There was early recognition that anticoagulation should be administered in COVID-19 patients, but heparin dosing has been controversial [436, 437] (Table 6). The International Society on Thrombosis and Haemostasis (ISTH) suggests risk stratification with dose escalation to intermediate (50% increase of prophylactic dose) for those with a BMI ≥ 30 or very high D-Dimers (≥ 3000) and *discourages the use of therapeutic doses for primary prevention* [222]. The ATTACC/ACTIV-4a/REMAP-CAP trial [438], where therapeutic anticoagulation was *inferior to usual care thromboprophylaxis* in the outcome of organ-support free survival, with a higher incidence of major bleeding complications, lends support to this approach. This sets critically ill COVID-19 patients apart from those with moderate illness (WHO stages 3,4,5) in whom therapeutic heparinization was not inferior (see above).

4. Aspirin (ASA)

ASA has a favorable anti-inflammatory effect on the neutrophil-platelet-endothelial interaction which results in microthrombi, VQ mismatch, and NETosis.

The data on treatment with ASA in non-COVID-19 ARDS in at-risk individuals is controversial [439, 440]. One study even showed an association with an increased risk of MI, VTE and stroke [440].

5. IL-6 Inhibitors

In addition to the use of IL-6 inhibitors as discussed under WHO stage 4, siltuximab (in one to two doses) was used in a small cohort study including 30 patients on either NIV support or MV matched to patients receiving SOC [441]. The majority received concomitant GCs (18/30). The 30-day mortality rate was significantly lower in the treatment group (HR 0.46, 95% CI 0.22–0.97; $p=0.04$). Though not all patients had completed the follow-up period, 16/30 were discharged, four remained on mechanical ventilation, and ten patients died. This contrasts the findings of the much larger Recovery trial on TCZ, and evidence on siltuximab will have to be revisited as more information becomes available.

6. IL-1 Inhibitors

In a cohort study comparing TCZ, Sarilumab and anakinra in patients at stages 5 and 6, IL-1 and IL-6 inhibition improved long-term (180 days) survival when initiated before the establishment of severe ARDS ($\text{PaO}_2/\text{FiO}_2 < 100$ mmHg). Notably, in this cohort that did not co-medicate patients with GCs, all three agents offered a survival benefit in patients requiring MV (180-day mortality risk. Anakinra HR 0.47 [0.26-0.87], sarilumab HR 0.55 [0.25-1.22], TCZ HR 0.57 [0.28-1.14]). In patients with severe

ARDS, the survival advantage offered by sarilumab and tocilizumab was lost (TCZ HR 1.02 [0.37-2.81], sarilumab HR 0.69 [0.25-1.75]), and while the efficacy of anakinra was reduced, it was still superior to SOC (HR 0.46 [0.22-0.94])[442].

Take home messages for this disease stage:

1. The use of GCs in patients with COVID-19 has been found to be most beneficial for patients in this disease stage. Careful monitoring for secondary infections in these patients is critical
2. Starting antiviral therapy in this disease stage is no longer recommended
3. Heparin at prophylactic dose remains indicated. A proposed risk stratification guiding heparin dosing is discussed above. The additional use of ASA and NSAIDs cannot be recommended
4. The use of IL-6 inhibitors may be beneficial
5. Drugs targeting Netosis might be critical in this disease stage but data from clinical trials are still pending.
6. Despite limited data the use of complement inhibitors for this stage cannot be recommended

8. WHO 9 point Scale, Patient Stage 7 – Ventilation and additional organ support

Stages 6 and 7 are pathophysiologically similar and characterized by gradual deterioration of widespread endothelial damage [443]. Approximately 33% of hospitalized patients may progress to COVID-19 associated ARDS [444].

Acute respiratory distress syndrome (ARDS) is the result of dysregulated inflammation in response to a pulmonary or systemic insult that impacts the endothelial and epithelial integrity of the alveolocapillary unit [445]. Clinical data suggest ARDS endotypes with distinct clinical features and disparate outcomes [445, 446]. The clinical course of ARDS is described as occurring in two stages [361, 447]:

- a. an inflammatory exudative phase characterized by alveolar-epithelial damage, recruitment of inflammatory cells with subsequent alveolar flooding with proteinaceous fluid, formation of hyaline membranes, and resultant hypoxemic respiratory failure (week 1-2)
- b. a fibroproliferative phase characterized by lung fibrosis and vascular remodeling (week 2-3ff)

The Berlin ARDS criteria define an international diagnostic standard [447, 448].

COVID-19 associated ARDS, as evidenced by autopsy studies, is consistently characterized by

- extensively affected microcirculation, alveoli infiltrated with neutrophils and/or monocytes/macrophages
- peripheral neutrophilia and decrease of most lymphocyte subsets (i.e., a high NLR), correlating with poor outcome, higher sequential organ failure assessment (SOFA) scores and death [28, 190, 202, 253, 255, 373, 374].
- a highly inflammatory pulmonary response, often in combination with ongoing viral RNA presence
- extensive diffuse alveolar damage
- widespread endothelial damage and thromboembolic events

The pandemic has put a spotlight on the fact that despite therapeutic advances, the overall mortality of ARDS remains unacceptably high[36]. Therefore, the most critical strategy in COVID-19 management is addressing the evolving inflammation-mediated tissue damage early. Ventilatory strategies, fluid balance and positioning are the most important points and foundations of ARDS management once it occurs but are well beyond the scope of this review. Pharmacologically, in addition to steroid administration, the therapeutic focus shifts to addressing the epithelial and endothelial barrier dysfunction – especially if the Angpt2/1 ratio or circulating VEGFR2 levels remain elevated. The patient's prognosis may be reflected in

NLR, coagulation parameters, D-Dimers, von Willebrand factors, Troponin, BNP, renal and liver function, CECs (circulating endothelial cells), and NETosis markers such as cell free DNA (see above).

Based on published evidence about this disease stage, therapeutic recommendations include:

Treatment recommendations in this disease stage are essentially identical to those for WHO stage 6.

1. **Steroids** are of benefit in COVID-19 patients who are mechanically ventilated

2. **Antibiotic and Antifungal treatment:** Prolonged immunosuppression in the critically ill will have to be navigated with caution. Secondary bacterial and fungal superinfections frequently complicate severe COVID-19. Patients must be closely monitored for secondary infections. Increasingly, COVID-19 associated aspergillosis (CAPA) is being recognized, resulting from profound lymphopenia, and as a complication of immunosuppressive therapies [449].

2. **Statins:** as discussed at WHO stage 5

3. **Mesenchymal stem cells (MSC):**

The use of MSC in severe ARDS is experimental and only included here for completeness and to introduce this novel treatment concept. It is a common misperception that MSCs in ARDS replace damaged alveolar cells. In fact, the proposed clinical benefit is ascribed to their immunomodulatory properties, skewing macrophages to M2, and exerting an antifibrotic effect. Available data is minimal. The COVID-19 Treatment Guidelines Panel of the NIH recommends against the use of mesenchymal stem cells for the treatment of COVID-19 outside of clinical trials.

Take home messages for this disease stage:

1. Ventilatory strategies, fluid balance and positioning are the most important points and foundations of ARDS management
2. The use of GCs are of benefit at this disease stage.
3. Due to prolonged immunosuppression and the critical condition of patients in this disease stage, active surveillance for secondary infections and antibiotic and antifungal treatment play an important role
4. Heparin remains indicated at prophylactic dose, with some data indicating that therapeutic dosing may inflict harm
5. The initiation of antiviral therapy in this disease stage is no longer recommended
6. The use of MSC in this disease stage is experimental and evidence insufficient to recommend it

9. Summary

Therapeutic options for patients with COVID-19 are rapidly evolving, and knowledge gained from currently ongoing clinical trials may change future treatment recommendations. We believe that sound treatment decisions are based on a thorough understanding of the immunopathology of COVID-19. This understanding will enable clinicians to develop a well-defined treatment strategy based on clinical risk scores, immune cell profiling, disease-stage specific biomarkers, laboratory and imaging findings. We recognize that during disease progression, pathological processes overlap, influence each other, and new ones may emerge. Especially at earlier disease stages, treatment target the prevention of a dysregulated hyperinflammatory state. We believe this occurs at the latest at WHO stage 4 in predisposed individuals. Once patients require mechanical ventilation, treatment becomes increasingly challenging with fewer effective treatment options and a higher risk of adverse outcomes. Consequently, a disease-stage specific treatment selection should not be made empirically but follow published evidence from the literature as summarized above.

References

1. Zhu N, Zhang D, Wang W, et al. A Novel Coronavirus from Patients with Pneumonia in China, 2019. *N Engl J Med*. 2020;382(8):727-33.
2. World Health Organization. WHO Coronavirus (COVID-19) Dashboard 2021 [Available from: <https://covid19.who.int>].
3. European Center for Disease Prevention and Control a. Data on SARS-CoV-2 variants in the EU/EEA 2021 [updated 09. Sept 2021. Available from: <https://www.ecdc.europa.eu/en/publications-data/data-virus-variants-covid-19-eueea>].
4. Centers for Disease Prevention and Control. COVID Data Tracker. 2021. Available from: <https://www.cdc.gov/coronavirus/2019-ncov/variants/index.html>]
5. Wan Y, Shang J, Graham R, et al. Receptor Recognition by the Novel Coronavirus from Wuhan: an Analysis Based on Decade-Long Structural Studies of SARS Coronavirus. *J Virol*. 2020;94(7).
6. Magrone T, Magrone M, Jirillo E. Focus on Receptors for Coronaviruses with Special Reference to Angiotensin- Converting Enzyme 2 as a Potential Drug Target - A Perspective. *Endocr Metab Immune Disord Drug Targets*. 2020;20(6):807-11.
7. Hou YJ, Okuda K, Edwards CE, et al. SARS-CoV-2 Reverse Genetics Reveals a Variable Infection Gradient in the Respiratory Tract. *Cell*. 2020;182(2):459-471.
8. Zhao Y, Zhao Z, Wang Y, et al. Single-Cell RNA Expression Profiling of ACE2, the Receptor of SARS-CoV-2. *Am J Respir Crit Care Med*. 2020;202(5):750-9.
9. Zhang H, Kang Z, Haiyi G.. The digestive system is a potential route of 2019-nCov infection: a bioinformatics analysis based on single-cell transcriptome. 2020 [Available from: Preprint at <https://www.biorxiv.org/content/10.1101/2020.01.30.921806v1>]
10. Hamming I, Timens W, Bulthuis ML, et al. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *J Pathol*. 2004;203(2):631-7.
11. Nascimento Conde J, Schutt WR, Corbunova EE, et al. Recombinant ACE2 Expression Is Required for SARS-CoV-2 To Infect Primary Human Endothelial Cells and Induce Inflammatory and Procoagulative Responses. *mBio*. 2020;11(5).
12. Monteil V, Kwon H, Prado P, et al. Inhibition of SARS-CoV-2 Infections in Engineered Human Tissues Using Clinical-Grade Soluble Human ACE2. *Cell*. 2020;181(4):905-13 e7.
13. Guan WJ, Ni ZY, Hu Y, et al. Clinical Characteristics of Coronavirus Disease 2019 in China. *N Engl J Med*. 2020; 382:1708-20.
14. Li Q, Guan X, Wu P, et al. Early Transmission Dynamics in Wuhan, China, of Novel Coronavirus-Infected Pneumonia. *N Engl J Med*. 2020;382(13):1199-207.
15. Lauer SA, Grantz KH, Bi Q, et al. The Incubation Period of Coronavirus Disease 2019 (COVID-19) From Publicly Reported Confirmed Cases: Estimation and Application. *Ann Intern Med*. 2020;172(9):577-82.
16. Gniazdowski V MP, Wohl S et al. Repeat COVID-19 Molecular Testing: Correlation with Recovery of Infectious Virus, Molecular Assay Cycle Thresholds, and Analytical Sensitivity. *medRxiv*. 2020. Available from: <https://doi.org/10.1101/2020.08.05.20168963>
17. Singanayagam A PM, Charlett M et al. Duration of infectiousness and correlation with RT-PCR cycle threshold values in cases of COVID-19, England, January to May 2020. *Euro Surveill*. 2020;25(32).
18. Jefferson T SE, Brassey J, Heneghan C. Viral cultures for COVID-19 infectivity assessment: Systematic review. *Clin Infect Dis*, 2020; ciaa1764, <https://doi.org/10.1093/cid/ciaa1764>
19. Wolfel R, Corman VM, Guggemos W, et al. Virological assessment of hospitalized patients with COVID-2019. *Nature*. 2020;581(7809):465-9.

20. Basile K MK, Carter I et al. Cell-based culture of SARS-CoV-2 informs infectivity and safe de-isolation assessments during COVID-19. *Clin Infect Dis*. 2020 Oct 24:ciaa1579. doi: 10.1093/cid/ciaa1579. Epub ahead of print.
21. van Kampen JJA, van de Vijver DAMC., Fraaij PLA, et al. Shedding of infectious virus in hospitalized patients with coronavirus disease-2019 (COVID-19): duration and key determinants. *Nat Commun* **12**, 267 (2021).
22. Homma Y, Katsuta T, Oka H, et al. The incubation period of the SARS-CoV-2 B.1.1.7 variant is shorter than that of other strains. *J Infect*. 2021;83(2):e15-e7.
23. Sah P, Fitzpatrick MC, Zimmer CF, et al. Asymptomatic SARS-CoV-2 infection: A systematic review and meta-analysis. *Proc Natl Acad Sci U S A*. 2021;118(34).
24. Wei WE, Li Z, Chiew CJ, et al. Presymptomatic Transmission of SARS-CoV-2 - Singapore, January 23-March 16, 2020. *MMWR Morb Mortal Wkly Rep*. 2020;69(14):411-5.
25. He X, Lau EHY, Wu P, et al. Temporal dynamics in viral shedding and transmissibility of COVID-19. *Nat Med*. 2020;26(5):672-5.
26. Arons MM, Hatfield KM, Reddy SC, et al. Presymptomatic SARS-CoV-2 Infections and Transmission in a Skilled Nursing Facility. *N Engl J Med*. 2020;382(22):2081-90.
27. Kimball A, Hatfield KM, Arons M, et al. Asymptomatic and Presymptomatic SARS-CoV-2 Infections in Residents of a Long-Term Care Skilled Nursing Facility, King County, Washington, March 2020. *MMWR Morb Mortal Wkly Rep*. 2020;69(13):377-81.
28. Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet*. 2020;395(10229):1054-62.
29. Yang X, Yu Y, Xu J, et al. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. *Lancet Respir Med*. 2020;8(5):475-81.
30. Nyberg T, Twohig KA, Harris RJ, et al. Risk of hospital admission for patients with SARS-CoV-2 variant B.1.1.7: cohort analysis. *BMJ*. 2021;373:n1412.
31. Twohig KA, Nyberg T, Zaidi A, et al. Hospital admission and emergency care attendance risk for SARS-CoV-2 delta (B.1.617.2) compared with alpha (B.1.1.7) variants of concern: a cohort study. *Lancet Infect Dis*. 2021.
32. Zhang JJY, Lee KS, Ang LW, et al. Risk Factors for Severe Disease and Efficacy of Treatment in Patients Infected With COVID-19: A Systematic Review, Meta-Analysis, and Meta-Regression Analysis. *Clin Infect Dis*. 2020;71(16):2192-205.
33. Potere N, Valeriani E, Candolero M, et al. Acute complications and mortality in hospitalized patients with coronavirus disease 2019: a systematic review and meta-analysis. *Crit Care*. 2020;24(1):389.
34. Rodriguez-Morales AJ, Cardona-Ospina JA, Gutierrez-Ocampo E, et al. Clinical, laboratory and imaging features of COVID-19: A systematic review and meta-analysis. *Travel Med Infect Dis*. 2020;34:101623.
35. Navaratnam AV GW, Day J, Wendon J, Briggs TWR. Patient factors and temporal trends associated with COVID-19 in-hospital mortality in England: an observational study using administrative data. *Lancet respir Med*. 2021.
36. Yeates EO, Nahmias J, Chinn J, et al. Improved outcomes over time for adult COVID-19 patients with acute respiratory distress syndrome or acute respiratory failure. *PLoS One*. 2021;16(6):e0253767.
37. To KK, Tsang OT, Leung WS, et al. Temporal profiles of viral load in posterior oropharyngeal saliva samples and serum antibody responses during infection by SARS-CoV-2: an observational cohort study. *Lancet Infect Dis*. 2020;20(5):565-74.
38. Lim ZJ, Subramaniam A, Ponnappa Reddy M, et al. Case Fatality Rates for Patients with COVID-19 Requiring Invasive Mechanical Ventilation. A Meta-analysis. *Am J Respir Crit Care Med*. 2021;203(1):54-66.

39. Tan E, Song J, Deane AM, et al. Global Impact of Coronavirus Disease 2019 Infection Requiring Admission to the ICU: A Systematic Review and Meta-analysis. *Chest*. 2021;159(2):524-36.
40. Docherty AB, Harrison EM, Green CA, et al. Features of 20 133 UK patients in hospital with covid-19 using the ISARIC WHO Clinical Characterisation Protocol: prospective observational cohort study. *BMJ*. 2020;369:m1985.
41. Horwitz LI, Jones SA, Cerfolio RJ, et al. Trends in COVID-19 Risk-Adjusted Mortality Rates. *J Hosp Med*. 2021;16(2):90-2.
42. Gray WK, Navaratnam AV, Day J, et al. Variability in COVID-19 in-hospital mortality rates between national health service trusts and regions in England: A national observational study for the Getting It Right First Time Programme. *EClinicalMedicine*. 2021;35:100859.
43. Centers for Disease Prevention and Control.. SARS-CoV-2 Infections and Hospitalizations Among Persons Aged ≥ 16 Years, by Vaccination Status — Los Angeles County, California, May 1–July 25, 2021 2021 [Available from: https://www.cdc.gov/mmwr/volumes/70/wr/mm7034e5.htm?s_cid=mm7034e5_w.
44. World Health Organization. Estimating mortality from COVID-19 2021 [Available from: <https://www.who.int/news-room/commentaries/detail/estimating-mortality-from-covid-19>.
45. Brazeau RV, S Jenks et al. . COVID-19 Infection Fatality Ratio: Estimates from Seroprevalence. Imperial College London (29-10-2020). Available from: [<https://www.imperial.ac.uk/mrc-global-infectious-disease-analysis/covid-19/report-34-ifr/>]
46. Wu Z, McGoogan JM. Characteristics of and Important Lessons From the Coronavirus Disease 2019 (COVID-19) Outbreak in China: Summary of a Report of 72314 Cases From the Chinese Center for Disease Control and Prevention. *JAMA*. 2020;323(13):1229-1242.
47. Wiersinga WJ, Rhodes A, Cheng AC, et al. Pathophysiology, Transmission, Diagnosis, and Treatment of Coronavirus Disease 2019 (COVID-19): A Review. *JAMA*. 2020;324(8):782-93.
48. Awortwe C, Cascorbi I. Meta-analysis on outcome-worsening comorbidities of COVID-19 and related potential drug-drug interactions. *Pharmacol Res*. 2020;161:105250.
49. Centers for Disease Control and Prevention. Risk for COVID-19 Infection, Hospitalization, and Death By Age Group. 2021. Available from: <https://www.cdc.gov/coronavirus/2019-ncov/covid-data/investigations-discovery/hospitalization-death-by-age.html>
50. Guan WJ, Liang WH, Zhao Y, et al. Comorbidity and its impact on 1590 patients with COVID-19 in China: a nationwide analysis. *Eur Respir J*. 2020;55(5): 2000547
51. Zheng Z, Peng F, Xu B, et al. Risk factors of critical & mortal COVID-19 cases: A systematic literature review and meta-analysis. *J Infect*. 2020;81(2):e16-e25.
52. Sattar N, Ho FK, Gill JM, et al. BMI and future risk for COVID-19 infection and death across sex, age and ethnicity: Preliminary findings from UK biobank. *Diabetes Metab Syndr*. 2020;14(5):1149-51.
53. Rep. MMMW. Severe Outcomes Among Patients with Coronavirus Disease 2019 (COVID-19) - United States, February 12-March 16, 2020. *MMWR Morb Mortal Wkly Rep*. 2020;2020 Mar 27;69(12):343-346.
54. Al-Samkari H, Karp Leaf RS, Dzik WH, et al. COVID-19 and coagulation: bleeding and thrombotic manifestations of SARS-CoV-2 infection. *Blood*. 2020;136(4):489-500.
55. Magro C, Mulvey JJ, Berlin D, et al. Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: A report of five cases. *Transl Res*. 2020;220:1-13.
56. Ackermann M, Verleden SE, Kuehnel M, et al. Pulmonary Vascular Endothelialitis, Thrombosis, and Angiogenesis in Covid-19. *N Engl J Med*. 2020;383(2):120-8.
57. Guo T, Fan Y, Chen M, et al. Cardiovascular Implications of Fatal Outcomes of Patients With Coronavirus Disease 2019 (COVID-19). *JAMA Cardiol*. 2020;5(7):811-8.

58. Jothimani D, Venugopal R, Abedin MF, et al. COVID-19 and the liver. *J Hepatol.* 2020;73(5):1231-40.
59. Iadecola C, Anrather J, Kamel H. Effects of COVID-19 on the Nervous System. *Cell.* 2020;183(1):16-27 e1.
60. Yang X, Jin Y, Li R, et al. Prevalence and impact of acute renal impairment on COVID-19: a systematic review and meta-analysis. *Crit Care.* 2020;24(1):356.
61. Di Minno A, Ambrosino P, Calcaterra I, et al. COVID-19 and Venous Thromboembolism: A Meta-analysis of Literature Studies. *Semin Thromb Hemost.* 2020;46(7):763-71.
62. Dietz M, Chironi G, Claessens YE, et al. COVID-19 pneumonia: relationship between inflammation assessed by whole-body FDG PET/CT and short-term clinical outcome. *Eur J Nucl Med Mol Imaging.* 2021;48(1):260-8.
63. Sollini M, Ciccarelli M, Cecconi M, et al. Vasculitis changes in COVID-19 survivors with persistent symptoms: an [(18)F]FDG-PET/CT study. *Eur J Nucl Med Mol Imaging.* 2021;48(5):1460-6.
64. World Health Organization. COVID-19 Therapeutic Trial Synopses 2020 [Available from: https://www.who.int/blueprint/priority-diseases/key-action/COVID-19_Treatment_Trial_Design_Master_Protocol_synopsis_Final_18022020.pdf].
65. WHO Working Group on the Clinical Characterization and Management of COVID-19 infection. A minimal common outcome measure set for COVID-19 clinical research. *Lancet Infect Dis.* 2020;20(8):e192-e7.
66. Chu DK, Akl EA, Duda S, et al. Physical distancing, face masks, and eye protection to prevent person-to-person transmission of SARS-CoV-2 and COVID-19: a systematic review and meta-analysis. *Lancet.* 2020;395(10242):1973-87.
67. World Health Organization. COVID-19 vaccine tracker and landscape 2021 [updated 07 Sept 2021. Available [<https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines>]
68. Alberer M, Gnad-Vogt U, Hong HS, et al. Safety and immunogenicity of a mRNA rabies vaccine in healthy adults: an open-label, non-randomised, prospective, first-in-human phase 1 clinical trial. *Lancet.* 2017;390(10101):1511-20.
69. Pardi N, Hogan MJ, Pelc RS, et al. Zika virus protection by a single low-dose nucleoside-modified mRNA vaccination. *Nature.* 2017;543(7644):248-51.
70. Kose N, Fox JM, Sapparat C, et al. A lipid-encapsulated mRNA encoding a potentially neutralizing human monoclonal antibody protects against chikungunya infection. *Sci Immunol.* 2019;4(35).
71. Peng Y, Mentzer AJ, Liu G, et al. Broad and strong memory CD4(+) and CD8(+) T cells induced by SARS-CoV-2 in UK convalescent individuals following COVID-19. *Nat Immunol.* 2020;21(11):1336-45.
72. Wang Z, Schmidt F, Weisblum Y, et al. mRNA vaccine-elicited antibodies to SARS-CoV-2 and circulating variants. *Nature.* 2021;592(7855):616-22.
73. Earle KA, Ambrosino DM, Fiore-Gartland A, et al. Evidence for antibody as a protective correlate for COVID-19 vaccines. *Vaccine.* 2021;39(32):4423-8.
74. Puranik A LP, Silvert E. et al. Comparison of two highly-effective mRNA vaccines for COVID-19 during periods of Alpha and Delta variant prevalence 2021. updated 08 Aug 2021. Available from: [<https://www.medrxiv.org/content/10.1101/2021.08.06.21261707v3>].
75. Bian L, Gao F, Zhang J, et al. Effects of SARS-CoV-2 variants on vaccine efficacy and response strategies. *Expert Rev Vaccines.* 2021;20(4):365-73.
76. Bian L, Gao Q, Gao F, et al. Impact of the Delta variant on vaccine efficacy and response strategies. *Expert Rev Vaccines.* 2021:1-9.

77. O'Brien MP, Forleo-Neto E, Musser BJ, et al. Subcutaneous REGEN-COV Antibody Combination to Prevent Covid-19. *N Engl J Med*. 2021 Aug 4;NEJMoa2109682. doi: 10.1056/NEJMoa2109682.
78. Administration FaD. FDA authorizes REGEN-COV monoclonal antibody therapy for post-exposure prophylaxis (prevention) for COVID-19 2021. Available from: [https://www.fda.gov/drugs/drug-safety-and-availability/fda-authorizes-regen-cov-monoclonal-antibody-therapy-post-exposure-prophylaxis-prevention-covid-19.]
79. Chen F, Chan KH, Jiang Y, et al. In vitro susceptibility of 10 clinical isolates of SARS coronavirus to selected antiviral compounds. *J Clin Virol*. 2004;31(1):69-75.
80. Chan JF, Chan KH, Kao RY, et al. Broad-spectrum antivirals for the emerging Middle East respiratory syndrome coronavirus. *J Infect*. 2013;67(6):606-16.
81. Zhongji Meng TW, Chen Li, Xinhe Chen, Longti Li, Xueqin Qin, Hai Li, Jie Luo. "The Effect of Recombinant Human Interferon Alpha Nasal Drops to Prevent COVID-19 Pneumonia for Medical Staff in an Epidemic Area", *Current Topics in Medicinal Chemistry* 2021; 21(10) . <https://doi.org/10.2174/1568026621666210429083050>
82. Hoagland DA, Moller R, Uhl SA, et al. Leveraging the antiviral type I interferon system as a first line of defense against SARS-CoV-2 pathogenicity. *Immunity*. 2021;54(3):557-70 e5.
83. Struyf T, Deeks JJ, Dinnes J, et al. Signs and symptoms to determine if a patient presenting in primary care or hospital outpatient settings has COVID-19 disease. *Cochrane Database Syst Rev*. 2020;7:CD013665.
84. Goyal A, Cardozo-Ojeda EF, Schiffer JT. Potency and timing of antiviral therapy as determinants of duration of SARS-CoV-2 shedding and intensity of inflammatory response. *Sci Adv*. 2020;6(47).
85. Liu Y, Yan LM, Wan L, et al. Viral dynamics in mild and severe cases of COVID-19. *Lancet Infect Dis*. 2020;20(6):656-7.
86. Borczuk AC, Salvatore SP, Seshan SV, et al. COVID-19 pulmonary pathology: a multi-institutional autopsy cohort from Italy and New York City. *Mod Pathol*. 2020;33(11):2156-68.
87. Sheahan TP, Sims AC, Zhou S, et al. An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 in human airway epithelial cell cultures and multiple coronaviruses in mice. *Sci Transl Med*. 2020;12(541).
88. Gordon CJ, Tchesnokov EP, Woolner E, et al. Remdesivir is a direct-acting antiviral that inhibits RNA-dependent RNA polymerase from severe acute respiratory syndrome coronavirus 2 with high potency. *J Biol Chem*. 2020;295(20):5785-97.
89. Siegel D, Hui HC, Doerffle E, et al. Discovery and Synthesis of a Phosphoramidate Prodrug of a Pyrrolo[2,1-f][triazin-4-amine], Adenine C-Nucleoside (GS-5734) for the Treatment of Ebola and Emerging Viruses. *J Med Chem*. 2017;60(5):1648-61.
90. Mulangu S, Dodd LE, Davey RT, Jr., et al. A Randomized, Controlled Trial of Ebola Virus Disease Therapeutics. *N Engl J Med*. 2019;381(24):2293-303.
91. Humeniuk R, Mathias A, Cao H, et al. Safety, Tolerability, and Pharmacokinetics of Remdesivir, An Antiviral for Treatment of COVID-19, in Healthy Subjects. *Clin Transl Sci*. 2020;13(5):896-906.
92. Schooley RT CA, Beadle JR et al. Rethinking Remdesivir: Synthesis of Lipid Prodrugs that Substantially Enhance Anti-Coronavirus Activity. *Antimicrob Agents Chemother*. 2021 Jul 26:AAC0115521. doi: 10.1128/AAC.01155-21.
93. Sahakijpijarn S, Moon C, Koleng JJ, et al. Development of Remdesivir as a Dry Powder for Inhalation by Thin Film Freezing. *Pharmaceutics*. 2020;12(11).
94. Wang Y, Zhang D, Du G, et al. Remdesivir in adults with severe COVID-19: a randomised, double-blind, placebo-controlled, multicentre trial. *Lancet*. 2020;395(10236):1569-78.
95. Beigel JH, Tomashek KM, Dodd LE, et al. Remdesivir for the Treatment of Covid-19 - Final Report. *N Engl J Med*. 2020;383(19):1813-26.

96. Spinner CD, Gottlieb RL, Criner GJ, et al. Effect of Remdesivir vs Standard Care on Clinical Status at 11 Days in Patients With Moderate COVID-19: A Randomized Clinical Trial. *JAMA*. 2020;324(11):1048-57.
97. Goldman JD, Lye DCB, Hui DS, et al. Remdesivir for 5 or 10 Days in Patients with Severe Covid-19. *N Engl J Med*. 2020;383(19):1827-37.
98. WHO Solidarity Trial Consortium, Pan H, Peto R, et al. Repurposed Antiviral Drugs for Covid-19 - Interim WHO Solidarity Trial Results. *N Engl J Med*. 2021;384(6):497-511.
99. Organization WH. Therapeutics and COVID-19: living guideline. 2021.
100. NIH B, Maryland. Therapeutic Management of patients with COVID-19 2020 [Available from: <https://www.covid19treatmentguidelines.nih.gov/therapeutic-management/>].
101. Wahl A, Gralinski LE, Johnson CE, et al. SARS-CoV-2 infection is effectively treated and prevented by EIDD-2801. *Nature*. 2021;591(7850):451-7.
102. Painter WP ST, Baric R et al. Reduction in infectious SARS-CoV-2 in treatment study of COVID-19 with molnupiravir 2021 [Available from: <https://www.croiconference.org/abstract/reduction-in-infectious-sars-cov-2-in-treatment-study-of-covid-19-with-molnupiravir/>].
103. Fisher W EJ, Homan W et al. Molnupiravir, an Oral Antiviral Treatment for COVID-19. *medRxiv*. 2021. [preprint] Jun 17:2021.06.17.21258639. doi: 10.1101/2021.06.17.21258639.
104. Mediarelease M. Merck and Ridgeback Biotherapeutics Provide Update on Progress of Clinical Development Program for Molnupiravir, an Investigational Oral Therapeutic for the Treatment of Mild-to-Moderate COVID-19 2021 [Available from: <https://www.merck.com/news/merck-and-ridgeback-biotherapeutics-provide-update-on-progress-of-clinical-development-program-for-molnupiravir-an-investigational-oral-therapeutic-for-the-treatment-of-mild-to-moderate-covid-19/>].
105. Ivashchenko AA DK, Vostokova NV, et al. AVIFAVIR for Treatment of Patients with Moderate COVID-19: Interim Results of a Phase II/III Multicenter Randomized Clinical Trial. *Clin Infect Dis*. 2021 Aug 2;73(3):531-534. doi: 10.1093/cid/ciaa1146.
106. Hanna CR, Blyth KG, Burley G, et al. Glasgow Early Treatment Arm Favirpiravir (GETAFIX) for adults with early stage COVID-19: A structured summary of a study protocol for a randomised controlled trial. *Trials*. 2020;21(1):935.
107. University of Florida. Harnessing genome editing for COVID-19 drug discovery. 2021. Available from [<https://www.epi.ufl.edu/articles/harnessing-crispr-for-covid-19-drug-discovery.html>].
108. Sun P, Lu X, Xu C, et al. CD-SPACE2 inclusion compounds: An effective treatment for coronavirus disease 2019 (COVID-19). *J Med Virol*. 2020 Oct;92(10):1721-1723.
109. Khan A, Benthin C, Zeno B, et al. A pilot clinical trial of recombinant human angiotensin-converting enzyme 2 in acute respiratory distress syndrome. *Crit Care*. 2017;21(1):234.
110. Walls AC, Park YJ, Torbieri MA, et al. Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. *Cell*. 2020;181(2):281-92 e6.
111. Hoffmann M, Schröder S, Kleine-Weber H, et al. Nafamostat Mesylate Blocks Activation of SARS-CoV-2: New Treatment Option for COVID-19. *Antimicrob Agents Chemother*. 2020;64(6).
112. Yamamoto M, Kiso M, Sakai-Tagawa Y, et al. The Anticoagulant Nafamostat Potently Inhibits SARS-CoV-2 S Protein-Mediated Fusion in a Cell Fusion Assay System and Viral Infection In Vitro in a Cell-Type-Dependent Manner. *Viruses*. 2020;12(6).
113. Ragia G, Manolopoulos VG. Inhibition of SARS-CoV-2 entry through the ACE2/TMPRSS2 pathway: a promising approach for uncovering early COVID-19 drug therapies. *Eur J Clin Pharmacol*. 2020;76(12):1623-30.
114. Balfour H. Camostat cuts recovery time in half for mild COVID-19 patients 2021 [updated 30. Jul 2021. Available from: <https://www.europeanpharmaceuticalreview.com/news/159375/camostat-cuts-recovery-time-in-half-for-mild-covid-19-patients/>].

115. Gunst JD, Staerke NB, Pahus MH, et al. Efficacy of the TMPRSS2 inhibitor camostat mesilate in patients hospitalized with Covid-19-a double-blind randomized controlled trial. *EClinicalMedicine*. 2021;35:100849.
116. Iwasaka S, Shono Y, Tokuda K, et al. Clinical improvement in a patient with severe coronavirus disease 2019 after administration of hydroxychloroquine and continuous hemodiafiltration with nafamostat mesylate. *J Infect Chemother*. 2020;26(12):1319-23.
117. Doi S, Akashi YJ, Takita M, et al. Preventing thrombosis in a COVID-19 patient by combinatorial therapy with nafamostat and heparin during extracorporeal membrane oxygenation. *Acute Med Surg*. 2020;7(1):e585. doi: 10.1002/ams2.585. Epub ahead of print
118. Asakura H, Ogawa H. Potential of heparin and nafamostat combination therapy for COVID-19. *J Thromb Haemost*. 2020;18(6):1521-2.
119. Shamsi A, Mohammad T, Anwar S, et al. Glecaprevir and Maraviroc are high-affinity inhibitors of SARS-CoV-2 main protease: possible implication in COVID-19 therapy. *Biosci Rep*. 2020;40(6).
120. Ferrero MR, Garcia CC, Dutra de Almeida M, et al. CCR5 Antagonist Maraviroc Inhibits Acute Exacerbation of Lung Inflammation Triggered by Influenza Virus in Cigarette Smoke-Exposed Mice. *Pharmaceuticals (Basel)*. 2021;14(7).
121. Regeneron. REGENERON'S REGN-COV2 ANTIBODY COCKTAIL REDUCED VIRAL LEVELS AND IMPROVED SYMPTOMS IN NON-HOSPITALIZED COVID-19 PATIENTS 2020 [Available from: <https://investor.regeneron.com/news-releases/news-release-details/regenerons-regn-cov2-antibody-cocktail-reduced-viral-levels-and>.]
122. Weinreich DM, Sivapalasingam S, Norton T, et al. REGN-COV2, a Neutralizing Antibody Cocktail, in Outpatients with Covid-19. *N Engl J Med*. 2021 Jan 21;384(3):238-251.
123. GlaxoSmithKline. Vir Biotechnology and GSK announce VIR-7831 reduces hospitalisation and risk of death in early treatment of adults with COVID-19 2021 [Available from: <https://www.gsk.com/en-gb/media/press-releases/vir-biotechnology-and-gsk-announce-vir-7831-reduces-hospitalisation-and-risk-of-death-in-early-treatment-of-adults-with-covid-19/>].
124. Xu J, Xu K, Jung S, et al. Nanobodies from camelid mice and llamas neutralize SARS-CoV-2 variants. *Nature*. 2021;595(7866):278-82.
125. Valenzuela Nieto G, Jara R, Watanabe D, et al. Potent neutralization of clinical isolates of SARS-CoV-2 D614 and G614 variants by a non-oligomeric, sub-nanomolar affinity nanobody. *Sci Rep*. 2021;11(1):3318.
126. Boulware DR, Pullen MF, Bangdiwala AS, et al. A Randomized Trial of Hydroxychloroquine as Postexposure Prophylaxis for Covid-19. *N Engl J Med*. 2020;383(6):517-25.
127. Avidan MS, Dehbi H, Delany-Moretlwe S. Hydroxychloroquine as Postexposure Prophylaxis for Covid-19. *N Engl J Med*. 2020;383(11):1087-8.
128. Skipper CP, Pastick KA, Engen NW, et al. Hydroxychloroquine in Nonhospitalized Adults With Early COVID-19: A Randomized Trial. *Ann Intern Med*. 2020;173(8):623-31.
129. Elsayah HK, Elsokary MA, Elrazzaz MG, et al. Hydroxychloroquine for treatment of nonsevere COVID-19 patients: Systematic review and meta-analysis of controlled clinical trials. *J Med Virol*. 2021;93(3):1265-75.
130. RECOVERY Collaborative Group. Effect of Hydroxychloroquine in Hospitalized Patients with Covid-19. *N Engl J Med*. 2020 Nov 19;383(21):2030-2040.
131. Cavalcanti AB, Zampieri FG, Rosa RG, et al. Hydroxychloroquine with or without Azithromycin in Mild-to-Moderate Covid-19. *N Engl J Med*. 2020;383(21):2041-52.
132. Cortegiani A, Ippolito M, Ingoglia G, et al. Update I. A systematic review on the efficacy and safety of chloroquine/hydroxychloroquine for COVID-19. *J Crit Care*. 2020;59:176-90.
133. Fiolet T, Guihur A, Rebeaud ME, et al. Effect of hydroxychloroquine with or without azithromycin on the mortality of coronavirus disease 2019 (COVID-19) patients: a systematic review and meta-analysis. *Clin Microbiol Infect*. 2021;27(1):19-27.

134. Axfors C, Schmitt AM, Janiaud P, et al. Mortality outcomes with hydroxychloroquine and chloroquine in COVID-19 from an international collaborative meta-analysis of randomized trials. *Nat Commun.* 2021;12(1):2349.
135. Kinobe RT, Owens L. A systematic review of experimental evidence for antiviral effects of ivermectin and an in-silico analysis of ivermectin's possible mode of action against SARS-CoV-2. *Fundam Clin Pharmacol.* 2021.
136. Roman YM, Burela PA, Pasupuleti V, Piscocoy A, Vidal JE, Hernandez AV. Ivermectin for the treatment of COVID-19: A systematic review and meta-analysis of randomized controlled trials. *Clin Infect Dis.* 2021 Jun 28:ciab591. doi: 10.1093/cid/ciab591. Epub ahead of print
137. Bryant A, Lawrie TA, Dowswell T, et al. Ivermectin for Prevention and Treatment of COVID-19 Infection: A Systematic Review, Meta-analysis, and Trial Sequential Analysis to Inform Clinical Guidelines. *Am J Ther.* 2021;28(4):e434-e60.
138. Jacobs RF, Tabor DR, Burks AW, et al. Elevated interleukin-1 release by human alveolar macrophages during the adult respiratory distress syndrome. *Am Rev Respir Dis.* 1989;140(6):1686-92.
139. Hussell T, Bell TJ. Alveolar macrophages: plasticity in a tissue-specific context. *Nat Rev Immunol.* 2014;14(2):81-93.
140. Snelgrove RJ, Goulding J, Didierlaurent AM, et al. A critical function for CD200 in lung immune homeostasis and the severity of influenza infection. *Nat Immunol.* 2006;9(9):1074-83.
141. Forrester SJ, Booz GW, Sigmund CD, et al. Angiotensin II Signal Transduction: An Update on Mechanisms of Physiology and Pathophysiology. *Physiol Rev.* 2018;98(3):1627-738.
142. Yugandhar VG, Clark MA. Angiotensin III: a physiological relevant peptide of the renin angiotensin system. *Peptides.* 2013;46:26-32.
143. Xie X, Chen J, Wang X, et al. Age- and gender-related difference of ACE2 expression in rat lung. *Life Sci.* 2006;78(19):2166-71.
144. Verdecchia P, Cavallini C, Spanevello A, et al. The pivotal link between ACE2 deficiency and SARS-CoV-2 infection. *Eur J Intern Med.* 2020;156:14-20.
145. Chen J, Jiang Q, Xia X, et al. Individual variation of the SARS-CoV-2 receptor ACE2 gene expression and regulation. *Aging Cell.* 2020;19(7).
146. Marshall RP, Gohlke P, Chambers RC, et al. Angiotensin II and the fibroproliferative response to acute lung injury. *Am J Physiol Lung Cell Mol Physiol.* 2004;286(1):L156-64.
147. Cheng ZJ, Vapaatalo H, Mervaala E. Angiotensin II and vascular inflammation. *Med Sci Monit.* 2005;11(6):RA194-205.
148. Zhang H, Penninger JM, Li Y, et al. Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanism and potential therapeutic target. *Intensive Care Med.* 2020;46(4):586-90.
149. Chong TJ, Victorino GP. Angiotensin II subtype AT1 and AT2 receptors regulate microvascular hydraulic permeability via cAMP and cGMP. *J Surg Res.* 2006;131(1):105-10.
150. Bernstein KE, Kher Z, Giani JF, et al. Angiotensin-converting enzyme in innate and adaptive immunity. *Nat Rev Nephrol.* 2018;14(5):325-36.
151. Yamamoto S, Yancey PG, Zuo Y, et al. Macrophage polarization by angiotensin II-type 1 receptor aggravates renal injury-acceleration of atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2011;31(12):2856-64.
152. Ruiz-Ortega M, Lorenzo O, Egido J. Angiotensin III increases MCP-1 and activates NF-kappaB and AP-1 in cultured mesangial and mononuclear cells. *Kidney Int.* 2000;57(6):2285-98.
153. Xu J, Sriramula S, Xia H, et al. Clinical Relevance and Role of Neuronal AT1 Receptors in ADAM17-Mediated ACE2 Shedding in Neurogenic Hypertension. *Circ Res.* 2017;121(1):43-55.
154. Carvajal G, Rodriguez-Vita J, Rodriguez-Diez R, et al. Angiotensin II activates the Smad pathway during epithelial mesenchymal transdifferentiation. *Kidney Int.* 2008;74(5):585-95.
155. Williamson EJ, Walker AJ, Bhaskaran K, et al. Factors associated with COVID-19-related death using OpenSAFELY. *Nature.* 2020;584(7821):430-6.

156. Clift AK, Coupland CAC, Keogh RH, et al. Living risk prediction algorithm (QCOVID) for risk of hospital admission and mortality from coronavirus 19 in adults: national derivation and validation cohort study. *BMJ*. 2020;371:m3731.
157. Dashti H, Roche E, Bates D, et al. SARS2 simplified scores to estimate risk of hospitalization and death among patients with COVID-19. *Scientific Reports*. 2021 Mar;11(1):4945.
158. Sun H, Jain A, Leone MJ, et al. CoVA: An Acuity Score for Outpatient Screening that Predicts Coronavirus Disease 2019 Prognosis. *J Infect Dis*. 2021;223(1):38-46.
159. Zhao Y, Qin L, Zhang P, et al. Longitudinal COVID-19 profiling associates IL-1RA and IL-10 with disease severity and RANTES with mild disease. *JCI Insight*. 2020;5(13):e139834.
160. Vartak R, Patil SM, Saraswat A, et al. Aerosolized nanoliposomal carrier of remdesivir: an effective alternative for COVID-19 treatment in vitro. *Nanomedicine (Lond)*. 2021;16(14):1187-202.
161. Lurie Y, Nevens F, Aprosina ZG, et al. A multicentre, randomized study to evaluate the safety and efficacy of histamine dihydrochloride and interferon-alpha-2b for the treatment of chronic hepatitis C. *J Viral Hepat*. 2002;9(5):346-53.
162. Park A, Iwasaki A. Type I and Type III Interferons - Induction, Signaling, Evasion, and Application to Combat COVID-19. *Cell Host Microbe*. 2020;27(6):870-8.
163. Feld JJ, Kandel C, Biondi MJ, et al. Peginterferon lambda for the treatment of outpatients with COVID-19: a phase 2, placebo-controlled randomised trial. *Lancet Respir Med*. 2021;9(5):498-510.
164. Jagannathan P, Andrews JR, Bonilla H, et al. Peginterferon Lambda-1a for treatment of outpatients with uncomplicated COVID-19: a randomized placebo-controlled trial. *Nat Commun*. 2021;12(1):1967.
165. Finney LJ, Glanville N, Farne H, et al. Inhaled corticosteroids downregulate the SARS-CoV-2 receptor ACE2 in COPD through suppression of type I interferon. *J Allergy Clin Immunol*. 2021;147(2):510-9 e5.
166. Ramakrishnan S, Nicolau DV, Jr., Lafford J B, et al. Inhaled budesonide in the treatment of early COVID-19 (STOIC): a phase 2, open-label, randomised controlled trial. *Lancet Respir Med*. 2021;9(7):763-72.
167. Gharbharan A, Jordans CCE, , Graatsen Kessel C et al. Effects of potent neutralizing antibodies from convalescent plasma in patients hospitalized for severe SARS-CoV-2 infection. *Nat Commun* **12**, 3189 (2021). <https://doi.org/10.1038/s41467-021-23469-2>
168. Rauch A, Dupont A, Goutay J, et al. Endotheliopathy is induced by plasma from critically-ill patients and associated with organ failure in severe COVID-19. *Circulation*. 2020. 142(19):1881-4
169. Bradfute SB, Hurwitz J, Yingling AV, et al. Severe Acute Respiratory Syndrome Coronavirus 2 Neutralizing Antibody Titer in Convalescent Plasma and Recipients in New Mexico: An Open Treatment Study in Patients With Coronavirus Disease 2019. *J Infect Dis*. 2020;222(10):1620-8.
170. Lucas C, Klein J, Sundaram M, et al. Kinetics of antibody responses dictate COVID-19 outcome. *medRxiv*. [Preprint]. 2020 Dec 22:2020.12.18.20248331. doi: 10.1101/2020.12.18.20248331.
171. Piechotta V, Iannizzi C, Chai KL, et al. Convalescent plasma or hyperimmune immunoglobulin for people with COVID-19: a living systematic review. *Cochrane Database Syst Rev*. 2021;5:CD013600.
172. Begin P, Callum J, Jamula E, et al. Convalescent plasma for hospitalized patients with COVID-19: an open-label, randomized controlled trial. *Nat Med*. 2021 Sep 9. doi: 10.1038/s41591-021-01488-2. Epub ahead of print. .
173. Lo KB, Bhargav R, Salacup G, et al. Angiotensin converting enzyme inhibitors and angiotensin II receptor blockers and outcomes in patients with COVID-19: a systematic review and meta-analysis. *Expert Rev Cardiovasc Ther*. 2020;18(12):919-30.
174. Haydar D, Cory TJ, Birket SE, et al. Azithromycin Polarizes Macrophages to an M2 Phenotype via Inhibition of the STAT1 and NF-kappaB Signaling Pathways. *J Immunol*. 2019;203(4):1021-30.
175. Cory TJ, Birket SE, Murphy BS, et al. Impact of azithromycin treatment on macrophage gene expression in subjects with cystic fibrosis. *J Cyst Fibros*. 2014;13(2):164-71.

176. Furtado RHM, Berwanger O, Fonseca HA, et al. Azithromycin in addition to standard of care versus standard of care alone in the treatment of patients admitted to the hospital with severe COVID-19 in Brazil (COALITION II): a randomised clinical trial. *Lancet*. 2020;396(10256):959-67.
177. Liang W, Liang H, Ou L, et al. Development and Validation of a Clinical Risk Score to Predict the Occurrence of Critical Illness in Hospitalized Patients With COVID-19. *JAMA Intern Med*. 2020;180(8):1081-9.
178. Caricchio R, Gallucci M, Dass C, et al. Preliminary predictive criteria for COVID-19 cytokine storm. *Ann Rheum Dis*. 2021; 80(1):88-95
179. Hao B, Sotudian S, Wang T, et al. Early prediction of level-of-care requirements in patients with COVID-19. *Elife*. 2020;9.
180. Bahl A, Van Baalen MN, Ortiz L, et al. Early predictors of in-hospital mortality in patients with COVID-19 in a large American cohort. *Intern Emerg Med*. 2020;15(8):1485-1499.
181. Simadibrata DM, Calvin J, Wijaya AD, et al. Neutrophil-to-lymphocyte ratio on admission to predict the severity and mortality of COVID-19 patients: A meta-analysis. *Am J Emerg Med*. 2021;42:60-9.
182. Rauch A, Labreuche J, Lassalle F, et al. Coagulation biomarkers are independent predictors of increased oxygen requirements in COVID-19. *J Thromb Haemost*. 2020;18(11):2942-53.
183. Henry B, Cheruiyot I, Vikse J, et al. Lymphopenia and neutrophilia at admission predicts severity and mortality in patients with COVID-19: a meta-analysis. *Acta Biom*. 2020;91(3):e2020008.
184. Varga Z, Flammer AJ, Steiger P, et al. Endothelial cell infection and endotheliitis in COVID-19. *Lancet*. 2020;395(10234):1417-8.
185. Zhang X, Jiang M, Yang J. Potential value of circulating endothelial cells for the diagnosis and treatment of COVID-19. *Int J Infect Dis*. 2021; 107:232-235.
186. Stauning MA, Altintas I, Kallemose T, et al. Soluble Urokinase Plasminogen Activator Receptor as a Decision Marker for Early Discharge of Patients with COVID-19 Symptoms in the Emergency Department. *J Emerg Med*. 2021; S0736-4679(21)00295-X.
187. Del Valle DM, Kim-Schulze S, Huang HH, et al. An inflammatory cytokine signature predicts COVID-19 severity and survival. *Nat Med*. 2020; 26(10):1636-1643
188. Li C, Jiang J, Wang F, et al. Longitudinal correlation of biomarkers of cardiac injury, inflammation, and coagulation to outcome in hospitalized COVID-19 patients. *J Mol Cell Cardiol*. 2020;147:74-87.
189. Rendeiro AF, Casano J, Morais CK, et al. Profiling of immune dysfunction in COVID-19 patients allows early prediction of disease progression. *Life Sci Alliance*. 2021;4(2).
190. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020; 395(10223):497-506.
191. Wichmann D, Sperhake JP, Lutgehetmann M, et al. Autopsy Findings and Venous Thromboembolism in Patients With COVID-19: A Prospective Cohort Study. *Ann Intern Med*. 2020;173(4):268-77.
192. Hagman K, Hedenstierna M, Gille-Johnson P, et al. SARS-CoV-2 RNA in serum as predictor of severe outcome in COVID-19: a retrospective cohort study. *Clin Infect Dis*. 2020. Aug 28;ciaa1285. doi: 10.1093/cid/ciaa1285.
193. Tang K, Wu L, Luo Y, et al. Quantitative assessment of SARS-CoV-2 RNAemia and outcome in patients with coronavirus disease 2019. *J Med Virol*. 2021;93(5):3165-75.
194. Balfanz P, Hartmann B, Muller-Wieland D, et al. Early risk markers for severe clinical course and fatal outcome in German patients with COVID-19. *PLoS One*. 2021;16(1):e0246182.
195. Yan L, Zhang, H., Goncalves, J. et al. An interpretable mortality prediction model for COVID-19 patients. *Nat Mach Intell*. 2020;2:282-8.
196. Li Q, Zhang J, Ling Y, et al. A simple algorithm helps early identification of SARS-CoV-2 infection patients with severe progression tendency. *Infection*. 2020;48(4):577-84.

197. Xu Z, Wu GM, Li Q, et al. Predictive Value of Combined LIPS and ANG-2 Level in Critically Ill Patients with ARDS Risk Factors. *Mediators Inflamm.* 2018;2018:1739615.
198. Ahmed ME, Hamed G, Fawzy S, et al. Lung injury prediction scores: Clinical validation and C-reactive protein involvement in high risk patients. *Med Intensiva.* 2020;44(5):267-74.
199. Festic E, Bansal V, Kor DJ, et al. SpO₂/FiO₂ ratio on hospital admission is an indicator of early acute respiratory distress syndrome development among patients at risk. *J Intensive Care Med.* 2015;30(4):209-16.
200. Gajic O, Dabbagh O, Park PK, et al. Early identification of patients at risk of acute lung injury: evaluation of lung injury prediction score in a multicenter cohort study. *Am J Respir Crit Care Med.* 2011;183(4):462-70.
201. Trillo-Alvarez C, Cartin-Ceba R, Kor DJ, et al. Acute lung injury prediction score: derivation and validation in a population-based sample. *Eur Respir J.* 2011;37(3):604-9.
202. Wang D, Hu B, Hu C, et al. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA.* 2020;323(11):1061-9.
203. Kopel J, Perisetti A, Gajendran M, et al. Clinical Insights into the Gastrointestinal Manifestations of COVID-19. *Dig Dis Sci.* 2020;65(7):1932-9.
204. Figliozzi S, Masci PG, Ahmadi N, et al. Predictors of adverse prognosis in COVID-19: A systematic review and meta-analysis. *Eur J Clin Invest.* 2020;50(10):e13362.
205. Singh K, Mittal S, Gollapudi S, et al. A meta-analysis of SARS-CoV-2 patients identifies the combinatorial significance of D-dimer, C-reactive protein, lymphocyte, and neutrophil values as a predictor of disease severity. *Int J Lab Hematol.* 2021;43(2):324-8.
206. Arnold DT, Attwood M, Barratt S, et al. Predicting outcomes of COVID-19 from admission biomarkers: a prospective UK cohort study. *Emerg Med J.* 2021; 38(7):543-8
207. Zhang Y, Xiao M, Zhang S, et al. Coagulation and Antiphospholipid Antibodies in Patients with Covid-19. *N Engl J Med.* 2020;382(17):e48.
208. Recovery Collaborative Group, Horby P, Lim WS, et al. Dexamethasone in Hospitalized Patients with Covid-19 - Preliminary Report. *N Engl J Med.* 2021 Feb 25;384(8):693-704.
209. Lee KH, Yoon S, Jeong GH, et al. Efficacy of Corticosteroids in Patients with SARS, MERS and COVID-19: A Systematic Review and Meta-analysis. *J Clin Med.* 2020;9(8).
210. Russell CD, Millar JE, Baillie JK. Clinical evidence does not support corticosteroid treatment for 2019-nCoV lung injury. *Lancet.* 2020;395(10223):473-5.
211. Pasin L, Navalesi P, Zangrillo A, et al. Corticosteroids for Patients With Coronavirus Disease 2019 (COVID-19) With Different Disease Severity: A Meta-Analysis of Randomized Clinical Trials. *J Cardiothorac Vasc Anesth.* 2021;35(2):578-84.
212. Leonard WJ, O'Hea JJ. Jaks and STATs: biological implications. *Annu Rev Immunol.* 1998;16:293-322.
213. Channappanavar R, Fehr AR, Vijay R, et al. Dysregulated Type I Interferon and Inflammatory Monocyte-Macrophage Responses Cause Lethal Pneumonia in SARS-CoV-Infected Mice. *Cell Host Microbe.* 2016;19(2):181-93.
214. Channappanavar R, Fehr AR, Zheng J, et al. IFN-I response timing relative to virus replication determines MERS coronavirus infection outcomes. *J Clin Invest.* 2019;130:3625-39.
215. Yoshikawa T, Hill TE, Yoshikawa N, et al. Dynamic innate immune responses of human bronchial epithelial cells to severe acute respiratory syndrome-associated coronavirus infection. *PLoS One.* 2010;5(1):e8729.
216. Hadjadj J, Yatim N, Barnabei L, et al. Impaired type I interferon activity and inflammatory responses in severe COVID-19 patients. *Science.* 2020;369(6504):718-24.
217. Chu H, Chan JF, Wang Y, et al. Comparative Replication and Immune Activation Profiles of SARS-CoV-2 and SARS-CoV in Human Lungs: An Ex Vivo Study With Implications for the Pathogenesis of COVID-19. *Clin Infect Dis.* 2020;71(6):1400-9.

218. World Health Organization. Solidarity clinical trial for COVID-19 treatments. 2020.
219. Rahmani H, Davoudi-Monfared E, Nourian A, et al. Interferon beta-1b in treatment of severe COVID-19: A randomized clinical trial. *Int Immunopharmacol*. 2020;88:106903.
220. Wang N, Zhan Y, Zhu L, et al. Retrospective Multicenter Cohort Study Shows Early Interferon Therapy Is Associated with Favorable Clinical Responses in COVID-19 Patients. *Cell Host Microbe*. 2020;28(3):455-64 e2.
221. Jalkanen J, Hollmen M, Jalkanen S. Interferon beta-1a for COVID-19: critical importance of the administration route. *Crit Care*. 2020;24(1):335.
222. Monk PD, Marsden RJ, Tear VJ, et al. Safety and efficacy of inhaled nebulised interferon beta-1a (SNG001) for treatment of SARS-CoV-2 infection: a randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet Respir Med*. 2021 Feb;9(2):196-206.
223. Spyropoulos AC, Levy JH, Ageno W, et al. Scientific and Standardization Committee communication: Clinical guidance on the diagnosis, prevention, and treatment of venous thromboembolism in hospitalized patients with COVID-19. *J Thromb Haemost*. 2020;18(8):1859-65.
224. Panel C-TG. Coronavirus Disease 2019 (COVID-19) Treatment Guidelines. National Institutes of Health. 2021.
225. ATTACC Investigators; ACTIV-4a Investigators; REMAP-CAP Investigators et al., et al. Therapeutic Anticoagulation with Heparin in Noncritically Ill Patients with Covid-19. *N Engl J Med*. 2021;385(9):790-802.
226. Di Castelnuovo A, Costanzo S, Antinori A, et al. Heparin in COVID-19 Patients Is Associated with Reduced In-Hospital Mortality: The Multicenter Italian CORIST Study. *Thromb Haemost*. 2021;121(8):1054-65.
227. The Scientist. 2020 [Available from: <https://www.the-scientist.com/news-opinion/eli-lilly-halts-antibody-trial-in-hospitalized-covid-19-patients-52092>]
228. Tuccori M, Ferraro S, Convertino I, et al. Anti-SARS-CoV-2 neutralizing monoclonal antibodies: clinical pipeline. *MAbs*. 2020;12(1):1854149.
229. Food and Drug Administration. Fact Sheet for health care providers [Available from: <https://www.fda.gov/media/145802/download>].
230. Daher A, Balfanz P, Aetou M, et al. Clinical course of COVID-19 patients needing supplemental oxygen outside the intensive care unit. *Sci Rep*. 2021;11(1):2256.
231. Marchesi C, Paradis P, Schiffrin EL. Role of the renin-angiotensin system in vascular inflammation. *Trends Pharmacol Sci*. 2008;29(7):367-74.
232. Scalia R, Gong Y, Bernins N, et al. A novel role for calpain in the endothelial dysfunction induced by activation of angiotensin II type 1 receptor signaling. *Circ Res*. 2011;108(9):1102-11.
233. Jafarzadeh A, Chauban P, Saha B, et al. Contribution of monocytes and macrophages to the local tissue inflammation and cytokine storm in COVID-19: Lessons from SARS and MERS, and potential therapeutic interventions. *Life Sci*. 2020;257:118102.
234. Channappanavar R, Perlman S. Evaluation of Activation and Inflammatory Activity of Myeloid Cells During Pathogenic Human Coronavirus Infection. *Methods Mol Biol*. 2020;2099:195-204.
235. Nathan CF, Murray HW, Wiebe ME, et al. Identification of interferon-gamma as the lymphokine that activates human macrophage oxidative metabolism and antimicrobial activity. *J Exp Med*. 1983;158(3):670-89.
236. Stein M, Keshav S, Harris N, et al. Interleukin 4 potently enhances murine macrophage mannose receptor activity: a marker of alternative immunologic macrophage activation. *J Exp Med*. 1992;176(1):287-92.
237. Shapouri-Moghaddam A, Mohammadian S, Vazini H, et al. Macrophage plasticity, polarization, and function in health and disease. *J Cell Physiol*. 2018;233(9):6425-40.
238. Murray PJ. Macrophage Polarization. *Annu Rev Physiol*. 2017;79:541-66.

239. Boada-Romero E, Martinez J, Heckmann BL, et al. The clearance of dead cells by efferocytosis. *Nat Rev Mol Cell Biol.* 2020;21(7):398-414.
240. Short KR, Kroeze E, Fouchier RAM, et al. Pathogenesis of influenza-induced acute respiratory distress syndrome. *Lancet Infect Dis.* 2014;14(1):57-69.
241. Herold S, Gabrielli NM, Vadasz I. Novel concepts of acute lung injury and alveolar-capillary barrier dysfunction. *Am J Physiol Lung Cell Mol Physiol.* 2013;305(10):L665-81.
242. Hettinger J, Richards DM, Hansson J, et al. Origin of monocytes and macrophages in a committed progenitor. *Nat Immunol.* 2013;14(8):821-30.
243. Kang K, Bachu M, Park SH, et al. IFN-gamma selectively suppresses a subset of TLR4-activated genes and enhancers to potentiate macrophage activation. *Nat Commun.* 2019;10(1):3320.
244. Borczuk AC, Salvatore SP, Seshan SV, et al. COVID-19 pulmonary pathology: a multi-institutional autopsy cohort from Italy and New York City. *Mod Pathol.* 2020.
245. Wang C, Xie J, Zhao L, et al. Alveolar macrophage dysfunction and cytokine storm in the pathogenesis of two severe COVID-19 patients. *EBioMedicine.* 2020;57:102833.
246. Murray PJ, Wynn TA. Protective and pathogenic functions of macrophage subsets. *Nat Rev Immunol.* 2011;11(11):723-37.
247. Rosseau S, Hammerl P, Maus U, et al. Phenotypic characterization of alveolar monocyte recruitment in acute respiratory distress syndrome. *Am J Physiol Lung Cell Mol Physiol.* 2000;279(1):L25-35.
248. Brittan M, Barr L, Conway Morris A, et al. A novel subpopulation of monocyte-like cells in the human lung after lipopolysaccharide inhalation. *Eur Respir J.* 2012;40(1):206-14.
249. Gerrick KY, Gerrick ER, Gupta A, et al. Transcriptional profiling identifies novel regulators of macrophage polarization. *PLoS One.* 2018;13(12):e0200602.
250. Chen X, Tang J, Shuai W, et al. Macrophage polarization and its role in the pathogenesis of acute lung injury/acute respiratory distress syndrome. *Inflamm Res.* 2020;69(9):883-95.
251. Song C, Li H, Li Y, et al. NETs promote ALI/ARDS inflammation by regulating alveolar macrophage polarization. *Exp Cell Res.* 2019;382(2):111486.
252. Smadja DM, Guerin CL, Chocron P, et al. Angiopoietin-2 as a marker of endothelial activation is a good predictor factor for intensive care unit admission of COVID-19 patients. *Angiogenesis.* 2020;23(4):611-20.
253. Wang H, Zhang Y, Mo P, et al. Neutrophil to CD4+ lymphocyte ratio as a potential biomarker in predicting virus negative conversion time in COVID-19. *Int Immunopharmacol.* 2020;85:106683.
254. Liu Y, Du X, Chen J, et al. Neutrophil-to-lymphocyte ratio as an independent risk factor for mortality in hospitalized patients with COVID-19. *J Infect.* 2020;81(1):e6-e12.
255. Liu J, Liu Y, Xiong P, et al. Neutrophil-to-lymphocyte ratio predicts critical illness patients with 2019 coronavirus disease in the early stage. *J Transl Med.* 2020;18(1):206.
256. Schulte-Schrepping J, Reusch N, Paclik D, et al. Severe COVID-19 Is Marked by a Dysregulated Myeloid Cell Compartment. *Cell.* 2020;182(6):1419-40 e23.
257. Tu GW, Shi Y, Zheng YJ, et al. Glucocorticoid attenuates acute lung injury through induction of type 2 macrophage. *J Transl Med.* 2017;15(1):181.
258. Lansbury LE, Rodrigo C, Leonardi-Bee J, et al. Corticosteroids as Adjunctive Therapy in the Treatment of Influenza: An Updated Cochrane Systematic Review and Meta-analysis. *Crit Care Med.* 2020;48(2):e98-e106.
259. WHO Rapid Evidence Appraisal for COVID-19 Therapies (REACT) Working Group, Sterne JAC, Murthy S, et al. Association Between Administration of Systemic Corticosteroids and Mortality Among Critically Ill Patients With COVID-19: A Meta-analysis. *JAMA.* 2020;324(13):1330-41.
260. Giamarellos-Bourboulis EJ, Netea MG, Rovina N, et al. Complex Immune Dysregulation in COVID-19 Patients with Severe Respiratory Failure. *Cell Host Microbe.* 2020;27(6):992-1000 e3.

261. Galvan-Roman JM, Rodriguez-Garcia SC, Roy-Vallejo E, et al. IL-6 serum levels predict severity and response to tocilizumab in COVID-19: An observational study. *J Allergy Clin Immunol*. 2021;147(1):72-80 e8.
262. Perrone F, Piccirillo MC, Ascierto PA, et al. Tocilizumab for patients with COVID-19 pneumonia. The single-arm TOCIVID-19 prospective trial. *J Transl Med*. 2020;18(1):405.
263. Piccirillo MC, Ascierto P, Atripaldi L, et al. TOCIVID-19 - A multicenter study on the efficacy and tolerability of tocilizumab in the treatment of patients with COVID-19 pneumonia. Study protocol. *Contemp Clin Trials*. 2020;98:106165.
264. Ramiro S, Mostard RLM, Magro-Checa C, et al. Historically controlled comparison of glucocorticoids with or without tocilizumab versus supportive care only in patients with COVID-19-associated cytokine storm syndrome: results of the CHIC study. *Ann Rheum Dis*. 2020;79(9):1143-51.
265. Recovery Collaborative Group. Tocilizumab in patients admitted to hospital with COVID-19 (RECOVERY): a randomised, controlled, open-label, platform trial. *Lancet*. 2021;397(10285):1637-45.
266. WHO Rapid Evidence Appraisal for COVID-19 Therapies (REACT) Working Group, Shankar-Hari M, Vale CL, et al. Association Between Administration of IL-6 Antagonists and Mortality Among Patients Hospitalized for COVID-19: A Meta-analysis. *JAMA*. 2021;326(6):499-518.
267. Tleyjeh IM, Kashour Z, Damlaj M, et al. Efficacy and safety of tocilizumab in COVID-19 patients: a living systematic review and meta-analysis. *Clin Microbiol Infect*. 2021;27(2):215-27.
268. National Institutes for Health, Bethesda, Maryland. Immunomodulators Under Evaluation for the Treatment of COVID-19. 2021. Available from [https://files.covid19treatmentguidelines.nih.gov/guidelines/section/section_70.pdf].
269. Huet T, Beaussier H, Voisin O, et al. Anakinra for severe forms of COVID-19: a cohort study. *Lancet Rheumatol*. 2020;2(7):e393-e400.
270. Cavalli G, De Luca G, Campochiaro C, et al. Interleukin-1 blockade with high-dose anakinra in patients with COVID-19, acute respiratory distress syndrome, and hyperinflammation: a retrospective cohort study. *Lancet Rheumatol*. 2020;2(6):e325-31.
271. Erden A, Ozdemir B, Karakas O, et al. Evaluation of 17 patients with COVID-19 pneumonia treated with anakinra according to HScore, COFA, MuLBSTA, and Brescia-COVID respiratory severity scale (BCRSS) scoring systems. *J Med Virol*. 2021;93(3):1532-7.
272. Aomar-Millan IF, Salvatierra J, Torres-Parejo U, et al. Anakinra after treatment with corticosteroids alone or with tocilizumab in patients with severe COVID-19 pneumonia and moderate hyperinflammation. A retrospective cohort study. *Intern Emerg Med*. 2021.
273. Bozzi G, Mangioni D, Minola F, et al. Anakinra combined with methylprednisolone in patients with severe COVID-19 pneumonia and hyperinflammation: An observational cohort study. *J Allergy Clin Immunol*. 2021;147(2):551-5 e4.
274. Balkhair A, Al-Zakwani I, Al Busaidi M, et al. Anakinra in hospitalized patients with severe COVID-19 pneumonia requiring oxygen therapy: Results of a prospective, open-label, interventional study. *Int J Infect Dis*. 2021;103:288-96.
275. CORIMUNO-19 Collaborative group. Effect of anakinra versus usual care in adults in hospital with COVID-19 and mild-to-moderate pneumonia (CORIMUNO-ANA-1): a randomised controlled trial. *Lancet Respir Med*. 2021;9(3):295-304.
276. Pontali E, Volpi S, Signori A, et al. Efficacy of early anti-inflammatory treatment with high doses of intravenous anakinra with or without glucocorticoids in patients with severe COVID-19 pneumonia. *J Allergy Clin Immunol*. 2021;147(4):1217-25.
277. Khan FA, Stewart I, Fabbri L, et al. Systematic review and meta-analysis of anakinra, sarilumab, siltuximab and tocilizumab for COVID-19. *Thorax*. 2021;76(9):907-19.
278. Rovina N, Akinosoglou K, Eugen-Olsen J, et al. Soluble urokinase plasminogen activator receptor (suPAR) as an early predictor of severe respiratory failure in patients with COVID-19 pneumonia. *Crit Care*. 2020;24(1):187.

279. Kyriazopoulou E, Panagopoulos P, Metallidis S, et al. An open label trial of anakinra to prevent respiratory failure in COVID-19. *Elife*. 2021;10.
280. Caricchio R, Abbate A, Gordeev I, et al. Effect of Canakinumab vs Placebo on Survival Without Invasive Mechanical Ventilation in Patients Hospitalized With Severe COVID-19: A Randomized Clinical Trial. *JAMA*. 2021;326(3):230-9.
281. Cron RQ, Caricchio R, Chatham WW. Calming the cytokine storm in COVID-19. *Nat Med*. 2021.
282. Lee JS, Shin EC. The type I interferon response in COVID-19: implications for treatment. *Nat Rev Immunol*. 2020;20(10):585-6.
283. Stebbing J, Phelan A, Griffin I, et al. COVID-19: combining antiviral and anti-inflammatory treatments. *Lancet Infect Dis*. 2020;20(4):400-2.
284. Walz L, Cohen AJ, Rebaza AP, et al. JAK-inhibitor and type I interferon ability to produce favorable clinical outcomes in COVID-19 patients: a systematic review and meta-analysis. *BMC Infect Dis*. 2021;21(1):47.
285. Cantini F, Niccoli L, Matarrese D, et al. Baricitinib therapy in COVID-19: A pilot study on safety and clinical impact. *J Infect*. 2020;81(2):318-56.
286. Cantini F, Niccoli L, Nannini C, et al. Beneficial impact of Baricitinib in COVID-19 moderate pneumonia; multicentre study. *J Infect*. 2020;81(4):647-79.
287. Kalil AC, Patterson TF, Mehta AK, et al. Baricitinib plus Remdesivir for Hospitalized Adults with Covid-19. *N Engl J Med*. 2021;384(9):795-807.
288. National Institutes of Health, Bethesda, Maryland. Kinase Inhibitors: Baricitinib and Other Janus Kinase Inhibitors, and Bruton's Tyrosine Kinase Inhibitors. 2021.
289. Kulkarni S, Fisk M, Kostapanos M, et al. Repurposed immunomodulatory drugs for Covid-19 in pre-ICU patients - mulTi-Arm Therapeutic study in pre-ICU patients admitted with Covid-19 - Repurposed Drugs (TACTIC-R): A structured summary of a study protocol for a randomised controlled trial. *Trials*. 2020;21(1):626.
290. Press release. Novartis provides update on RUXCOVID study of ruxolitinib for hospitalized patients with COVID-19 2021 [Available from. <https://www.novartis.com/news/media-releases/novartis-provides-update-ruxcovid-study-ruxolitinib-hospitalized-patients-covid-19>.
291. Guimaraes PO, Quirk D, Furtado RH, et al. Tofacitinib in Patients Hospitalized with Covid-19 Pneumonia. *N Engl J Med*. 2021;385(3):406-15.
292. Stallmach A, Kortgen A, Gonnert F, et al. Infliximab against severe COVID-19-induced cytokine storm syndrome with organ failure- a cautionary case series. *Crit Care*. 2020;24(1):444.
293. Hue S, Beldi-Ferchichi A, Bendib I, et al. Uncontrolled Innate and Impaired Adaptive Immune Responses in Patients with COVID-19 ARDS. *Am J Respir Crit Care Med*. 2020; 2020(11). <https://doi.org/10.1164/rccm.202005-1885OC>
294. Hamilton JA. GM-CSF in inflammation. *J Exp Med*. 2020;217(1): e20190945
295. Matute-Bello G, Liles WC, Radella F, 2nd, et al. Modulation of neutrophil apoptosis by granulocyte colony-stimulating factor and granulocyte/macrophage colony-stimulating factor during the course of acute respiratory distress syndrome. *Crit Care Med*. 2000;28(1):1-7.
296. Overgaard CE, Schlingmann B, Dorsainvil White S, et al. The relative balance of GM-CSF and TGF-beta1 regulates lung epithelial barrier function. *Am J Physiol Lung Cell Mol Physiol*. 2015;308(12):L1212-23.
297. Rosler B, Herold S. Lung epithelial GM-CSF improves host defense function and epithelial repair in influenza virus pneumonia-a new therapeutic strategy? *Mol Cell Pediatr*. 2016;3(1):29.
298. Paine R, 3rd, Standiford TJ, Dechert RE, et al. A randomized trial of recombinant human granulocyte-macrophage colony stimulating factor for patients with acute lung injury. *Crit Care Med*. 2012;40(1):90-7.

299. Hall MW, Geyer SM, Guo CY, et al. Innate immune function and mortality in critically ill children with influenza: a multicenter study. *Crit Care Med.* 2013;41(1):224-36.
300. Bosteels C, Maes B, Van Damme K, et al. Sargramostim to treat patients with acute hypoxic respiratory failure due to COVID-19 (SARPAC): A structured summary of a study protocol for a randomised controlled trial. *Trials.* 2020;21(1):491.
301. Biospace. Second Randomized Trial of Leukine® (sargramostim) in COVID-19 Demonstrates Improvement in Lung Function 2021 [updated 28.Jun 2021. Available from: <https://www.biospace.com/article/releases/second-randomized-trial-of-leukine-sargramostim-in-covid-19-demonstrates-improvement-in-lung-function/>.
302. De Luca G, Cavalli G, Campochiaro C, et al. GM-CSF blockade with mavrilimumab in severe COVID-19 pneumonia and systemic hyperinflammation: a single-centre, prospective cohort study. *Lancet Rheumatol.* 2020;2(8):e465-e73.
303. Cremer PC, Abbate A, Hudock K, et al. Mavrilimumab in patients with severe COVID-19 pneumonia and systemic hyperinflammation (MASH-COVID): an investigator initiated, multicentre, double-blind, randomised, placebo-controlled trial. *Lancet Rheumatol.* 2021;3(6):e410-e8.
304. Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. *Science.* 2004;303(5663):1532-5.
305. de Bont CM, Boelens WC, Pruijn GJM. NETosis, complement, and coagulation: a triangular relationship. *Cell Mol Immunol.* 2019;16(1):19-27.
306. Schonrich G, Raftery MJ. Neutrophil Extracellular Traps Go Viral. *Front Immunol.* 2016;7:366.
307. Lv D, Xu Y, Cheng H, et al. A novel cell-based assay for dynamically detecting neutrophil extracellular traps-induced lung epithelial injuries. *Exp Cell Res.* 2020;394(2):112101.
308. Saffarzadeh M, Juenemann C, Queisser MA, et al. Neutrophil extracellular traps directly induce epithelial and endothelial cell death: a predominant role of histones. *PLoS One.* 2012;7(2):e32366.
309. Jimenez-Alcazar M, Rangaswamy C, Pandey R, et al. Host DNases prevent vascular occlusion by neutrophil extracellular traps. *Science.* 2017;358(6367):1202-6.
310. Gupta AK, Joshi MB, Philippova M, et al. Activated endothelial cells induce neutrophil extracellular traps and are susceptible to NETosis-mediated cell death. *FEBS Lett.* 2010;584(14):3193-7.
311. You Y, Liu Y, Li F, et al. Anti-beta2GPI/beta2GPI induces human neutrophils to generate NETs by relying on ROS. *Cell Biochem Funct.* 2019;37(2):56-61.
312. Leppkes M, Knopf J, Naschberger E, et al. Vascular occlusion by neutrophil extracellular traps in COVID-19. *EBioMedicine.* 2020;58:102925.
313. Bonow RO, Fonarow GC, O'Gara PT, et al. Association of Coronavirus Disease 2019 (COVID-19) With Myocardial Injury and Mortality. *JAMA Cardiol.* 2020;5(7):751-3.
314. Zuo Y, Yalavarti S, Shih H, et al. Neutrophil extracellular traps (NETs) as markers of disease severity in COVID-19. *JCI Insight.* 2020 Jun 4;5(11):e138999.
315. Middleton EA, He X, Denorme F, et al. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. *Blood.* 2020;136(10):1169-79.
316. Petit E, Falcinelli E, Paliani U, et al. Neutrophil more than platelet activation associates with thrombotic complications in COVID-19 patients. *J Infect Dis.* 2021;223(6):933-944.
317. Yang C, Montgomery M. Dornase alfa for cystic fibrosis. *Cochrane Database Syst Rev.* 2018;9:CD001127.
318. Earhart AP, Holliday ZM, Hofmann HV, et al. Consideration of dornase alfa for the treatment of severe COVID-19 acute respiratory distress syndrome. *New Microbes New Infect.* 2020;35:100689.
319. Okur HK, Yalcin K, Tastan C, et al. Preliminary report of In vitro and In vivo Effectiveness of Dornase alfa on SARS-CoV-2 infection. *New Microbes New Infect.* 2020:100756.
320. Desilles JP, Gregoire C, Le Cossec C, et al. Efficacy and safety of aerosolized intra-tracheal dornase alfa administration in patients with SARS-CoV-2-induced acute respiratory distress syndrome

(ARDS): a structured summary of a study protocol for a randomised controlled trial. *Trials*. 2020;21(1):548.

321. Toma A, Darwish C, Taylor M, et al. The Use of Dornase Alfa in the Management of COVID-19-Associated Adult Respiratory Distress Syndrome. *Crit Care Res Pract*. 2021;2021:8881115.
322. India MoD. DCGI approves anti-COVID drug developed by DRDO for emergency use 2021 [Available from: <https://pib.gov.in/PressReleasePage.aspx?PRID=1717007>.
323. Verma A, Adhikary A, Woloschak G, et al. A combinatorial approach of a polypharmacological adjuvant 2-deoxy-D-glucose with low dose radiation therapy to quell the cytokine storm in COVID-19 management. *Int J Radiat Biol*. 2020;96(11):1323-8.
324. Grant MC, Geoghegan L, Arbyn M, et al. The prevalence of symptoms in 24,410 adults infected by the novel coronavirus (SARS-CoV-2; COVID-19): A systematic review and meta-analysis of 148 studies from 9 countries. *PLoS One*. 2020;15(6):e0234765.
325. Milam KE, Parikh SM. The angiopoietin-Tie2 signaling axis in the vascular leakage of systemic inflammation. *Tissue Barriers*. 2015;3(1-2):e957508.
326. Ware LB, Matthay MA. Alveolar fluid clearance is impaired in the majority of patients with acute lung injury and the acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 2001;163(6):1376-83.
327. Martin S, Maruta K, Burkart V, et al. IL-1 and IFN-gamma increase vascular permeability. *Immunology*. 1988;64(2):301-5.
328. Fahey E, Doyle SL. IL-1 Family Cytokine Regulation of Vascular Permeability and Angiogenesis. *Front Immunol*. 2019;10:1426.
329. Barratt S, Medford AR, Millar AB. Vascular endothelial growth factor in acute lung injury and acute respiratory distress syndrome. *Respiration*. 2014;87(4):329-42.
330. Kaner RJ, Crystal RG. Compartmentalization of vascular endothelial growth factor to the epithelial surface of the human lung. *Mol Med*. 2001;7(4):240-6.
331. Mura M, dos Santos CC, Stewart D, et al. Vascular endothelial growth factor and related molecules in acute lung injury. *J Appl Physiol* (1985). 2004;97(5):1605-17.
332. Abadie Y, Bregeon F, Papazian I, et al. Decreased VEGF concentration in lung tissue and vascular injury during ARDS. *Eur Respir J*. 2005;25(1):139-46.
333. Zhang L, Liu H, Peng YM, et al. Vascular endothelial growth factor increases GEnC permeability by affecting the distributions of occludin, ZO-1 and tight junction assembly. *Eur Rev Med Pharmacol Sci*. 2015;19(14):2621-7.
334. Thickett DR, Armstrong L, Millar AB. A role for vascular endothelial growth factor in acute and resolving lung injury. *Am J Respir Crit Care Med*. 2002;166(10):1332-7.
335. Thickett DR, Armstrong L, Christie SJ, et al. Vascular endothelial growth factor may contribute to increased vascular permeability in acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 2001;164(9):1601-5.
336. Wada T, Jesmin S, Gando S, et al. The role of angiogenic factors and their soluble receptors in acute lung injury (ALI)/ acute respiratory distress syndrome (ARDS) associated with critical illness. *J Inflamm (Lond)*. 2013;10(1):6.
337. Loughna S, Sato TN. Angiopoietin and Tie signaling pathways in vascular development. *Matrix Biol*. 2001;20(5-6):319-25.
338. Jones N, Dumont DJ. Tek/Tie2 signaling: new and old partners. *Cancer Metastasis Rev*. 2000;19(1-2):13-7.
339. Hughes DP, Marron MB, Brindle NP. The antiinflammatory endothelial tyrosine kinase Tie2 interacts with a novel nuclear factor-kappaB inhibitor ABIN-2. *Circ Res*. 2003;92(6):630-6.
340. Pizurki L, Zhou Z, Glynos K, et al. Angiopoietin-1 inhibits endothelial permeability, neutrophil adherence and IL-8 production. *Br J Pharmacol*. 2003;139(2):329-36.

341. Hwang JA, Lee EH, Lee SD, et al. COMP-Ang1 ameliorates leukocyte adhesion and reinforces endothelial tight junctions during endotoxemia. *Biochem Biophys Res Commun*. 2009;381(4):592-6.
342. Parikh SM, Mammoto T, Schultz A, et al. Excess circulating angiopoietin-2 may contribute to pulmonary vascular leak in sepsis in humans. *PLoS Med*. 2006;3(3):e46.
343. Thurston G, Suri C, Smith K, et al. Leakage-resistant blood vessels in mice transgenically overexpressing angiopoietin-1. *Science*. 1999;286(5449):2511-4.
344. Hakanpää L, Sipilä T, Leppanen VM, et al. Endothelial destabilization by angiopoietin-2 via integrin β 1 activation. *Nat Commun*. 2015;6:5962.
345. Witzénbichler B, Westermann D, Kneuppel S, et al. Protective role of angiopoietin-1 in endotoxic shock. *Circulation*. 2005;111(1):97-105.
346. Bhandari V, Choo-Wing R, Lee CG, et al. Hyperoxia causes angiopoietin 2-mediated acute lung injury and necrotic cell death. *Nat Med*. 2006;12(11):1286-93.
347. Thomas M, Felcht M, Kruse K, et al. Angiopoietin-2 stimulation of endothelial cells induces α 5 β 3 integrin internalization and degradation. *J Biol Chem*. 2010;285(31):23842-9.
348. Lukasz A, Hillgruber C, Oberleithner H, et al. Endothelial glycocalyx breakdown is mediated by angiopoietin-2. *Cardiovasc Res*. 2017;113(6):671-80.
349. Sun NN, Li C, Zhou L, et al. Lentivirus-mediated angiopoietin 2 gene silencing decreases TNF- α induced apoptosis of alveolar epithelium cells. *Biochem Cell Biol*. 2016;94(5):491-7.
350. Syed M, Das P, Pawar A, et al. Hyperoxia causes miR-34a mediated injury via angiopoietin-1 in neonatal lungs. *Nat Commun*. 2017;8(1):1173.
351. Fiedler U, Reiss Y, Scharpfenecker M, et al. Angiopoietin-2 sensitizes endothelial cells to TNF- α and has a crucial role in the induction of inflammation. *Nat Med*. 2006;12(2):235-9.
352. Dumont DJ, Yamaguchi TP, Conlon RA, et al. Tie-2, a novel tyrosine kinase gene located on mouse chromosome 4, is expressed in endothelial cells and their presumptive precursors. *Oncogene*. 1992;7(8):1471-80.
353. Saharinen P, Eklund L, Miettinen J, et al. Angiopoietins assemble distinct Tie2 signalling complexes in endothelial cell-cell and cell-matrix contacts. *Nat Cell Biol*. 2008;10(5):527-37.
354. van der Heijden M, van Nieuw Amerongen GP, Koolwijk P, et al. Angiopoietin-2, permeability oedema, occurrence and severity of ALI/ARDS in septic and non-septic critically ill patients. *Thorax*. 2008;63(10):903-9.
355. Meyer NJ, Li M, Feng R, et al. ANGPT2 genetic variant is associated with trauma-associated acute lung injury and altered plasma angiopoietin-2 isoform ratio. *Am J Respir Crit Care Med*. 2011;183(10):1344-53.
356. Reilly JP, Wang F, Jones TK, et al. Plasma angiopoietin-2 as a potential causal marker in sepsis-associated ARDS development: evidence from Mendelian randomization and mediation analysis. *Intensive Care Med*. 2018;23(11):1849-58.
357. Gallagher DC, Parikh SM, Balonov K, et al. Circulating angiopoietin 2 correlates with mortality in a surgical population with acute lung injury/adult respiratory distress syndrome. *Shock*. 2008;29(6):656-61.
358. Zinter MS, Spicer A, Orwoll BO, et al. Plasma angiopoietin-2 outperforms other markers of endothelial injury in prognosticating pediatric ARDS mortality. *Am J Physiol Lung Cell Mol Physiol*. 2016;310(3):L224-31.
359. Agrawal A, Matthay MA, Kangelaris KN, et al. Plasma angiopoietin-2 predicts the onset of acute lung injury in critically ill patients. *Am J Respir Crit Care Med*. 2013;187(7):736-42.
360. Calfee CS, Gallagher D, Abbott J, et al. Plasma angiopoietin-2 in clinical acute lung injury: prognostic and pathogenetic significance. *Crit Care Med*. 2012;40(6):1731-7.
361. Thille AW, Esteban A, Fernandez-Segoviano P, et al. Chronology of histological lesions in acute respiratory distress syndrome with diffuse alveolar damage: a prospective cohort study of clinical autopsies. *Lancet Respir Med*. 2013;1(5):395-401.

362. Matthay MA, Zemans RL, Zimmerman GA, et al. Acute respiratory distress syndrome. *Nat Rev Dis Primers*. 2019;5(1):18.
363. Pierce RW, Shabanova V, Canarie M, et al. Angiopoietin Level Trajectories in Toddlers With Severe Sepsis and Septic Shock and Their Effect on Capillary Endothelium. *Shock*. 2019;51(3):298-305.
364. Kumpers P, Lukasz A, David S, et al. Excess circulating angiopoietin-2 is a strong predictor of mortality in critically ill medical patients. *Crit Care*. 2008;12(6):R147.
365. Giuliano JS, Jr., Wheeler DS. Excess circulating angiopoietin-2 levels in sepsis: harbinger of death in the intensive care unit? *Crit Care*. 2009;13(1):114.
366. Dekker NAM, van Leeuwen ALI, van Strien WWJ, et al. Microcirculatory perfusion disturbances following cardiac surgery with cardiopulmonary bypass are associated with in vitro endothelial hyperpermeability and increased angiopoietin-2 levels. *Crit Care*. 2019;23(1):117.
367. Parikh SM. Dysregulation of the angiopoietin-Tie-2 axis in sepsis and ARDS. *Virulence*. 2013;4(6):517-24.
368. Whitcomb DC, Muddana V, Langmead CJ, et al. Angiopoietin-2, a regulator of vascular permeability in inflammation, is associated with persistent organ failure in patients with acute pancreatitis from the United States and Germany. *Am J Gastroenterol*. 2010;105(10):2787-92.
369. Benest AV, Kruse K, Savant S, et al. Angiopoietin-2 is critical for cytokine-induced vascular leakage. *PLoS One*. 2013;8(8):e70459.
370. Liao M, Liu Y, Yuan J, et al. Single-cell landscape of bronchoalveolar immune cells in patients with COVID-19. *Nat Med*. 2020;26(6):842-4.
371. Kolaczkowska E, Kubes P. Neutrophil recruitment and function in health and inflammation. *Nat Rev Immunol*. 2013;13(3):159-75.
372. Laforge M, Elbim C, Frere C, et al. Tissue damage from neutrophil-induced oxidative stress in COVID-19. *Nat Rev Immunol*. 2020;20(9):515-6.
373. Shi L, Wang Y, Liang X, et al. Is neutrophilia associated with mortality in COVID-19 patients? A meta-analysis and meta-regression. *Int J Lab Hematol*. 2020 ;42(6):e244-e247.
374. Yan X, Li F, Wang X, et al. Neutrophil to lymphocyte ratio as prognostic and predictive factor in patients with coronavirus disease 2019: A retrospective cross-sectional study. *J Med Virol*. 2020.
375. Sanchez-Cerrillo I, Landete P, Adrio B, et al. COVID-19 severity associates with pulmonary redistribution of CD1c+ DCs and inflammatory transitional and nonclassical monocytes. *J Clin Invest*. 2020; 130(12):6290-6300.
376. Fox SE, Akmatbekov A, Harbert JL, et al. Pulmonary and cardiac pathology in African American patients with COVID-19: an autopsy series from New Orleans. *Lancet Respir Med*. 2020;8(7):681-6.
377. Tomar B, Anders H, Desai J, et al. Neutrophils and Neutrophil Extracellular Traps Drive Necroinflammation in COVID-19. *Cells*. 2020;9(6).
378. Duan F, Guo L, Yang L, et al. Modeling COVID-19 with Human Pluripotent Stem Cell-Derived Cells Reveals Synergistic Effects of Anti-inflammatory Macrophages with ACE2 Inhibition Against SARS-CoV-2. *Res Sq*. 2020. rs.3.rs-62758.
379. Cortjens B, Ingelse SA, Calis JC, et al. Neutrophil subset responses in infants with severe viral respiratory infection. *Clin Immunol*. 2017;176:100-6.
380. Pillay J, Ramakers BP, Kamp VM, et al. Functional heterogeneity and differential priming of circulating neutrophils in human experimental endotoxemia. *J Leukoc Biol*. 2010;88(1):211-20.
381. Pillay J, Kamp VM, van Hoffen E, et al. A subset of neutrophils in human systemic inflammation inhibits T cell responses through Mac-1. *J Clin Invest*. 2012;122(1):327-36.
382. Vassallo A, Wood AJ, Subburayalu J, et al. The counter-intuitive role of the neutrophil in the acute respiratory distress syndrome. *Br Med Bull*. 2019;131(1):43-55.
383. Villanueva E, Yalavarthi S, Berthier CC, et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. *J Immunol*. 2011;187(1):538-52.

384. Hamam HJ, Palaniyar N. Post-Translational Modifications in NETosis and NETs-Mediated Diseases. *Biomolecules*. 2019;9(8).
385. Rossaint J, Herter JM, Van Aken H, et al. Synchronized integrin engagement and chemokine activation is crucial in neutrophil extracellular trap-mediated sterile inflammation. *Blood*. 2014;123(16):2573-84.
386. Goodman RB, Strieter RM, Martin DP, et al. Inflammatory cytokines in patients with persistence of the acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 1996;154(3 Pt 1):602-11.
387. Muller AM, Cronen C, Muller KM, et al. Heterogeneous expression of cell adhesion molecules by endothelial cells in ARDS. *J Pathol*. 2002;198(2):270-5.
388. Burns JA, Issekutz TB, Yagita H, et al. The beta2, alpha4, alpha5 integrins and selectins mediate chemotactic factor and endotoxin-enhanced neutrophil sequestration in the lung. *Am J Pathol*. 2001;158(5):1809-19.
389. Burns JA, Issekutz TB, Yagita H, et al. The alpha 4 beta 1 (very late antigen (VLA)-4, CD49d/CD29) and alpha 5 beta 1 (VLA-5, CD49e/CD29) integrins mediate beta 2 (CD11/CD18) integrin-independent neutrophil recruitment to endotoxin-induced lung inflammation. *J Immunol*. 2001;166(7):4644-9.
390. Summers C, Singh NR, White JF, et al. Pulmonary retention of primed neutrophils: a novel protective host response, which is impaired in the acute respiratory distress syndrome. *Thorax*. 2014;69(7):623-9.
391. Vogt KL, Summers C, Chilvers ER, et al. Priming and de-priming of neutrophil responses in vitro and in vivo. *Eur J Clin Invest*. 2018;48 Suppl 2:e12967.
392. Vitte J, Diallo AB, Boumaza A, et al. A Granulocytic Signature Identifies COVID-19 and Its Severity. *J Infect Dis*. 2020;222(12):1985-96.
393. Klok FA, Kruip M, van der Meer NJM, et al. Incidence of thrombotic complications in critically ill ICU patients with COVID-19. *Thromb Res*. 2020;191:145-7.
394. Helms J, Tacquard C, Severac F, et al. High risk of thrombosis in patients with severe SARS-CoV-2 infection: a multicenter prospective cohort study. *Intensive Care Med*. 2020;46(6):1089-98.
395. Suh YJ, Hong H, Ohana M, et al. Pulmonary Embolism and Deep Vein Thrombosis in COVID-19: A Systematic Review and Meta-Analysis. *Radiology*. 2020:203557.
396. Poissy J, Goutay J, Caplan M, et al. Pulmonary Embolism in Patients With COVID-19: Awareness of an Increased Prevalence. *Circulation*. 2020;142(2):184-6.
397. Middeldorp S, Coppens M, van Haaps TF, et al. Incidence of venous thromboembolism in hospitalized patients with COVID-19. *J Thromb Haemost*. 2020;18(8):1995-2002.
398. Oxley TJ, Mocco J, Majid S, et al. Large-Vessel Stroke as a Presenting Feature of Covid-19 in the Young. *N Engl J Med*. 2020;382(20):e60.
399. Ladikou EE, Sivaloganathan H, Milne KM, et al. Von Willebrand factor (vWF): marker of endothelial damage and thrombotic risk in COVID-19? *Clin Med (Lond)*. 2020;20(5):e178-e82.
400. Bazzan M, Montaruli B, Sciascia S, et al. Low ADAMTS 13 plasma levels are predictors of mortality in COVID-19 patients. *Intern Emerg Med*. 2020;15(5):861-3.
401. Philippe A, Chocron R, Gendron N, et al. Circulating Von Willebrand factor and high molecular weight multimers as markers of endothelial injury predict COVID-19 in-hospital mortality. *Angiogenesis*. 2021;24 (3):505-517.
402. Ruberto F, Chistolini A, Curreli M, et al. Von Willebrand factor with increased binding capacity is associated with reduced platelet aggregation but enhanced agglutination in COVID-19 patients: another COVID-19 paradox? *J Thromb Thrombolysis*. 2021; 52(1):105-110.
403. Rovas A, Osiaevi I, Buscher K, et al. Microvascular dysfunction in COVID-19: the MYSTIC study. *Angiogenesis*. 2021;24(1):145-57.
404. Agrawal P, Nawadkar R, Ojha H, et al. Complement Evasion Strategies of Viruses: An Overview. *Front Microbiol*. 2017;8:1117.

405. Peffault de Latour R, Bergeron A, Lengline E, et al. Complement C5 inhibition in patients with COVID-19 - a promising target? *Haematologica*. 2020;105(12):2847-50.
406. Diao B. Human kidney is a target for novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection. *Nat Commun*. 2021 May 4;12(1):2506
407. Shen B, Yi X, Sun Y, et al. Proteomic and Metabolomic Characterization of COVID-19 Patient Sera. *Cell*. 2020;182(1):59-72 e15.
408. Carvelli J, Demaria O, Vely F, et al. Association of COVID-19 inflammation with activation of the C5a-C5aR1 axis. *Nature*. 2020;588(7836):146-50.
409. Yan B, Freiwald T, Chauss D, et al. SARS-CoV-2 drives JAK1/2-dependent local complement hyperactivation. *Sci Immunol*. 2021;6(58).
410. Hidalgo M, Martinez-Garcia M, Le Tourneau C, et al. First-in-Human Phase I Study of Single-agent Vanucizumab, A First-in-Class Bispecific Anti-Angiopoietin-2/Anti-VEGF-A Antibody, in Adult Patients with Advanced Solid Tumors. *Clin Cancer Res*. 2018;24(7):1535-45.
411. Pang J, Xu F, Aondio G, et al. Efficacy and tolerability of bevacizumab in patients with severe Covid-19. *Nat Commun*. 2021;12(1):814.
412. Islam MA, Mazumder MA, Akhter N, et al. Extraordinary Survival Benefits of Severe and Critical Patients with COVID-19 by Immune Modulators: The Outcome of a Clinical Trial in Bangladesh. *Euroasian J Hepatogastroenterol*. 2020;10(2):68-75.
413. Sugiyama MG, Armstrong SM, Wang C, et al. The Tie-2-agonist Vasculotide rescues mice from influenza virus infection. *Sci Rep*. 2015;5:11030.
414. David S, Ghosh CC, Kumpers P, et al. Effects of a synthetic PEG-ylated Tie-2 agonist peptide on endotoxemic lung injury and mortality. *Am J Physiol Lung Cell Mol Physiol*. 2011;300(6):L851-62.
415. Dekker NAM, van Meurs M, van Leeuwen AL, et al. Vasculotide, an angiopoietin-1 mimetic, reduces pulmonary vascular leakage and preserves microcirculatory perfusion during cardiopulmonary bypass in rats. *Br J Anaesth*. 2018;121(5):1047-51.
416. Annane D, Heming N, Grimaldi-Bensoussan L, et al. Eculizumab as an emergency treatment for adult patients with severe COVID-19 in the intensive care unit: A proof-of-concept study. *EClinicalMedicine*. 2020;28:100590.
417. Diurno F, Numis FG, Porta G, et al. Eculizumab treatment in patients with COVID-19: preliminary results from real life ASI Napoli 2 Nord experience. *Eur Rev Med Pharmacol Sci*. 2020;24(7):4040-7.
418. Mastellos DC, Pires da Silva BGP, Fonseca BAL, et al. Complement C3 vs C5 inhibition in severe COVID-19: Early clinical findings reveal differential biological efficacy. *Clin Immunol*. 2020;220:108598.
419. Alexion Ap. Alexion Provides Update on Phase 3 Study of ULTOMIRIS® (ravulizumab-cwvz) in Hospitalized Patients with Severe COVID-19 2021 [Available from: <https://ir.alexion.com/news-releases/news-release-details/alexion-provides-update-phase-3-study-ultomiris-ravulizumab>.
420. Vlaar APJ, de Bruin S, Busch M, et al. Anti-C5a antibody IFX-1 (vilobelimab) treatment versus best supportive care for patients with severe COVID-19 (PANAMO): an exploratory, open-label, phase 2 randomised controlled trial. *Lancet Rheumatol*. 2020; (12):e764-e773.
421. Chaturvedi S, Brodsky RA, McCrae KR. Complement in the Pathophysiology of the Antiphospholipid Syndrome. *Front Immunol*. 2019;10:449.
422. Guo RF, Ward PA. Role of C5a in inflammatory responses. *Annu Rev Immunol*. 2005;23:821-52.
423. Kow CS, Hasan SS. Meta-analysis of Effect of Statins in Patients with COVID-19. *Am J Cardiol*. 2020;134:153-5.
424. Calfee CS, Delucchi KL, Sinha P, et al. Acute respiratory distress syndrome subphenotypes and differential response to simvastatin: secondary analysis of a randomised controlled trial. *Lancet Respir Med*. 2018;6(9):691-8.

425. Rizzo AN, Sammani S, Esquinca AE, et al. Imatinib attenuates inflammation and vascular leak in a clinically relevant two-hit model of acute lung injury. *Am J Physiol Lung Cell Mol Physiol*. 2015;309(11):L1294-304.
426. Rizzo AN, Aman J, van Nieuw Amerongen GP, et al. Targeting Abl kinases to regulate vascular leak during sepsis and acute respiratory distress syndrome. *Arterioscler Thromb Vasc Biol*. 2015;35(5):1071-9.
427. Aman J, Duijvelaar E, Botros L, et al. Imatinib in patients with severe COVID-19: a randomised, double-blind, placebo-controlled, clinical trial. *Lancet Respir Med*. 2021;9(9):957-68.
428. Karagiannidis C, Mostert C, Hentschker C, et al. Case characteristics, resource use, and outcomes of 10 021 patients with COVID-19 admitted to 920 German hospitals: an observational study. *Lancet Respir Med*. 2020;8(9):853-62.
429. Koehler P, Bassetti M, Chakrabarti A, et al. Defining and managing COVID-19-associated pulmonary aspergillosis: the 2020 ECMM/ISHAM consensus criteria for research and clinical guidance. *Lancet Infect Dis*. 2021;21(6):e149-e62.
430. Birocchi S, Manzoni M, Podda GM, et al. High rates of pulmonary artery occlusions in COVID-19. A meta-analysis. *Eur J Clin Invest*. 2021;51(1):e13433.
431. Tang N, Bai H, Chen X, et al. Anticoagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy. *J Thromb Haemost*. 2020;18(5):1094-9.
432. Lu YF, Pan LY, Zhang WW, et al. A meta-analysis of the incidence of venous thromboembolic events and impact of anticoagulation on mortality in patients with COVID-19. *Int J Infect Dis*. 2020;100:34-41.
433. Sridharan GK, Vegunta R, Rokkam VRP, et al. Venous Thromboembolism in Hospitalized COVID-19 Patients. *Am J Ther*. 2020;27(6):e599-e610.
434. White D, MacDonald S, Bull T, et al. Heparin resistance in COVID-19 patients in the intensive care unit. *J Thromb Thrombolysis*. 2020;50(2):287-91.
435. Levy JH, Connors JM. Heparin Resistance - Clinical Perspectives and Management Strategies. *N Engl J Med*. 2021;385(9):826-32.
436. Spyropoulos AC. The management of venous thromboembolism in hospitalized patients with COVID-19. *Blood Adv*. 2020;4(16):4023.
437. Hippensteel JA, LaRiviere WB, Colbert JF, et al. Heparin as a therapy for COVID-19: current evidence and future possibilities. *Am J Physiol Lung Cell Mol Physiol*. 2020;319(2):L211-L7.
438. Investigators R-C, Investigators AC-a, Investigators A, et al. Therapeutic Anticoagulation with Heparin in Critically Ill Patients with Covid-19. *N Engl J Med*. 2021;385(9):777-89.
439. Panka BA, de Groot NJ, Spoelstra-de Man AM, et al. Prevention or Treatment of Ards With Aspirin: A Review of Preclinical Models and Meta-Analysis of Clinical Studies. *Shock*. 2017;47(1):13-21.
440. Kor DJ, Carter RE, Mark PK, et al. Effect of Aspirin on Development of ARDS in At-Risk Patients Presenting to the Emergency Department: The LIPS-A Randomized Clinical Trial. *JAMA*. 2016;315(22):2406-14.
441. Gritti G, Raimondi F, Bottazzi B, et al. Siltuximab downregulates interleukin-8 and pentraxin 3 to improve ventilatory status and survival in severe COVID-19. *Leukemia*. 2021;35(9):2710-4.
442. Della-Torre E, Lanzillotta M, Campochiaro C, et al. Respiratory Impairment Predicts Response to IL-1 and IL-6 Blockade in COVID-19 Patients With Severe Pneumonia and Hyper-Inflammation. *Front Immunol*. 2021;12:675678.
443. Libby P, Luscher T. COVID-19 is, in the end, an endothelial disease. *Eur Heart J*. 2020;41(32):3038-44.
444. Tzotzos SJ, Fischer B, Fischer H, et al. Incidence of ARDS and outcomes in hospitalized patients with COVID-19: a global literature survey. *Crit Care*. 2020;24(1):516.

445. Peck TJ, Hibbert KA. Recent advances in the understanding and management of ARDS. *F1000Res*. 2019;8.
446. Bellani G, Laffey JG, Pham T, et al. Epidemiology, Patterns of Care, and Mortality for Patients With Acute Respiratory Distress Syndrome in Intensive Care Units in 50 Countries. *JAMA*. 2016;315(8):788-800.
447. Force ADT, Ranieri VM, Rubenfeld GD, et al. Acute respiratory distress syndrome: the Berlin Definition. *JAMA*. 2012;307(23):2526-33.
448. Ferguson ND, Fan E, Camporota L, et al. The Berlin definition of ARDS: an expanded rationale, justification, and supplementary material. *Intensive Care Med*. 2012;38(10):1573-82.
449. Bartoletti M, Pascale R, Cricca M, et al. Epidemiology of invasive pulmonary aspergillosis among COVID-19 intubated patients: a prospective study. *Clin Infect Dis*. 2020.
450. Li L, Zhang W, Hu Y, et al. Effect of Convalescent Plasma Therapy on Time to Clinical Improvement in Patients With Severe and Life-threatening COVID-19: A Randomized Clinical Trial. *JAMA*. 2020;324(5):460-70.
451. Agarwal A, Mukherjee A, Kumar G, et al. Convalescent plasma in the management of moderate covid-19 in adults in India: open label phase II multicentre randomised controlled trial (PLACID Trial). *BMJ*. 2020;371:m3939.
452. Abolghasemi H, Eshghi P, Cheraghali AM, et al. Clinical efficacy of convalescent plasma for treatment of COVID-19 infections: Results of a multicenter clinical study. *Transfus Apher Sci*. 2020;59(5):102875.
453. Simonovich VA, Burgos Pratz LD, Scibona P, et al. A Randomized Trial of Convalescent Plasma in Covid-19 Severe Pneumonia. *N Engl J Med*. 2020.
454. Joyner MJ, Senefeld JW, Klassen SA, et al. Effect of Convalescent Plasma on Mortality among Hospitalized Patients with COVID-19: Initial Three-Month Experience. *medRxiv*. 2020.
455. JoynerMJ CR, Senefeld JW et al. Convalescent Plasma Antibody Levels and the Risk of Death from Covid-19. *NEJM*. 2021;Jan 2021.
456. Chai KL, Valk SJ, Piechotta V, et al. Convalescent plasma or hyperimmune immunoglobulin for people with COVID-19: a living systematic review. *Cochrane Database Syst Rev*. 2020;10:CD013600.
457. Cicceri F, Castagna A, Rovere-Queiro P, et al. Early predictors of clinical outcomes of COVID-19 outbreak in Milan, Italy. *Clin Immunol*. 2020;217:108509.
458. Stefanini GG, Chiarito M, Perente G, et al. Early detection of elevated cardiac biomarkers to optimise risk stratification in patients with COVID-19. *Heart*. 2020;106(19):1512-8.
459. Masetti C, Generali E, Colanietro F, et al. High mortality in COVID-19 patients with mild respiratory disease. *Eur J Clin Invest*. 2020;50(9):e13314.
460. Feng Z, Yu Q, Yao S, et al. Early prediction of disease progression in COVID-19 pneumonia patients with chest CT and clinical characteristics. *Nat Commun*. 2020;11(1):4968.
461. Jain V, Yuan JM. Predictive symptoms and comorbidities for severe COVID-19 and intensive care unit admission: a systematic review and meta-analysis. *Int J Public Health*. 2020;65(5):533-46.
462. consortium WSt. Repurposed antiviral drugs for COVID-19 –interim WHO SOLIDARITY trial results. *MedRxiv*. 2020;October 15 version.
463. Davoudi-Monfared E, Rahmani H, Khalili H, et al. A Randomized Clinical Trial of the Efficacy and Safety of Interferon beta-1a in Treatment of Severe COVID-19. *Antimicrob Agents Chemother*. 2020;64(9).
464. Estebanez M R-OG, Mata T et al. Clinical evaluation of IFN beta1b in COVID-19 pneumonia: a retrospective study 2020 [Available from: <https://doi.org/10.1101/2020.05.15.20084293>].
465. Hung IF, Lung KC, Tso EY, et al. Triple combination of interferon beta-1b, lopinavir-ritonavir, and ribavirin in the treatment of patients admitted to hospital with COVID-19: an open-label, randomised, phase 2 trial. *Lancet*. 2020;395(10238):1695-704.

466. Pereda R, Gonzalez D, Rivero HB, et al. Therapeutic Effectiveness of Interferon-alpha2b Against COVID-19: The Cuban Experience. *J Interferon Cytokine Res.* 2020;40(9):438-42.
467. Zhou Q, Chen V, Shannon CP, et al. Interferon-alpha2b Treatment for COVID-19. *Front Immunol.* 2020;11:1061.
468. Rosas IO, Brau N, Waters M, et al. Tocilizumab in Hospitalized Patients with Severe Covid-19 Pneumonia. *N Engl J Med.* 2021.
469. Salama C, Han J, Yau L, et al. Tocilizumab in Patients Hospitalized with Covid-19 Pneumonia. *N Engl J Med.* 2021;384(1):20-30.
470. Stone JH, Frigault MJ, Serling-Boyd NJ, et al. Efficacy of Tocilizumab in Patients Hospitalized with Covid-19. *N Engl J Med.* 2020;383(24):2333-44.
471. Hermine O, Mariette X, Tharaux PL, et al. Effect of Tocilizumab vs Usual Care in Adults Hospitalized With COVID-19 and Moderate or Severe Pneumonia: A Randomized Clinical Trial. *JAMA Intern Med.* 2021;181(1):32-40.
472. Bronte V, Ugel S, Tinazzi E, et al. Baricitinib restrains the immune dysregulation in patients with severe COVID-19. *J Clin Invest.* 2020;130(12):6409-16.
473. Rosas J, Liano FP, Canto ML, et al. Experience With the Use of Baricitinib and Tocilizumab Monotherapy or Combined, in Patients With Interstitial Pneumonia Secondary to Coronavirus COVID19: A Real-World Study. *Reumatol Clin.* 2020.
474. Cao Y, Wei J, Zou L, et al. Ruxolitinib in treatment of severe coronavirus disease 2019 (COVID-19): A multicenter, single-blind, randomized controlled trial. *J Allergy Clin Immunol.* 2020;146(1):137-46 e3.
475. Giudice V, Pagliano P, Vatrella A, et al. Combination of Ruxolitinib and Eculizumab for Treatment of Severe SARS-CoV-2-Related Acute Respiratory Distress Syndrome: A Controlled Study. *Front Pharmacol.* 2020;11:857.
476. Kalil AC, Patterson TF, Mehta AK, et al. Baricitinib plus Remdesivir for Hospitalized Adults with Covid-19. *N Engl J Med.* 2020.
477. Pavoni V, Ganesello L, Pazzi M, et al. Venous thromboembolism and bleeding in critically ill COVID-19 patients treated with higher than standard low molecular weight heparin doses and aspirin: A call to action. *Thromb Res.* 2020;196:313-7.
478. Chow JH KA, Kethireddy S et al. Aspirin Use is Associated with Decreased Mechanical Ventilation, ICU Admission, and In-Hospital Mortality in Hospitalized Patients with COVID-19. *Anesth Analg.* 2020;Oct 21.
479. Yuan S, Chen P, Li H, et al. Mortality and pre-hospitalization use of low-dose aspirin in COVID-19 patients with coronary artery disease. *J Cell Mol Med.* 2020.
480. Salah HM, Mekki J. Meta-Analysis of the Effect of Aspirin on Mortality in COVID-19. *Am J Cardiol.* 2021.
481. Hasan SS, Radford S, Kow CS, et al. Venous thromboembolism in critically ill COVID-19 patients receiving prophylactic or therapeutic anticoagulation: a systematic review and meta-analysis. *J Thromb Thrombolysis.* 2020;50(4):814-21.
482. Hall DA, Hanrott K, Badorrek P, et al. Effects of Recombinant Human Angiotensin-Converting Enzyme 2 on Response to Acute Hypoxia and Exercise: A Randomised, Placebo-Controlled Study. *Pulm Ther.* 2021 Jun 26:1–15. doi: 10.1007/s41030-021-00164-7. Epub ahead of print.
483. Hifumi T, Isokawa S, Otani N, Ishimatsu S. Adverse events associated with nafamostat mesylate and favipiravir treatment in COVID-19 patients. *Crit Care.* 2020;24(1):497.
484. Marconi VC, Ramanan AV, de Bono S. et al. Efficacy and safety of baricitinib for the treatment of hospitalised adults with COVID-19 (COV-BARRIER): a randomised, double-blind, parallel-group, placebo-controlled phase 3 trial. *Lancet Respir Med.* 2021; [https://doi.org/10.1016/S2213-2600\(21\)00331-3](https://doi.org/10.1016/S2213-2600(21)00331-3)

485. Pupim L, Wang TS, Hudock K, et al. Mavrililumab improves outcomes in phase 2 trial in non-mechanically ventilated patients with severe COVID-19 pneumonia and systemic hyperinflammation. *Ann Rheum Dis* 2021;80:198-9

Journal Pre-proof

Journal Pre-proof

Table 1. Relevant trials assessing convalescent Plasma (CP) in COVID-19 (selection)				
Study	Design, n	WHO stage of included patients, administered dose	outcomes	
Li [450]	RCT, n=103	4-13 mL/kg, variable titers Severe COVID (23/22), life threatening COVID (29/29)	Time to improvement at 28d by 2OSP: overall Severe COVID Life threatening COVID-19 28D mortality	HR 1.4 (0.79-2.49) HR 2.15 (1.07-4.32) HR 0.88 (0.3-2.63) HR 0.59 (0.22-1.59) No effect on time to improvement or mortality, possible signal for clinical benefit in severe but not life threatening COVID-19
Agarwal[451]	PLACID, RCT open label n=464	moderate COVID-19 (SaO ₂ ≤93% in RA, PaO ₂ /FiO ₂ 200-300)	Progression to severe COVID (PaO ₂ /FiO ₂ ≤100) 28D mortality	HR 1.04 (0.54-1.98) HR 1.04 (0.66-1.63) No effect on mortality or disease progression
Gharbharan [167]	ConCOVID, n=86, RCT	Hospitalized, not MV≥4d, but otherwise not well defined	Mortality Clinical improvement D15	79% of patients had antibodies at baseline HR 0.95 (0.2-4.7) HR 1.3 (0.52-3.32)
Abolghasemi [452]	open label RCT, N=189	Mod. COVID-19 (stages 4,5), hospit. line 1 for ≤3d, O ₂ requirement, not intubated	28D mortality Progression to MV	14.8% vs 24.3% , p=0.09 7% vs 20.3%, p=0.006
Simonovich [453]	PlasmAR, RCT, n=333	Hospitalized, with O ₂ requirement (any). Almost all received steroids	30D mortality Improvement on ordinal scale D14	HR 0.83 (0.52-1.46) HR 1.00 (0.65-1.55) No significant benefit in severe COVID
Joyner [454]	Observational, n=35,522	Hospitalized, ICU: 52.3% MV: 27.5%	7D mortality pts who received high-titer CP, no MV 7D mortality in pts treated with CP within 3d, no MV 7D mortality in those treated < vs ≥3d after diagnosis 30D mortality in those	14% vs 11%, p=0.03 6.3% vs 11.3%, p=0.0008 8.7% (8.3-9.2%) vs 11.9% (11.4-12.2%), p≤0.001 21.6% vs 26.7% , p≤0.001

			treated < vs ≥3d after diagnosis	
Joyner [455]	retrospective, n=3082	WHO stage 4,5,6,7	30D mortality (high titer CP) 30D mortality (high titer CP), not MV 30D mortality (high titer CP) MV (low titer and/or already on MV – no benefit. Data not shown)	HR 0.75 (0.61-0.93) HR 0.64 (0.46-0.88) HR 0.93 (0.73-1.19)
Chai [456]	Cochrane review, 19 observational studies and RCTs	N=38.160 patients (36.081 received CP) RCTs: n=189 (95 received CP)	Mortality Improvement of clinical symptoms at D7 Improvement of clinical symptoms at D15 Improvement of clinical symptoms at D30	HR 0.64 (0.33-1.25) RR 0.98 (0.3-3.19) RR 1.34 (0.85-2.11) RR 1.13 (0.88-1.43)
Recovery	RCT, open label, n=10406	Not released, pre-publication communication to investigators	28D mortality	HR 1.04 (0.95-1.14)

Table. 2 Clinical Risk Score in hospitalized patients with COVID-19

	Symptom/marker on	OR disease progression to ICU	OR death
--	-------------------	-------------------------------	----------

	admission	or critical illness	
Liang [177] AUC 0.88	Chest x-ray abnormal	3.39	
	Hemoptysis	4.53	
	Dyspnea	1.88	
	Level of consciousness	4.71	
	History of malignancy	4.07	
	NLR raised	1.06	
	LDH raised	1.02	
	Bilirubin raised	1.15	
	Number of comorbidities	1.60	
Ciceri [457]	>65 years of age		3.17
	History of coronary disease		2.93
	Lymphocytes $<0.9 \times 10^9$		1.83
	Higher RALE score		1.05
	LDH above median		2.95
	D-Dimer above median		2.54
Stefanini [458] AUC 0.88	hsTroponin		1.32
	Lymphocytes		0.52
	Age		1.1
	O ₂ requirement		2.55
	Tachypnoea $>20/\text{min}$		1.84
	Tachycardia $>100/\text{min}$		0.36
	Fever		2.12
	GFR $<60\text{mL}/\text{min} \times 1.73\text{m}^2$		2.19
	Malignancy		2.38
	D-Dimer		1.51
AUC 0.92	Age		1.13
	GFR $<60\text{mL}/\text{min} \times 1.73\text{m}^2$		2.66
	hsTroponin AND BNP		3.24
	D-Dimer		1.00
	Lymphocyte		0.19
	SaO ₂ desaturation		2.07
Masetti [459]	Age $>75\text{years}$		10.6
	Thrombocytopenia $<150 \times 10^9/\text{L}$		3.64

	Ferritin >750ng/mL		3.33
Henry [183], (metanalysis)	Lymphopenia	4.2	3.7
	Neutrophilia	7.99	7.87
	Lymphopenia <0.5 x10 ⁹ /L		12.0
Hao [179] Hospitalization	SpO ₂	5.67	
	Fever	2.36	
	Age	2.4	
	Tachycardia	2	
	Diastolic BP	4.51	
	Dyspnoea	7.41	
	Chronic kidney disease	2.25	
ICU	Chest x-ray opacity	4.08	
	Tachypnoea	1.66	
	Age	1.76	
	Fever	1.83	
	Male	1.62	
	Hypoalbuminemia	1.78	
	SpO ₂	2.29	
	LDH	2.62	
	Ca ²⁺	1.73	
Mechanical ventilaion	CRP	1.53	
	LDH	6.47	
	Ca ²⁺	1.79	
Feng [460]	Age	1.06	
	NLR	1.74	
	CT severity score	1.19	
Jain [461]	Progression to severe disease	Dyspnea 3.7	
	Progression to ITU	Dyspnea 6.5	
Li [188]	hsTrop, CK, LDH	See text	
Caricchio [178]	Six criteria predicting cytokine storm, see text	Predicted cytokine storm/ use of cytokine blockade	

Table 3.: Studies assessing interferon for use in COVID-19 (selection)

Study	DESIGN, N	WHO STAGE OF INCLUDED	OUTCOMES	
-------	-----------	-----------------------	----------	--

		PATIENTS, ADMINISTERED DOSE		
Solidarity [462]	RCT, open label, n=2050	INFβ1a 3x 0.44mcg s/c or iv for 1week. WHO stages 3-6 In air n=482/490 O ₂ req. n=1429/1430 Ventilated n=139/130	28 day mortality MV No MV	HR 1.16 (0.96-1.39) HR 1.4 (0.82-2.4) HR 1.1 (0.84-1.45)
Rhamani [219]	RCT, open label, n=80 (33/33)	INFβ1b 250 mcg s/c for 2 weeks, combined with LPV/r/ ATV/r and HCQ. WHO stage 4 (6% IFN group), 5 (75% IFN group), 6ff (18% IFN group)	Time to clinical improvement Discharge D14 ICU admission 28D mortality	9(6-10) vs 11 (9-15), HR 2.30, p=0.002 78.8% vs 54.6%, OR 3.09, p=0.03 14 (66.7%) vs 22 (42.4%), p=0.04 2 (6%) vs 6 (18.2%), p=0.12
Davoudi-Monfared [463]	RCT, n=92 (46/46)	INFβ1a 0.44mcg s/c, 3x weekly for 2 weeks. combined with LPV/r, HCQ, GCs. SaO ₂ ≤90%, median symptom duration 10d	D28 discharge D28 overall mortality Progression to MV Mortality early IFN (<10d) Mortality late IFN (>10d)	31 (73.8%) vs 23(58.9%), OR 1.96(0.8-5) 8 (19%) vs 15 (43.6%), p=0.015 35% vs 44%, p=0.33 OR, 13.5;95%CI 1.5-118) OR, 2.1; 95%CI 0.48-9.6
Estebanez [464]	Observational retrospective. N=256 (106/150)	INFβ1b at 250mcg s/c for 1-2 weeks on alternate days, combined LPV/r, HCQ, or TCZ, GCs. (mild 26%, moderate 36%, severe 18%) median symptom duration 7d	Mortality	20.8% vs 27.3% p=0.229
Hung [465]	RCT open label, n= 127 (86/41)	INFβ1b s/c 500 IU for 1-3 doses. Combined with LPV/r, ribavirin. Most WHO stage 3	Time to SARS-CoV-2 PCR neg Clinical improvement Length of hospitalization	7d vs 12d (RR 4.37 (1.86-10.24) p=0.001 4d (3-8) vs 8d (7-9), p<0.0001 9d vs 14.5d
Wang [220]	Retrospective, observational	IFNα2b, Early= within 5d (48%) Late= after 5d (5.8%) No IFN (45.7%). Most WHO stage 3,4,5	In-hospital mortality Early IFN vs no IFN Late IFN vs no IFN Age >60y	Early (0.9%), late (15.4%), non (4.9%) aHR mortality 0.05 (0.01-0.37), p=0.004 aHR mortality 6.82 (1.14-40.8), p=0.005 HR mortality 6.87 (p≤0.001) – treatment independent.
Pereda [466]	Observational N=814	IFNα2b 3x per week for 2 weeks, i.m.	Note: 75% of control group but 5.5% of treatment group on ICU	

		Majority combined with LPV/r, HCQ	at inclusion Discharge Fatality rate overall Fatality rate for severe disease	145 (95.4%) vs 6(26.1%) 7 (0.9%) vs 17 (32.1%) 7 (21.9%) vs 17 (48.5%)
Monk [222]	Blinded, placebo controlled RCT, n=101 (50/51)	Nebulized IFN β 1a 6mio IU once daily for 14d, WHO stage 3, 4,5, median symptom duration 10d (7-11d)	Recovery D15 Recovery D28 Discharge D15 Discharge D28 Improvement D15 Improvement D28 Progression to ICU/severe disease	OR 3.19 (1.24-8.24) OR 3.58 (1.41-9.04) OR 1.63 (0.61-4.35) OR 1.84 (0.64-5.29) OR 2.32 (1.07-5.04) OR 3.15 (1.39-7.14) OR 0.21 (0.04-0.97) p=0.046

Table 4. Interleukin-6 inhibition in COVID-19 (selection)

Study	DESIGN, N	WHO STAGE INCLUDED, DRUG ADMINISTERED	OUTCOMES	RESULT
COVACTA [468]	Multinational RCT, N=452	8mg/kg Tocilizumab iv once or twice. Hospitalized patients at WHO stage ≥ 4 . Co-administration of SOC except: immunomodulators other than GCs	Median ordinal scale D28 Median ordinal scale D14 Mortality overall D28 Median ordinal scale D28 if MV Need for ICU transfer	1.0 (TCZ); 2.0 (1.0-4.0) placebo, p=0.31 3.0 (2.0-4.0) TCZ; 4.0 (3.0-5.0) placebo 19.7% TCZ; 19.4% placebo; p=0.94 5.0 (3.0-5.0) TCZ; 5.0 (4.0-6.0) placebo 21.3% TCZ; 35.9% placebo
EMPACTA [469]	Double-blinded, placebo-controlled RCT, n=249	8mg/kg Tocilizumab i.v. Hospitalized patients at WHO stage ≥ 4 , excluded if requiring pressure support, >50% received steroids	Progression to MV or death, (composite) overall Mortality Overall	12.0 (8.5-16.9)% TCZ; 19.3 (13.3-27.4)% placebo; HR 0.56; p=0.04 11.6% TCZ; 11.8% placebo; p=N.S.
BACC BAY[470]	Double-blinded, placebo-controlled RCT, n=243	8mg/kg Tocilizumab single dose. majority WHO stage 3 (supplemental oxygen only). GCs in 6% placebo, 11% TCZ	Mortality D28 Time to ICU admission or death Oxygen weaned at D14	10.6 (6.7-16.6) TCZ; 12.5 (6.9-22)% placebo, p=0.64 15.9 TCZ; 15.8% placebo, p=0.97 75.4% TCZ; 78.8% placebo, p=N.S.
CORIMUNO-TOCI[471]	RCT, n=131	Tocilizumab 8mg/kg, repeat if no improvement	28D mortality	7/64 (89%) TCZ, 8/67 (88%) SOC; HR 0.92 (0.33-2.53)

		GCs in 33% Patients at WHO stage ≥ 3	Oxygen weaned by D28	89% TCZ; 75% SOC; HR 1.41 (0.98-2.01)
--	--	--	----------------------	---------------------------------------

Table 5. Jak-inhibitor trials in COVID-19 (selection)

Study	DESIGN, N	WHO STAGE INCLUDED, DRUG ADMINISTERED	OUTCOMES	RESULT
Bronte [472]	Observational, n=96 (n=20 treatment/ n=76 control)	Baricitinib 4mg BD for 2d, then 4mg OD for 1 week. Clinical stage not specified.	narrative	Faster reduction in O2 supplementation
Cantini [286]	Observational, retrospective. N=192 (78/113)	Baricitinib. Moderate COVID-19. FiO ₂ 200-300. No GCs given	14D mortality ICU admission at 2 weeks Discharge at 2 weeks	0% vs 6.4% , p=0.01 0.88% vs 17.9%, p \leq 0.001 77.8% vs 12.8%, p \leq 0.0001
Cantini [285]	observational, n=24 (12/12)	Baricitinib 2 weeks, combined LPV/r, HCQ. mild-moderate COVID-19, SaO ₂ <93%	Mortality ICU admission Discharge at 14D	1/20 (5%) vs 25/56 (45%) 0% vs 33%, p=0.093 58% vs 8%, p=0.027
Rosas [473]	Retrospective N=60	Baricitinib, TCZ or combination of baricitinib and TCZ. Moderate-severe disease	2/12 deaths on baricitinib monotherapy 4/20 deaths on TCZ monotherapy 3/11 deaths on baricitinib +TCZ	Mortality lowest on baricitinib monotherapy. No serious adverse events were observed
Cao [474]	RCT open label, n=43 (22/21)	Ruxolitinib (10mg BD for 14d) WHO stages 4 (most) and 5	Mortality Clinical improvement D14	3 (7.3%) vs 3 (14.3%), p=0.23 21 (51.2%) vs 9 (42.9%), p=0.35
Giudice [475]	Observational, n=17 (7/10)	Ruxolitinib (10mg BD for 14d) and Eculizumab (D7 and D14), hospitalized, severe COVID-19. Combined with GCs, antivirals.	Mortality Progression to ARDS	1/7 vs 1/10 1/7 vs 4/10
Kalil [476]	double-blinded, placebo controlled RCT N=1033	Baricitinib +/- temdesivir WHO stage 4ff	Clinical improvement at D15 Mortality at 28D all Mortality at 28D stage 4 (suppl O2) Mortality at 28D stage 5 (HF or NIV)	OR 1.3 (1.0-1.6) 5.1% vs 7.8% (HR 0.65 (0.39-1.09)) 1.9% vs 4.7% (HR 0.4 (0.14-1.14)) 7.5% vs 12.9%, HR 0.55 (0.22-1.38)

	(515/518)		Time to recovery WHO stage 3 Time to recovery WHO stage 4 Time to recovery WHO stage 5 Time to recovery MV	RR 0.88 (0.63-1.23) RR 1.17 (0.98-1.39) RR 1.51 (1.1-2.08) RR 1.08 (0.59-1.97)
Marconi [484]	Double blinded, placebo- controlled RCT	Baricitinib 4mg OD for 14d WHO stages 3, 4, 5	28d Mortality overall 28d Mortality WHO stage 4 28d Mortality WHO stage 5	HR 0.57 [0.41-0.78] HR 0.75 [0.45-1.16] HR 0.52 [0.33-0.80]
Guimaraes [291]	Placebo- controlled, open label RCT	Tofacitinib 10mgBD for 14d WHO stage 4, 5 (high flow but no pressure support)	Death or MV day 28 Death 28 d (any cause)	18.1% vs 29% (HR0.63 [0.41-0.97]) 2.8% vs 5.5% (HR 0.49 [0.15-1.63])

Table 6. Trials assessing heparin and Aspirin use in COVID-19 (selection)			
Study	Design, intervention, n	Parameters	Outcome
Pavoni [477]	Observational, n=42 WHO stage ≥ 5 ff, high risk group: 90% MV, low risk group: 23% MV	DD \leq 3000 n=22: ASA, LMWH 4000-6000IU DD \geq 3000 n=20: ASA, LMWH 1000IU/kg	LR group: 14% VTE, 4.5% PE; Mortality: 18% HR group: 65% VTE, 10% PE; Mortality: 25%
Chow [478]	Observational retrospective cohort, n=412 WHO stages 4, 5	N=314 no aspirin N=98 aspirin prior to admission Progression to MV Progression to ICU In-hospital mortality	aHR 0.56, 0.37-0.85, p=0.007 aHR 0.57, 0.38-0.85, p=0.005 aHR 0.53, 0.31-0.90, p=0.02
Yuan [479]	Observational, n=183 (52/131) patients with coronary artery disease (all WHO stages) who were either on ongoing ASA or not	WHO stages 5 (HF O2) 84.6% (ASA), 80.9% (no ASA) 5 (NIV) 19.2% (ASA), 26% (no ASA) 6ff (MV) 1.9% (no ASA), 11.5% (no ASA) All-cause mortality	OR 0.94 (0.41-2.17), p=0.89
Petito [316]	Observational, Netosis markers in n=36 COVID-19 patients, n=31 healthy controls	Prediction of VTE: MPO-DNA Cit3H	AUC 0.77, p<0.001 AUC 0.79, p<0.001
Hasan [481]	Metanalysis of 12 studies. ICU COVID-19 patients, UFH or LMWH	Prophylactic vs therapeutic anticoagulation of patients with COVID-19 on ICU Pooled prevalence of VTE (all)	31% (21-43%)

		VTE in prophylactic VTE in therapeutic (and prophylactic)	38% (10-70%) 27% (17-40%)
Lu [432]	Metanalysis, 20 observational (VTE incidence) 5 observational (VTE and mortality)	Incidence VTE (pooled, all) Incidence VTE (pooled, ICU) Incidence PE (pooled, all) Incidence PE (pooled ICU) Incidence DVT (pooled, all) Incidence DVT (pooled ICU) Mortality (with/without heparinization n=2886/5647)	255/ 1808, 21% (15-27%) 169/656, 27% (16-38%) 238/1793 15% (10-20%) 148/690 20% (9-31%) 212/1243 27% (19-36%) 99/579 33% (19-47%) RR 0.86 (0.69-1.09)
Birocchi [430]	Metanalysis, 26 studies (17 COVID-19 studies, n=3224; 7 non-COVID-19 studies, n=11,985)	67% COVID-19 on heparin prophylaxis 16% COVID-19 on therapeutic heparin DVT prevalence (pooled) PE (pooled) Non ICU DVT PE ICU patients only DVT PE	15.4% (4.08-31.8%) vs 4.2% (2.3-6.7%) p=0.046 4.9% (0.3-13%) vs 0.2% (0.03-0.6%) p=0.013 2.63%(0.7-5.6%) vs 3.64 (1.9-5.8%) p=0.48 2.83% (1.2-5.1%) vs 0.11 (0.0-0.3), p<0.0001 9.1% (3.6-16.7%) vs 7.4% (6.2-8.7%) p=0.63 11.7% (5.3-20.1) vs 0.96% (0.57-1.5%) p=0.0001 22.2% (5.3-44.6%) vs 6.4% (3.2-10.4%) p=0.48 57% (38-78%) vs 11.5% (6.9-17.6%) p=0.0002
Sridharan [433]	Metanalysis, 11 studies	VTE in hospitalized COVID-19 patients Prophylactic heparin dose Therapeutic heparin dose	12.5% 17.2% OR 0.33 (0.14-0.75), p=0.008

Figure 1.: WHO Ordinal 9 Point Scale and therapeutic options recommended and under investigation during the different disease stages.